Kinetics of Plasma Cell-Free DNA and Creatine Kinase in a Canine Model of Tissue Injury


Background: Cell-free DNA (cfDNA) comprises short, double-stranded circulating DNA sequences released from damaged cells. In people, cfDNA concentrations correlate well with disease severity and tissue damage. No reports are available regarding cfDNA kinetics in dogs.

Objectives/Hypothesis: Cell-free DNA will have a short biological half-life and would be able to stratify mild, moderate, and severe tissue injury. Our study aims were to determine the kinetics and biological half-life of cfDNA and to contrast them with those of creatine kinase (CK).

Animals: Three groups of 10 dogs undergoing open ovariohysterectomy, surgery for cranial cruciate ligament rupture (CCLR), or hemilaminectomy.

Methods: Plasma for cfDNA and CK analysis was collected at admission, at induction of anesthesia, postsurgery (time 0) and at 6, 12, 24, 36, 48, 60, and 72 hours after surgery.

Results: The biological half-life of plasma cfDNA and CK were 5.64 hours (95% confidence interval [CI] 4.36–7.98 hours) and 28.7 hours (CI 25.3–33.3 hours), respectively. In the hemilaminectomy group, cfDNA concentrations differed significantly from admission at 6–12 hours after surgery. Creatine kinase activity differed among the surgical groups and reached a peak 6 hours after surgery. In the ovariohysterectomy and CCLR groups, plasma CK activity 72 hours after surgery did not differ from admission activity of the ovariohysterectomy group. In contrast, in the hemilaminectomy group, plasma CK activity after 72 hours did not return to the ovariohysterectomy group admission activity.

Conclusions and Clinical Importance: Plasma CK activity has a longer biological half-life than previously thought. In contrast to plasma CK activity, cfDNA has a short half-life and could be a useful marker for peracute severe tissue injury.

Key words: Biological half-life; Cell-free DNA; Creatine kinase; Dog; Surgery.

Cell-free DNA (cfDNA) comprises short, double-stranded DNA sequences, circulating unbound in plasma. The source of cfDNA in the circulation still is largely unknown. Release of cfDNA from cells after apoptosis and necrosis, lysis of cells in the bloodstream, and spontaneous release from cells under stress are possible sources of cfDNA. In people, DNA hydrolysis by DNase I nuclease is the predominant pathway of cfDNA clearance from the blood, leading to a reported biological half-life of 157.6 minutes at 37°C.

In 1948, the presence of cfDNA was reported in human serum. Since then, increased concentrations of cfDNA in blood and body fluids have been linked to systemic lupus erythematosus and various cancers. Cell-free DNA concentrations correlated well with tumor burden and response to radiation therapy. Cell-free DNA has become widely studied in human medicine and has utility as a clinical noninvasive biomarker not only in cancer research but also in prenatal diagnostics, organ transplantation, and in several emergency conditions including stroke, myocardial infarction, sepsis, and severe trauma. Because of its short half-life, cfDNA is a reliable approximation of the current status of tissue injury and abates as resolution occurs.

In veterinary medicine, the focus of research on cfDNA has mainly centered on its potential value in the diagnosis, prognosis, and monitoring of the response to treatment of cancer in dogs. Dogs with mammary tumors or lymphoma have high concentrations of cfDNA in the blood, and the concentration of cfDNA is inversely correlated with remission time. The ability to detect cfDNA in canine blood led to the investigation of cfDNA in other disease states. A significant increase in the blood concentration of cfDNA was found in diseased dogs, as compared to healthy dogs, and a positive association between cfDNA concentration and disease severity and survival was observed. Both dogs with sepsis and those with

Abbreviations:

