

Characterization of the Whole-Genome Sequence of a Beak and Feather Disease Virus Isolate from a Mallee Ringneck Parrot (*Barnardius zonarius barnardi*)

Shubhagata Das,^{a,c} Subir Sarker,^{a,c} Jade K. Forwood,^{b,c} Seyed A. Ghorashi,^{a,c} Shane R. Raidal^{a,c}

School of Animal and Veterinary Sciences, Charles Sturt University, Wagga Wagga, New South Wales, Australia^a; School of Biomedical Sciences, Charles Sturt University, Wagga Wagga, New South Wales, Australia^b; Graham Centre for Agricultural Innovation, Wagga Wagga, New South Wales, Australia^c

The complete genome sequence of beak and feather disease virus (BFDV) from a wild Australian Mallee ringneck parrot (*Barnardius zonarius barnardi*) was characterized. The genome consists of 1,995 nucleotides and encodes two major proteins in opposing directions. This is the first evidence of BFDV infectivity and the first complete genome sequence for this novel host.

Received 20 June 2014 Accepted 3 July 2014 Published 24 July 2014

Citation Das S, Sarker S, Forwood JK, Ghorashi SA, Raidal SR. 2014. Characterization of the whole-genome sequence of a beak and feather disease virus isolate from a Mallee ringneck parrot (*Barnardius zonarius barnardi*). *Genome Announc.* 2(4):e00708-14. doi:10.1128/genomeA.00708-14.

Copyright © 2014 Das et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Shane R. Raidal, shraidal@csu.edu.au, or Subir Sarker, ssarker@csu.edu.au.

Psittacine beak and feather disease (PBFD) is a common viral infection that occurs in a wide variety of psittacine birds, globally affecting >60 different species (1–4). The disease is caused by one of the smallest and simplest viruses belonging to the family *Circoviridae*, beak and feather disease virus (BFDV), a nonenveloped icosahedral virus with an approximately 2.0-kb circular single-stranded DNA (ssDNA) genome. The genome typically encompasses two major bidirectionally transcribed open reading frames (ORFs) encoding replication-associated protein (Rep) and capsid protein (Cap), with a potential stem-loop structure located between them (5–7). Clinically, BFDV infection exhibits as either sudden death or a chronic progressive illness characterized by symmetrical feather loss and deformity in beak and claw (5, 6, 8). Under natural conditions, horizontal transmission is likely the major route of spread for this virus (9). Here, we characterize the complete BFDV genome isolated from a wild Mallee ringneck parrot (*Barnardius zonarius barnardi*) in Australia. Feather samples collected from a Mallee ringneck parrot (ID, 14-1195/001; year of sampling, 2014; global positioning system [GPS] location, 31.9558°S 141.4651°E) were used as a source of genomic DNA, for which extraction of DNA was performed according to established protocols (10–12). The whole-genome sequence was amplified using the primers and PCR conditions developed in previous studies (13, 14). Briefly, the optimized reaction mixture contained 3 μ l extracted genomic DNA, 2.5 μ l of 10- μ l high-fidelity PCR buffer (Invitrogen), 1 μ l of 25 μ M (each) primers, 1 μ l of 50 mM MgSO₄, 4 μ l of 1.25 mM dinucleoside triphosphates (dNTPs), 1 U Platinum *Taq* DNA polymerase high-fidelity (Invitrogen), and distilled water added for a final volume of 25 μ l. The optimized PCR conditions were 95°C for 3 min, followed by 40 cycles of 95°C for 30 s, 57°C for 45 s, and 68°C for 2 min, and finally 68°C for 5 min. The amplified PCR products were TA cloned into pGEM-T vector (Promega, USA) and sequenced at the Australian Genome Research Facility (AGRF), Ltd. (Sydney, Australia). The sequenced contigs were assembled, and the entire BFDV genome was constructed using Geneious software (version 6.1.7).

The newly amplified BFDV genome (GenBank accession no. KJ866054) comprises 1,995 nucleotides (nt), with a G+C content of 53.3%. Similar to other BFDV genomes, the basic structure includes two major open reading frames (ORFs), ORF1 (870 nt) and ORF2 (708 nt), encoding a replication-associated protein (Rep) and the capsid protein (Cap), respectively. Phylogenetic analysis of this newly sequenced genome with all other BFDV genomes available on GenBank revealed the closest relationship (99% bootstrap support and 96% nucleotide sequence identity) with one of the Australian BFDV isolates obtained from a glossy black cockatoo (GenBank accession no. AF385408) (15). The overall nucleotide identity of this new isolate ranges from 92 to 96% compared with the BFDV genomes available on GenBank (16). This is the first report of a BFDV genome identification and characterization for this host species. This study documents the genomic characteristics and diversity of BFDV in a novel host (*B. zonarius barnardi*), which may facilitate further research on viral evolution and recombination events in this host species.

Nucleotide sequence accession number. The complete genome sequence of BFDV has been deposited at GenBank under the accession no. [KJ866054](https://www.ncbi.nlm.nih.gov/nuccore/KJ866054).

ACKNOWLEDGMENT

This research was supported by the Australian Research Council's Discovery Projects funding scheme (grant no. DP1095408).

