



Genome Sequence of a Beak and Feather Disease Virus from an Unusual Novel Host, Australian Boobook Owl (*Ninox boobook*)

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ABSTRACT The beak and feather disease virus (BFDV) is a pathogen of psittacine birds. BFDVs infecting nonpsittacine birds remain largely uncharacterized. We report the genome of a BFDV from a boobook owl (*Ninox boobook*), a nonpsittacine bird. The genome consisted of 1,993 bp containing two major bidirectionally transcribed open reading frames.

Beak and feather disease virus (BFDV) is a member of the family *Circoviridae*. Like other circoviruses, BFDV possesses a circular single-stranded, approximately 2.0-kb DNA genome that is encapsidated into a nonenveloped, spherical icosahedral virion (1), and it contains two bidirectionally transcribed genes. BFDV infection was thought to be restricted to Psittaciformes (2–6), but evidence of infection in distantly related Australian avian species was demonstrated in the rainbow bee-eater (*Merops ornatus*) (7), powerful owl (*Ninox strenua*) (8), and finches (9). Many other nonpsittacine birds are also likely susceptible to sporadic spillover infection (10). Here, we report the characterization of a BFDV genome in a nonpsittacine bird, the boobook owl (*Ninox boobook*), a species of Strigiformes.

Kidney tissue was collected from a dead boobook owl (*Ninox boobook*) submitted to the Avian, Reptile, and Exotic Pet Hospital of the University of Sydney, Camden Campus (34°0'10.61"S, 150°37'27.84"E), between December 2018 and April 2019. Total genomic DNA was extracted using a PureLink genomic DNA minikit (Invitrogen, CA). The library was prepared using Illumina DNA prep (Illumina, San Diego, CA), starting with 250 ng of DNA (11). The quality and quantity of the prepared library were assessed by the Australian Genome Research Facility, Melbourne, Australia, and the library was sequenced using the Illumina NovaSeq sequencing platform, generating 150-bp paired-end reads.

Sequencing data were analyzed as per established pipeline (12–15) using Geneious (version 10.2.2; Biomatters, New Zealand) and CLC Genomics Workbench (version 9.5.4). Briefly, a total of 37,770,262 raw reads were preprocessed to remove the Illumina adapter, ambiguous base calls, and poor-quality reads (trim using quality score, limit 0.05; trim ambiguous nucleotides up to 15 using CLC Genomics Workbench), followed by mapping against barn owl (*Tyto alba*) (16) and *Escherichia coli* (GenBank accession no. U00096) to remove nonviral DNA. A total of 37,612,162 trimmed and unmapped reads were used as input data for *de novo* default assembly in CLC Genomics Workbench (version 9.5.4). This resulted in the generation of a 1,993-bp BFDV genome with an average coverage of 39.61×. Annotation and circularization of the assembled genome were performed using in Geneious (version 10.2.2). All software was used with default parameters except where stated.

The genome has 1,993 bp, with a G+C content of 53.8%. A BLASTn analysis (under GenBank database parameters, maximum target sequences: 100) (17) of the sequenced

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BFDV genome in this study showed an overall 78.75 to 98.54% pairwise identity with other BFDV genomes, showing highest sequence similarity to BFDV (98.54%) from a little corella (*Cacatua sanguinea*) from Australia (GenBank accession no. [KY189060.1](https://doi.org/10.1093/jmm.0.000516)). In addition, when we compared a segment of capsid gene of BFDV sequenced (~407 bp) in this study, we found a 82.3% pairwise identity with a previously sequenced partial capsid gene from a boobook owl (*Ninox boobook*) (GenBank accession no. [KY410375.1](https://doi.org/10.1093/jmm.0.000516)) (10). The genome architecture of boobook BFDV sequence had characteristics of other *Circoviridae*, including two major open reading frames (ORFs): ORF1, encoding a replication-associated protein (293 amino acids), and ORF2, encoding the capsid protein (247 amino acids).

This study provides evidence of BFDV infection in an Australian boobook owl, as an unusual nonpsittacine host. Its apparent origin from a corella host suggests that the infection occurred from a contaminated nest hollow.

Data availability. The complete beak and feather disease virus genome sequence of *N. boobook* has been deposited in DDBJ/ENA/GenBank under accession no. [OL762453](https://doi.org/10.1093/jmm.0.000516). The version described in this paper is the first version, [OL762453.1](https://doi.org/10.1093/jmm.0.000516). The data that support the findings of this study are accessible via GenBank accession no. [OL762453](https://doi.org/10.1093/jmm.0.000516). The raw sequencing data from this study have been deposited in the NCBI Sequence Read Archive (SRA) under accession no. [SRR17163735](https://doi.org/10.1093/jmm.0.000516) (BioProject accession no. [PRJNA787018](https://doi.org/10.1093/jmm.0.000516)).