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
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<tbody>
<tr>
<td>CCLR</td>
<td>cranial cruciate ligament rupture</td>
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<tr>
<td>cfDNA</td>
<td>cell-free DNA</td>
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<tr>
<td>CK</td>
<td>creatine kinase</td>
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<tr>
<td>HL</td>
<td>hemilaminectomy</td>
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<tr>
<td>OVH</td>
<td>ovariohysterectomy</td>
</tr>
<tr>
<td>TPLO</td>
<td>tibial plateau leveling osteotomy</td>
</tr>
<tr>
<td>TTA</td>
<td>tibial tuberosity advancement</td>
</tr>
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</table>
moderate-to-severe trauma had significantly increased cfDNA concentrations compared to healthy dogs,22 and cfDNA concentration was associated with death in dogs with immune-mediated hemolytic anemia.23 A short half-life and a good correlation with disease severity would render cfDNA a useful tool to quantify the extent of tissue injury. Based on current knowledge, we hypothesized that, in dogs, cfDNA would have a short biological half-life and its concentration would be able to stratify mild, moderate, and severe tissue injury. Accordingly, the aims of our study were to determine the kinetics of cfDNA and its biological half-life. To contrast with cfDNA, we estimated similar variables for creatine kinase activity. We chose to approach the hypothesis and aims using a model of controlled tissue injury in dogs that underwent 3 types of surgeries associated with mild, moderate, and severe tissue injury.

Materials and Methods

Animals

We collected plasma samples from dogs presented to the Massey University Veterinary Teaching Hospital between October 2015 and April 2017. Inclusion criteria were client-owned dogs presented for cranial cruciate ligament rupture (CCLR) surgically managed by tibial tuberosity advancement (TTA) or tibial plateau leveling osteotomy (TPLO) surgical techniques, thoracolumbar disk disease decompressed by hemilaminectomy surgery (HL), and bitches presented for an elective, open ovariohysterectomy surgery (OVH). The surgeries were stratified according to perceived tissue injury (mild, moderate, and severe) on the basis of the extent of dissection required for each procedure. In an open OVH, the incision is primarily through the linear alba although there can be some injury to the rectus abdominis muscle and crushing and stretching of uterine muscle. In a TPLO, there is more extensive dissection with elevation of the pes anserinus, cranial tibial, and popliteal muscles in addition to an arthrotomy. The HL was considered the most severe trauma because of the need to extensively elevate and transect epaxial musculature and use Gelpi retractors for extended time periods during surgery. Enrollment of 10 patients for each category in the study gave a total of 30 patients. Table 1 presents the descriptive characteristics of the dogs included in the study. The Massey University Animal Ethics Committee approved the study (MUAEC protocol 15/50) and enrollment of patients required informed, signed client consent.

Sample Collection

Each dog enrolled in the study had a 2 mL sample of whole blood in ethylenediaminetetraacetic acid (EDTA) collected by jugular venipuncture upon arrival (admission sample). A 16 G, 20 cm long indwelling IV catheter was placed in the lateral saphenous vein after induction of anesthesia for the intended surgical procedure. For each blood sample, we irrigated the indwelling IV catheter with 5 mL 0.9% sodium chloride and aspirated back 1 mL of whole blood that was discarded. Plasma was harvested from an additional 2 mL of whole blood that was placed in an EDTA tube. At the end of blood collection, the indwelling IV catheter was irrigated with 0.9% sodium chloride. Collection of

Table 1. The descriptive characteristics of the dogs included in the study.

<table>
<thead>
<tr>
<th></th>
<th>HL</th>
<th>CCLR</th>
<th>OVH</th>
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<tbody>
<tr>
<td>Age (months)*</td>
<td>42 (9–128)</td>
<td>40 (18–96)</td>
<td>69 (5–144)</td>
</tr>
<tr>
<td>Sex</td>
<td>M</td>
<td>7</td>
<td>5</td>
</tr>
<tr>
<td></td>
<td>F</td>
<td>3</td>
<td>5</td>
</tr>
<tr>
<td>Neuter</td>
<td>E</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>N</td>
<td>9</td>
<td>8</td>
</tr>
<tr>
<td>Weight (kg)**</td>
<td>25 (7.7–43.5)</td>
<td>36 (8–45.6)</td>
<td>17.5 (7.7–33.4)</td>
</tr>
<tr>
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<td>1</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>Cocker Spaniel</td>
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<td>1</td>
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<tr>
<td></td>
<td>Dachshund</td>
<td>4</td>
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</tr>
<tr>
<td></td>
<td>Dogue de Bordeaux</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>German Shorthaired Pointer</td>
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<td>1</td>
</tr>
<tr>
<td></td>
<td>Golden Retriever</td>
<td>1</td>
<td>1</td>
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<tr>
<td></td>
<td>Harrier</td>
<td>5</td>
<td>1</td>
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<tr>
<td></td>
<td>New Zealand Heading Dog</td>
<td>1</td>
<td>1</td>
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<tr>
<td></td>
<td>Huntaway</td>
<td>1</td>
<td>1</td>
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<tr>
<td></td>
<td>Labrador Retriever</td>
<td>1</td>
<td>4</td>
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<td></td>
<td>Lhasa Apso</td>
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<td>Mastiff</td>
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<tr>
<td></td>
<td>Miniature Schnauzer</td>
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<td></td>
<td>Pekingese</td>
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<tr>
<td></td>
<td>Rottweiler</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>Staffordshire Bull Terrier</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>Toy Poodle</td>
<td>2</td>
<td>2</td>
</tr>
</tbody>
</table>

