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Host anemone size as a determinant of social group size and structure in the orange clownfish (*Amphiprion percula*)

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ABSTRACT

The size and structure of social groups of animals can be governed by a range of ecological factors and behavioral interactions. In small, highly site-attached coral reef fishes, group size is often constrained by the size of the habitat patch they are restricted to. However, group size may also be influenced by changes in abundance along important environmental gradients, such as depth or distance offshore. In addition, the body size and sex structure within social groups can be determined by the size of the habitat patch and the dominance relationships among group members. Here we examined the roles of ecological factors and behavioral interactions in governing group size and structure in the orange clownfish, Amphiprion percula, on inshore reefs in Kimbe Bay, Papua New Guinea. We quantified relationships between ecological variables (anemone size, depth, and distance from shore) and social group variables (group size, and total body length of the three largest individuals (ranks 1, 2, and 3)). Anemone size explained the greatest amount of variation in group variables, with strong, positive relationships between anemone surface area and group size, and total length of individuals ranked 1, 2, and 3. Group structure was also weakly correlated with increasing depth and distance from shore, most likely through the indirect effects of these environmental gradients on anemone size. Variation in group size and the lengths of ranks 2 and 3 were all closely related to the length of rank 1. Path analysis indicated that anemone size has a strong direct effect on the length of rank 1. In turn, the length of rank 1 directly affects the size of the subordinate individuals and indirectly affects the group size through its influence on subordinates. Hence, anemone size directly and indirectly controls social group size and structure in this space-limited fish species. It is also likely that anemonefish have feedback effects on anemone size, although this could not be differentiated in the path analysis.

Subjects Animal Behavior, Ecology, Marine Biology, Environmental ImpactsKeywords Papua New Guinea, Social hierarchy, Coral reefs, Anemonefish, Depth, Patch size, Pomacentridae, Amphiprion percula, Heteractis magnifica

INTRODUCTION

Animals often form social groups to benefit from cooperation in finding food, avoiding predators, and reproducing (*Wilson, 1975; Krebs & Davies, 1981; Alcock, 2001*).

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There are numerous ecological factors that influence optimal group size, including variation in resource availability and predation risk (*Cody*, 1971; *Hamilton*, 1971; *Caraco & Wolf*, 1975; *Caraco, Martindale & Whitlam*, 1980; *Markham et al.*, 2015). Group size can also be determined by the size and quality of the habitat patches occupied—the well-supported "ecological constraints" model (*Crook*, 1970; *Chapman & Chapman*, 2000; *Smith & Chapman*, 2007). As the distribution and abundance of most animals vary along key environmental gradients such as temperature, light, rainfall, and depth (*Krebs*, 1972; *Ricklefs & Miller*, 1999), group size in social animals is also likely to vary as a consequence. Behavioral interactions among group members can also influence group size and the reproductive status and body size of group members, with competing individuals evicted from groups or prevented from joining them (*Wilson*, 1975; *Gilmour*, *DiBattista & Thomas*, 2005; *Wong et al.*, 2007; *Fitzpatrick et al.*, 2008). Given the multitude of factors that can affect a group's size and structure, the relative importance of different factors and the relationships among direct and indirect effects is not always known.

Many small, specialized, and highly site attached coral reef fishes form small social groups associated with discrete patches of habitat, such as corals, sponges, or anemones (Fautin & Allen, 1992; Munday, Jones & Caley, 1997; Wong, Munday & Jones, 2005; D'Aloia, Majoris & Buston, 2011). Individuals are confined to the habitat patch, which provides shelter from predators, access to specialized food resources, and breeding sites. Habitat patch size and quality are likely to represent ecological constraints on group size, with several studies demonstrating that group size can be correlated with the size of the habitat patch (Wong, Munday & Jones, 2005; Thompson, Munday & Jones, 2007; D'Aloia, Majoris & Buston, 2011; Noonan, Jones & Pratchett, 2012). Because such specialized species live in relatively small social groups, their behavior is frequently characterized by strong social hierarchies, with the dominant member of a group often aggressively regulating the status, size, sex, and reproductive status of subordinates (Fricke, 1979; Shapiro, 1981; Buston, 2003a; Hobbs, Munday & Jones, 2004). Dominant individuals may control group size by evicting subordinates and preventing recruitment to the habitat patch (Elliott, Elliott & Mariscal, 1995; Buston, 2003b). Subordinates may restrict their food intake to prevent potential conflict with dominant individuals and avoid eviction, thereby affecting their body size (Wong et al., 2008). However, the different roles of patch size and behavioral interactions in determining group size and structure are often unknown.