REFERENCES

1. Bassami MR, Ypelaar I, Berryman D, Wilcox GE, Raidal SR. 2001. Genetic diversity of beak and feather disease virus detected in psittacine species in Australia. *Virology* 279:392–400. <http://dx.doi.org/10.1006/viro.2000.0847>.
2. Peters A, Patterson EI, Baker BG, Holdsworth M, Sarker S, Ghorashi SA, Raidal SR. 2014. Evidence of psittacine beak and feather disease virus spillover into wild critically endangered orange-bellied parrots (*Neophema chrysogaster*). *J. Wildl. Dis.* 50:288–296. <http://dx.doi.org/10.7589/2013-05-121>.
3. Sarker S, Patterson EI, Peters A, Baker GB, Forwood JK, Ghorashi SA, Holdsworth M, Baker R, Murray N, Raidal SR. 2014. Mutability dy-

- namics of an emergent single stranded DNA virus in a naïve host. PLoS One 9:e85370. <http://dx.doi.org/10.1371/journal.pone.0085370>.
4. Sarker S, Forwood JK, Ghorashi SA, McLelland D, Peters A, Raidal SR. 2014. Whole-genome sequence characterization of a beak and feather disease virus in a wild regent parrot (*Polytelis anthopeplus monarchoides*). Genome Announc. 2(1):e01243-13. <http://dx.doi.org/10.1128/genomeA.01243-13>.
 5. Ritchie BW, Niagro FD, Lukert PD, Steffens WL, Latimer KS. 1989. Characterization of a new virus from cockatoos with psittacine beak and feather disease. Virology 171:83–88. [http://dx.doi.org/10.1016/0042-6822\(89\)90513-8](http://dx.doi.org/10.1016/0042-6822(89)90513-8).
 6. Ritchie BW, Niagro FD, Latimer KS, Lukert PD, Steffens WL, Rakich PM, Pritchard N. 1990. Ultrastructural, protein composition, and antigenic comparison of psittacine beak and feather disease virus purified from 4 genera of psittacine birds. J. Wildl. Dis. 26:196–203. <http://dx.doi.org/10.7589/0090-3558-26.2.196>.
 7. Bassami MR, Berryman D, Wilcox GE, Raidal SR. 1998. Psittacine beak and feather disease virus nucleotide sequence analysis and its relationship to porcine circovirus, plant circoviruses, and chicken anaemia virus. Virology 249:453–459. <http://dx.doi.org/10.1006/viro.1998.9324>.
 8. Todd D. 2004. Avian circovirus diseases: lessons for the study of PMWS. Vet. Microbiol. 98:169–174. <http://dx.doi.org/10.1016/j.vet-mic.2003.10.010>.
 9. Raidal SR, Sabine M, Cross GM. 1993. Laboratory diagnosis of psittacine beak and feather disease by haemagglutination and haemagglutination inhibition. Aust. Vet. J. 70:133–137. <http://dx.doi.org/10.1111/j.1751-0813.1993.tb06104.x>.
 10. Ypelaar I, Bassami MR, Wilcox GE, Raidal SR. 1999. A universal polymerase chain reaction for the detection of psittacine beak and feather disease virus. Vet. Microbiol. 68:141–148. [http://dx.doi.org/10.1016/S0378-1135\(99\)00070-X](http://dx.doi.org/10.1016/S0378-1135(99)00070-X).
 11. Khalesi B, Bonne N, Stewart M, Sharp M, Raidal S. 2005. A comparison of haemagglutination, haemagglutination inhibition and PCR for the detection of psittacine beak and feather disease virus infection and a comparison of isolates obtained from lorriids. J. Gen. Virol. 86:3039–3046. <http://dx.doi.org/10.1099/vir.0.81275-0>.
 12. Bonne N, Clark P, Shearer P, Raidal S. 2008. Elimination of false-positive polymerase chain reaction results resulting from hole punch carryover contamination. J. Vet. Diagn. Invest. 20:60–63. <http://dx.doi.org/10.1177/104063870802000111>.
 13. Sarker S, Ghorashi SA, Forwood JK, Metz S, Raidal SR. 2013. Characterization of the complete genome sequence of a beak and feather disease virus from a Moluccan red lory (*Eos bornea*). Genome Announc. 1(6):e00844-13. <http://dx.doi.org/10.1128/genomeA.00844-13>.
 14. Sarker S, Ghorashi SA, Forwood JK, Raidal SR. 2013. Whole-genome sequences of two beak and feather disease viruses in the endangered swift parrot (*Lathamus discolor*). Genome Announc. 1(6):e00842-13. <http://dx.doi.org/10.1128/genomeA.00842-13>.
 15. Sarker S, Ghorashi SA, Forwood JK, Bent JS, Peters A, Raidal SR. 2014. Phylogeny of beak and feather disease virus in cockatoos demonstrates host generalism and multiple-variant infections within *Psittaciformes*. Virology 460–461:72–82. <http://dx.doi.org/10.1016/j.virol.2014.04.021>.
 16. Benson DA, Cavanaugh M, Clark K, Karsch-Mizrachi I, Lipman DJ, Ostell J, Sayers EW. 2013. GenBank. Nucleic Acids Res. 41:D36–D42. <http://dx.doi.org/10.1093/nar/gks1035>.