HL, hemilaminectomy; CCLR, surgeries for cranial cruciate ligament rupture; OVH, open ovariohysterectomy; M, male; F, female; E, entire; N, neuter.

*Age and weight are presented as median (range).

**There is a statistically significant difference between the CCLR and OVH groups ($P < 0.01$).

**There were statistically significant differences between groups ($P < 0.01$).
blood samples took place at the time of admission, after induction of anesthesia, immediately after surgery ended and then at 6, 12, 24, 36, 48, 60, and 72 hours after surgery. At 72 hours, we also collected a sample from the bag of 0.9% sodium chloride that had been used to irrigate the catheter throughout the sampling period. This sample served as a negative control to ensure that there was no DNA contamination in the 0.9% sodium chloride before sampling.

**Sample Handling**

Centrifugation of whole blood samples at 3,000 × g for 15 minutes at 4°C facilitated harvesting the plasma. Equal volumes of the harvested plasma were stored at −80°C until the time of analysis.

**Cell-free DNA Analysis**

We used the Qubit dsDNA HS Assay Kit and Qubit 2.0 fluorimeter and quantified the plasma concentration of cell-free DNA (cfDNA) as previously described. Plasma samples were thawed in batches of 15–50 samples, and a volume of 20 μL was used for the analysis. The Qubit assay utilizes a dye that fluoresces with a higher intensity when bound to double-strand DNA (dsDNA), and the recorded amount of fluorescence is proportional to the amount of dsDNA in the sample. The dilution algorithm provided by the manufacturer within the Qubit 2.0 determined the concentration of the cfDNA. Calibration of the Qubit 2.0 with the provided standards preceded each run. We assayed a single sample with a previously measured concentration of cfDNA that had been previously separated into aliquots and stored at −20°C with every batch of samples as an interassay control. The coefficient of variation determined the intra-assay precision on 22 samples run in triplicates.

**Creatine Kinase Analysis**

A commercial veterinary diagnostic laboratory measured the activity of plasma creatine kinase (CK) on a Roche/Hitachi analyzer.

**Statistical Analysis**

A priori power sample size analysis was performed by G*Power version 3.1.9.8. The analysis indicated that 9 dogs in each group would suffice to detect a difference of 20% in plasma CK activity on repeated measures of plasma CK activity on the same dog with a power of 0.8 and alpha probability error of 0.05, assuming that the correlation for the repeated measurements on the same dog, as described above, did not contribute significantly enough to the model to be included in the final model ($P > 0.05$).

The dependent variable CK was analyzed with the same model as cfDNA but also included the fixed effects of “age” and “weight” with the same distributional properties for the residuals of repeated measures on the same dog, as described above. “Sex” effect did not contribute significantly enough to the model to be included in the final model ($P > 0.05$).

The lsmeans() function of the lsmeans package was used to obtain the least squares means and standard errors and was used for Tukey’s pairwise mean comparisons.