The distribution and abundance of coral reef fishes often varies along major environmental gradients on coral reefs, such as distance offshore (*Williams, 1991*; *Gust, Choat & McCormick, 2001*; *Pittman & Brown, 2011*; *MacDonald, Bridge & Jones, 2016*) and depth (*Choat & Bellwood, 1985*; *McGehee, 1994*; *Srinivasan, 2003*; *Jankowski, Gardiner & Jones, 2015*; *Jankowski, Graham & Jones, 2015*; *MacDonald, Bridge & Jones, 2016*; *Smallhorn-West et al., 2017*). Group size is also likely to vary along the same gradients, with larger groups located in areas of highest abundance. The morphology of habitat patches, such as corals, sponges, and anemones, may also vary along depth or turbidity gradients (D'Aloia, Majoris & Buston, 2011). Coral colony size can increase with depth in response to declining light intensity, with more foliose and open morphologies persisting in deeper habitats to maximize energy acquisition (*Hoogenboom*, *Connolly & Anthony*, 2008). Anemones also symbiotically associate with zooxanthellae and likely maximize surface area at greater depths to increase light capture for photosynthesis (*Sebens & DeRiemer*, 1977). However, the relative effects of distance offshore or depth on habitat patch size and group size and structure in specialized reef fish have not been assessed.

Anemonefishes represent a tractable model system for examining the interrelationships among habitat patch size, location, and the size and structure of social groups. They live in an obligate association with anemones, a small and discrete habitat where fish must derive all critical resources. Larger anemones are likely to provide more shelter, as well as an increased area for reproduction and laying eggs. While the specific species of the anemonefish and the host play an important role in determining the range of potential fish body lengths and group sizes (Fautin & Allen, 1992; Huebner et al., 2012), group size does appear to be positively related to anemone size in a number of species (Fricke, 1980; Fautin, 1992; Mitchell & Dill, 2005). The anemonefish simultaneously provides reciprocal benefits to the anemone, which may reinforce relationships between anemone size, and group size, and group structure. For example, anemonefish provide protection against anemone predators, such as butterflyfish, through aggressive behavior (Godwin & Fautin, 1992; Porat & Chadwick-Furman, 2004). Anemonefish have also been shown to transfer carbon and nitrogen, at least partially in the form of ammonia, to their host and the host's symbiodinium (Cleveland, Verde & Lee, 2010). The transfer of ammonia from fish to host results in increased the zooxanthellae concentrations and tissue regeneration potential in anemones (Porat & Chadwick-Furman, 2005; Roopin & Chadwick, 2009). Nocturnal anemonefish movement aerates the anemone, potentially facilitating a range of benefits, including gas exchange and debris removal (Szczebak et al., 2013). Overall, anemones hosting anemonefish exhibit greater growth, more frequent asexual reproduction, and lower mortality than those without resident anemonefish (Holbrook & Schmitt, 2005; Frisch et al., 2016).

Anemonefishes are protandrous hermaphrodites, the largest individual (rank 1) in the group generally being the female and the second largest (rank 2) the breeding male, the latter undergoing sex change following the death of the female (*Fricke & Fricke, 1977*; *Fricke, 1979, 1983*; *Buston & Elith, 2011*). In anemonefish groups, reproduction is restricted to the two dominant individuals regardless of the group size. There is a strong dominance and size hierarchy between all members of the group, with non-breeders essentially in a queue for reproductive status (*Buston, 2003a*). The body size structure of groups may also be determined by anemone size (*Mitchell & Dill, 2005*), although *Buston* (*2003a*) determined that anemone diameter had no effect on the body lengths of subordinates in *Amphiprion percula*. Group size may be directly influenced by density dependent processes impacting the whole group or may be dictated by the size of the largest individual (*Buston, 2003c*; *Mitchell & Dill, 2005*). However, cause and effect can be difficult to assess when there is strong covariation between anemone size, group size, and body size. To date, few studies have examined the potential effects of both environmental and social control on relationships among group size and body size structure in clownfishes.

The aim of this study was to quantify relationships among anemone size, depth, distance offshore, group size, and body size structure in the orange clownfish, *A. percula* on inshore reefs in Kimbe Bay, Papua New Guinea. In addition, we conducted a path analysis to determine whether anemone size has a direct effect on group size or whether anemone size controls the size of the largest individual, who then dictates the size and structure of the social group. The following specific questions were addressed: (1) Are group size and structure positively related to anemone size? (2) Does anemone depth and/or distance offshore explain variation in group size and structure, and can this be attributed to spatial variation in anemone size? (3) Are the group size, and the size of rank 2 and rank 3 individuals related to the size of rank 1? (4) Does anemone size directly determine group size (and the size of subordinates) or is this mediated by the size of the rank 1 individual?