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REFERENCES

- Sarker S, Terron MC, Khandokar Y, Aragao D, Hardy JM, Radjainia M, Jimenez-Zaragoza M, de Pablo PJ, Coulibaly F, Luque D, Raidal SR, Forwood JK. 2016. Structural insights into the assembly and regulation of distinct viral capsid complexes. *Nat Commun* 7:13014. <https://doi.org/10.1038/ncomms13014>.
- Sarker S, Forwood JK, Ghorashi SA, Peters A, Raidal SR. 2015. Beak and feather disease virus genotypes in Australian parrots reveal flexible host-switching. *Aust Vet J* 93:471–475. <https://doi.org/10.1111/avj.12389>.
- Sarker S, Patterson EI, Peters A, Baker BG, Forwood JK, Ghorashi SA, Holdsworth M, Baker R, Murray N, Raidal SR. 2014. Mutability dynamics of an emergent single stranded DNA virus in a naïve host. *PLoS One* 9: e85370. <https://doi.org/10.1371/journal.pone.0085370>.
- Sarker S, Ghorashi SA, Forwood JK, Bent JS, Peters A, Raidal SR. 2014. Phylogeny of beak and feather disease virus in cockatoos demonstrates host generalism and multiple-variant infections within *Psittaciformes*. *Virology* 460:461:72–82. <https://doi.org/10.1016/j.virol.2014.04.021>.
- Sarker S, Ghorashi SA, Forwood JK, Raidal SR. 2013. Whole-genome sequences of two beak and feather disease viruses in the endangered swift parrot (*Lathamus discolor*). *Genome Announc* 1:e00842-13. <https://doi.org/10.1128/genomeA.00842-13>.
- Sarker S, Ghorashi SA, Forwood JK, Metz S, Raidal SR. 2013. Characterization of the complete genome sequence of a beak and feather disease virus from a moluccan red lory (*Eos bornea*). *Genome Announc* 1:e00844-13. <https://doi.org/10.1128/genomeA.00844-13>.
- Sarker S, Moylan KG, Ghorashi SA, Forwood JK, Peters A, Raidal SR. 2015. Evidence of a deep viral host switch event with beak and feather disease virus infection in rainbow bee-eaters (*Merops ornatus*). *Sci Rep* 5:14511. <https://doi.org/10.1038/srep14511>.
- Sarker S, Lloyd C, Forwood J, Raidal SR. 2016. Forensic genetic evidence of beak and feather disease virus infection in a powerful owl, *Ninox strenua*. *Emu* 116:71–74. <https://doi.org/10.1071/MU15063>.
- Circella E, Legretto M, Pugliese N, Caroli A, Bozzo G, Accogli G, Lavazza A, Camarda A. 2014. Psittacine beak and feather disease-like illness in Gouldian finches (*Chloebea gouldiae*). *Avian Dis* 58:482–487. <https://doi.org/10.1637/10745-121113Case.1>.
- Amery-Gale J, Marenda MS, Owens J, Eden PA, Browning GF, Devlin JM. 2017. A high prevalence of beak and feather disease virus in non-psittacine Australian birds. *J Med Microbiol* 66:1005–1013. <https://doi.org/10.1099/jmm.0.000516>.
- Sarker S. 2021. Metagenomic detection and characterisation of multiple viruses in apparently healthy Australian Neophema birds. *Sci Rep* 11: 20915. <https://doi.org/10.1038/s41598-021-00440-1>.
- Sarker S, Das S, Lavers JL, Hutton I, Helbig K, Imbery J, Upton C, Raidal SR. 2017. Genomic characterization of two novel pathogenic avipoxviruses isolated from pacific shearwaters (*Ardenna* spp.). *BMC Genomics* 18:298. <https://doi.org/10.1186/s12864-017-3680-z>.
- Athukorala A, Phalen DN, Das A, Helbig KJ, Forwood JK, Sarker S. 2021. Genomic characterisation of a highly divergent siadenovirus (psittacine siadenovirus F) from the critically endangered orange-bellied parrot (*Neophema chrysogaster*). *Viruses* 13:1714. <https://doi.org/10.3390/v13091714>.
- Sutherland M, Sarker S, Vaz PK, Legione AR, Devlin JM, Macwhirter PL, Whiteley PL, Raidal SR. 2019. Disease surveillance in wild Victorian caca-tuids reveals co-infection with multiple agents and detection of novel avian viruses. *Vet Microbiol* 235:257–264. <https://doi.org/10.1016/j.vetmic.2019.07.012>.
- Sarker S, Isberg RS, Moran LJ, Araujo DR, Elliott N, Melville L, Beddoe T, Helbig JK. 2019. Crocodilepox virus evolutionary genomics supports observed poxvirus infection dynamics on saltwater crocodile (*Crocodylus porosus*). *Viruses* 11:1116. <https://doi.org/10.3390/v11121116>.
- Zhang G, Li B, Li C, Gilbert MTP, Jarvis ED, The Avian Genome Consortium, Wang J. 2014. Genomic data of the barn owl (*Tyto alba*). *GigaScience Database*. <https://doi.org/10.5524/101039>.
- Benson DA, Cavanaugh M, Clark K, Karsch-Mizrachi I, Lipman DJ, Ostell J, Sayers EW. 2013. GenBank. *Nucleic Acids Res* 41:D36–D42. <https://doi.org/10.1093/nar/gks1195>.