To calculate the biological half-life of plasma cfDNA and CK, we used the lm4 package lm() function to perform simple linear regression on the log-transformed down slope of the least squares means curves of plasma cfDNA and CK. The half-life was calculated from the following formula: $t_{1/2} = \frac{\log(1/2)}{\text{coefficient of regression slope}}$.

The correlation between plasma cfDNA and CK was evaluated by the Spearman’s rank correlation $\rho$ and Kendall’s rank correlation $\tau$.

**Table 2.** Least squares mean of plasma cell-free DNA (μg/L) in 30 dogs, stratified by the perceived severity of tissue trauma at surgery (10 dogs per group).

<table>
<thead>
<tr>
<th>Time</th>
<th>Lsmean</th>
<th>SE</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>HL</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Admission</td>
<td>648</td>
<td>144</td>
<td>363–932</td>
</tr>
<tr>
<td>Induction</td>
<td>570</td>
<td>144</td>
<td>286–854</td>
</tr>
<tr>
<td>Post-Op</td>
<td>734</td>
<td>138</td>
<td>462–1,007</td>
</tr>
<tr>
<td>6</td>
<td>1,458</td>
<td>138</td>
<td>1,185–1,730</td>
</tr>
<tr>
<td>12</td>
<td>1,446</td>
<td>138</td>
<td>1,174–1,719</td>
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<tr>
<td>24</td>
<td>1,074</td>
<td>144</td>
<td>789–1,358</td>
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<td>36</td>
<td>770</td>
<td>144</td>
<td>485–1,055</td>
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<tr>
<td>48</td>
<td>645</td>
<td>144</td>
<td>360–929</td>
</tr>
<tr>
<td>60</td>
<td>675</td>
<td>144</td>
<td>390–960</td>
</tr>
<tr>
<td>72</td>
<td>607</td>
<td>144</td>
<td>322–892</td>
</tr>
<tr>
<td>CCLR</td>
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<tr>
<td>Admission</td>
<td>776</td>
<td>138</td>
<td>503–1,048</td>
</tr>
<tr>
<td>Induction</td>
<td>749</td>
<td>138</td>
<td>477–1,022</td>
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<td>Post-Op</td>
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<td>486–1,031</td>
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<td>756</td>
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<td>819</td>
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<td>856</td>
<td>138</td>
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<tr>
<td>36</td>
<td>873</td>
<td>138</td>
<td>601–1,146</td>
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<td>48</td>
<td>859</td>
<td>138</td>
<td>587–1,132</td>
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<tr>
<td>60</td>
<td>735</td>
<td>138</td>
<td>463–1,008</td>
</tr>
<tr>
<td>72</td>
<td>835</td>
<td>138</td>
<td>563–1,108</td>
</tr>
<tr>
<td>OVH</td>
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<td></td>
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<tr>
<td>Admission</td>
<td>611</td>
<td>138</td>
<td>338–883</td>
</tr>
<tr>
<td>Induction</td>
<td>623</td>
<td>138</td>
<td>350–895</td>
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<tr>
<td>Post-Op</td>
<td>675</td>
<td>138</td>
<td>400–945</td>
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<tr>
<td>6</td>
<td>787</td>
<td>138</td>
<td>514–1,059</td>
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<tr>
<td>12</td>
<td>794</td>
<td>138</td>
<td>521–1,066</td>
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<td>24</td>
<td>789</td>
<td>138</td>
<td>516–1,061</td>
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<td>36</td>
<td>773</td>
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<td>501–1,045</td>
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<td>723</td>
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<td>451–996</td>
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<td>688</td>
<td>138</td>
<td>416–960</td>
</tr>
<tr>
<td>72</td>
<td>695</td>
<td>138</td>
<td>422–967</td>
</tr>
</tbody>
</table>

HL, hemilaminectomy; CCLR, surgery for cranial cruciate ligament rupture; OVH, open ovariohysterectomy; Lsmean, least squares mean; SE, standard error; CI, confidence interval.
Results

**Plasma Cell-free DNA**

The least squares means of plasma cell-free DNA (cfDNA) concentrations at the various time points of the study are shown in Table 2.

**Between-group Comparisons**

Plasma cfDNA concentrations (μg/L) were significantly higher in the hemilaminectomy (HL) group, compared to both the CCLR and open ovariohysterectomy (OVH) groups at 6 hours (1,458 ± 138 versus 756 ± 138 and 787 ± 138; \( P = 0.001 \) and \( P = 0.002 \)) and 12 hours (1,446 ± 138 versus 819 ± 138 and 794 ± 138; \( P = 0.004 \) and \( P = 0.003 \); Fig 1). No significant differences were identified between the CCLR and open OVH groups at any time.

**Within-group Comparisons**

In the HL group, plasma cfDNA reached a peak at 6 hours through 12 hours and rapidly returned to admission concentrations by 72 hours. In contrast, in the open OVH and CCLR groups, plasma cfDNA reached a shallow peak at 12 and 36 hours, respectively.

In the HL group, significant differences were observed between times 6 hours through 12 hours compared to the other time points (other than 24 hours postoperatively; Fig 1). Within the CCLR and open OVH groups, there were no differences between any of the time points.

The calculated biological half-life of plasma cfDNA was 5.64 hours (CI 95, 4.36–7.98 hours).

**Plasma CK**

The geometric means are shown in Table 3, which were derived from back transformation of the least squares means of log plasma CK at the various time points of the study.

**Between-group Comparisons**

Overall, a significant difference was identified between the HL and open OVH groups (estimate 1.03, \( P = 0.013 \)).

A significant difference in the log plasma CK activity was observed between the HL and open OVH groups at 6 hours through 72 hours (estimate 1.182, \( P = 0.017 \); estimate 1.178, \( P = 0.017 \); estimate 1.536, \( P = 0.002 \); estimate 1.284, \( P = 0.010 \); estimate 1.302, \( P = 0.008 \); estimate 1.063, \( P = 0.037 \); estimate 1.129, \( P = 0.025 \), respectively).

**Within-group Comparisons**

In the HL and open OVH groups, plasma CK reached a peak at 6 hours in comparison with the CCLR group in which plasma CK reached a peak at 12 hours. In the HL group, plasma CK did not return to admission activity by 72 hours. In contrast, in the CCLR and open OVH groups, plasma CK reached admission activity at 72 hours.

Within the HL and CCLR groups, the period of 6 hours through 24 hours was significantly different
Correlation between cfDNA and CK

Moderate-to-strong correlation was observed between plasma CK and cfDNA for all surgeries and time intervals, for all surgeries at the down slope from 6 to 72 hours, and for the HL surgery at the down slope from 6 to 72 hours (Table 4).

Discussion

The aims of our study were to determine the kinetics of plasma cfDNA and its biological half-life. The results indicate that after substantial tissue damage, plasma cfDNA concentrations increased sharply at 6 hours for a period of approximately 6 hours and then decreased abruptly to baseline concentrations. Plasma cfDNA did not increase significantly when mild or moderate tissue damage occurred. The lack of increase in cfDNA post-surgery in the CCLR and OVH groups also could be a consequence of the different trauma type experienced during these surgeries, which is mainly traction and compression of soft tissues as opposed to HL where there is substantial blunt dissection and retraction of the muscle by Gelpi retractors to expose the surgical site. Alternatively, the differences between the CCLR and OVH groups and the HL group could be secondary to the proportion of injured tissue. The fact that cfDNA plasma concentrations were increased for only a short period of time implies that in dogs, plasma cfDNA is a marker of substantial peracute tissue injury. Also, it is less sensitive to milder forms of tissue injury, and it is less sensitive in the later phases after an acute insult, as was apparent in our results in which cfDNA plasma concentrations returned to baseline after 12 hours (Fig 1).

Our results are in agreement with the previous studies indicating that plasma cfDNA is a marker associated with peracute inflammation. In our study, plasma cfDNA did not differ among groups in the immediate postoperative period even though the times it took to complete HL were different from the CCLR and TTA groups. We speculate that there is a temporal association between the early phase of inflammation and the increase in plasma cfDNA concentration at 6–12 hours. In the early phase of inflammation, inflammatory cytokines increase vascular permeability because of activation of the endothelium. Therefore, cfDNA that leaked from injured cells into the interstitial space could diffuse rapidly along its concentration gradient and its plasma concentration would increase. Thus, we argue that cfDNA is a good indicator of early inflammation and tissue damage. In contrast, we expect that in chronic inflammation, despite increased permeability of the inflamed vascular bed, there would be insufficient amounts of cfDNA to diffuse into the circulation because of fibrosis and scarring. Hence, we postulate that chronic inflammation would not be associated with high plasma cfDNA concentrations. This hypothesis is supported by a recent meta-analysis that compared plasma cfDNA among 4 groups: healthy controls, acute inflammation, chronic inflammation, and acute infections. In the metaanalysis, the cfDNA concentration in chronic inflammatory conditions was substantially lower than in acute inflammation and infection, yet still higher than the concentration in healthy controls. A similar trend was recently identified in dogs. In that study, small but statistically significant differences were observed between healthy control dogs when compared to dogs with chronic and acute disease conditions. Dogs with acute disease conditions had significantly higher plasma cfDNA concentrations than did dogs with chronic disease conditions. The authors defined chronic

Table 3. Geometric mean (GM) of plasma creatine kinase activity (IU/L) in 30 dogs, stratified by the perceived severity of tissue trauma at surgery (10 dogs per group).

<table>
<thead>
<tr>
<th>Time</th>
<th>GM</th>
<th>95% CI</th>
</tr>
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<tbody>
<tr>
<td>HL</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Admission</td>
<td>229.5</td>
<td>125.8–419</td>
</tr>
<tr>
<td>Induction</td>
<td>202.6</td>
<td>111–370</td>
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<td>Post-Op</td>
<td>373.5</td>
<td>207.5–672</td>
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<td>1192.9</td>
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<td>1129.7</td>
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<td>24</td>
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<td>36</td>
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<td>48</td>
<td>497.4</td>
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<td>60</td>
<td>315.1</td>
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<td>72</td>
<td>298.9</td>
<td>163.4–547</td>
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<td>CCLR</td>
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<td>Admission</td>
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<td>Induction</td>
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<td>110.1–361</td>
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<td>60.4–201</td>
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<td>OVH</td>
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<td>Induction</td>
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<td>76.9–252</td>
</tr>
<tr>
<td>Post-Op</td>
<td>195.8</td>
<td>108.1–355</td>
</tr>
<tr>
<td>6</td>
<td>365.8</td>
<td>202–665</td>
</tr>
<tr>
<td>12</td>
<td>347.9</td>
<td>191.6–632</td>
</tr>
<tr>
<td>24</td>
<td>231.2</td>
<td>127.4–419</td>
</tr>
<tr>
<td>36</td>
<td>159</td>
<td>87.8–288</td>
</tr>
<tr>
<td>48</td>
<td>135.3</td>
<td>74.6–245</td>
</tr>
<tr>
<td>60</td>
<td>108.8</td>
<td>59.8–198</td>
</tr>
<tr>
<td>72</td>
<td>96.7</td>
<td>53.2–176</td>
</tr>
</tbody>
</table>

HL, hemilaminectomy; CCLR, surgeries for cranial cruciate ligament rupture; OVH, open ovariohysterectomy; GM, geometric mean; CI, confidence interval.
versus acute disease based on the history but did not report the associate conditions.

Previous studies have characterized the kinetics of plasma CK. In 1 study, the 3 canine isoforms of CK were purified and their half-lives determined after an IV injection. This pharmacokinetic study indicated that the half-life of plasma CK in dogs was 119.5 minutes. We calculated the time plasma CK would reach 50% of its peak activity after surgery. In marked contrast to the previously described study, we found that for all surgeries, the biological half-life of plasma CK was 28.7 hours (CI 25.3–33.3 hours). This estimated biological half-life should be considered when monitoring trends in plasma CK activity. The estimated biological half-life is different than the pharmacologic half-life. The biological half-life takes into account CK released from the primary insult and CK that further leaks from myofiber damage induced by local ischemia, thrombosis, and inflammation. Thus, in a clinical setting, the biological half-life would better predict the decrease in serum CK activity after an isolated insult. Deviation from that prediction should alert the clinician to possible ongoing muscle damage. In dogs, plasma CK activities are assumed to be proportional to the extent of muscle injury. This assumption is based on a single pharmacokinetic study in which the supernatants of dog muscle homogenates were injected IV and IM into 6 dogs. In another study, serum CK activity was measured over 48 hours after HL and OVH. Similar to our study, clear stratification in serum CK activity was observed according to the type of surgery. In that study, serum CK activity after HL did not return to baseline at 48 hours. We followed plasma CK activity longer than in the previous study and found that, in the HL group, plasma CK activity did not return to baseline even after 72 hours. Both our study and the previous study indicate that plasma CK activity is proportional to the extent of muscle injury and that the biological half-life is approximately 28 hours.

To our knowledge, ours is the first study to estimate the time it takes plasma cfDNA to reach 50% of its peak concentration in a clinical setting. We found that the biological half-life of plasma cfDNA was 5.64 hours. The short biological half-life of plasma cfDNA combined with the long half-life of plasma CK could permit differentiation of progressive tissue injury from acute nonprogressive injury. For example, after substantial trauma, progressive tissue injury might result from thrombosis or compartment syndrome. Monitoring plasma CK activity might not be useful because of the duration of time that it would take to return to baseline. In that scenario, co-measurement of plasma cfDNA along with CK activity would indicate if there is substantial ongoing tissue injury.

Table 4. Correlation coefficients between plasma creatine kinase and cell-free DNA.

<table>
<thead>
<tr>
<th></th>
<th>Spearman’s Rank Correlation ρ</th>
<th>P Value</th>
<th>Kendall’s Rank Correlation τ</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>All surgery</td>
<td>0.53</td>
<td>0.003</td>
<td>0.38</td>
<td>0.003</td>
</tr>
<tr>
<td>All surgery</td>
<td>0.47</td>
<td>0.040</td>
<td>0.35</td>
<td>0.030</td>
</tr>
<tr>
<td>6–72 hours</td>
<td>0.96</td>
<td>0.003</td>
<td>0.9</td>
<td>0.003</td>
</tr>
<tr>
<td>HL surgery</td>
<td>0.96</td>
<td>0.003</td>
<td>0.9</td>
<td>0.003</td>
</tr>
</tbody>
</table>

HL, hemilaminectomy.

Fig. 2. Least squares means of log plasma creatine kinase. Different letters for the same number indicate $P < 0.05$ within the group. “∗” indicates $P < 0.05$ between hemilaminectomy and the ovariohysterectomy group per time period. “#” indicates $P < 0.05$ between hemilaminectomy and the cranial cruciate ligament rupture surgery group per time period. Post-op, postoperative period; HL, hemilaminectomy; CCLR, surgeries for cranial cruciate ligament rupture; OVH, open ovariohysterectomy.
In conclusion, plasma cfDNA has a short biological half-life, and plasma CK has a longer biological half-life than previously thought. Plasma cfDNA could be a useful marker for peracute severe tissue injury. Combining measurement of plasma cfDNA with CK activity may allow differentiation of progressive tissue injury from acute nonprogressive injury.

**Author Contributions**

Gal A, Wilson II, and Burchell RK formulated the hypothesis, designed the study, collected samples, analyzed the data, and wrote the manuscript. Worth AJ, Burton SE, Gedye KR, Clark KJ, Crosse KR, Jack M, Odom TF, De Grey SJ, McGlade KMS, Tomlin SC, and Lopez-Villalobos N contributed to the development of the hypothesis, design of the study, collection and processing of the samples, analysis of the data, and critically contributed to the writing of the manuscript.

**Footnotes**

* Qubit dsDNA HS Assay Kit and Qubit 2.0 fluorimeter, Life Sciences, Carlsbad, CA, USA
* IDEXX New Zealand, Palmerston North, New Zealand
* Cobas CK Roche Diagnostics GmbH, Mannheim, Germany

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**Conflict of Interest Declaration:** Authors declare no conflict of interest.

**Off-label Antimicrobial Declaration:** Authors declare no off-label use of antimicrobials.

**References**