MATERIALS AND METHODS

Study site and species

This study was undertaken in April 2016 on the inshore coral reefs near the Mahonia Na Dari Research and Conservation Centre, near the town of Kimbe, in Kimbe Bay, Papua New Guinea ($05^{\circ}26'S$, $150^{\circ}52'E$) (Figs. 1A and 1B). It focused on an area of the fringing reef approximately five km long as well as 19 discrete emergent reefs ranging a distance of 0.2–1.4 km from the coastline (Fig. 1C). Kimbe Bay experiences a distinct wet season from December to January, and a windy season from June to July, but there is little variation in temperature year-round (*Srinivasan, 2003*).

This study focused on the orange clownfish, *A. percula*, which is associated primarily with *Heteractis magnifica* and *Stichodactlya gigantea* anemones in this region, usually at depths shallower than 12 m (*Almany et al., 2007; Planes, Jones & Thorrold, 2009*). Due to the relatively low numbers of *S. gigantea* on coastal reefs, this study focused primarily on *A. percula* groups associated with *H. magnifica. A. percula* lives in small social groups with a strong dominance hierarchy, whereby individuals of sequential rank follow a fixed size ratio (*Buston, 2003a, 2003b, 2003c*). Reproduction in *A. percula* is also restricted to the two highest rank individuals: the highest rank generally being the female and the second highest rank generally being the male, if they have achieved the size required for sexual maturation (*Buston & Elith, 2011*). Recruitment to the group is restricted until the lowest ranked juvenile reaches a critical size (*Elliott, Elliott & Mariscal, 1995; Buston, 2003b*). Because this study did not incorporate gonad analyses, and the length of certain largest individuals were potentially below the length for reproductive maturity, individuals measured are referred to by their size ranking: rank 1 (the largest individual), rank 2 (the second largest individual), and rank 3 (the third largest individual).

The research in this study was conducted in compliance with the Queensland Animal Care and Protection Act 2001, and with the consent of the Mahonia Na Dari Research and Conservation Center, and the Local Marine Management Committee and traditional owners in Kilu.



Figure 1 Maps showing (A and B) the location of Kimbe Bay and (C) the 19 inshore reefs. Full-size DOI: 10.7717/peerj.5841/fig-1

Field methods

The majority of the *A. percula* groups were located on the fringing reef and 19 adjacent reefs by systematic searches of all the reef area shallower than 20 m (Figs. 1B and 1C). The depth and GPS coordinates were recorded for all located anemones and *A. percula* groups. For each anemone, a photograph of the entire anemone, with calipers for scale, was taken to estimate the effective surface area. The number of *A. percula* in the anemone was recorded. All individuals in each social group were caught using hand nets, and the total length (TL) of each individual was measured using plastic calipers before releasing it back onto the host anemone.

Environmental variables

All anemones were mapped using QGIS software, and distance from shore was calculated as the linear distance between each anemone and the closest point on the shore. Using Digital Earth Watch software, tentacle crown surface area was calculated from the anemone photographs (*Dixon, McVay & Chadwick, 2017*). This process involved virtual drawing along the contour of the anemone, including specific tentacles, to capture the full extent of the surface area. Anemones that appeared to have contracted tentacles and oral discs when photographed were not included in statistical analyses of anemone size, as measurements of surface area would have been underestimated.

Group size and structure

Four variables were measured to quantify the social group structure of *A. percula*: (1) social group size, or the number of fish per anemone, (2) TL of rank 1 (the largest individual), (3) TL of rank 2 (the second largest individual), and (4) TL of rank 3 (the third largest individual). Only the lengths of the three largest individuals were assessed, because few *A. percula* groups contained more than three individuals per anemone in the area surveyed.

Effects of environmental variables on group size and structure

Separate linear regressions between each environmental variable (anemone size, depth, and distance from shore) and each social structure variable (group size, TL of rank 1, TL of rank 2, and TL of rank 3) were fitted. The environmental variables were log transformed to ensure a linear relationship with social structure variables and to ensure the data conformed to the assumptions of linear regression. The potential relationships between anemone size and the other environmental variables (depth and distance offshore) were also assessed.

As there were significant correlations between environmental variables, correlation coefficients were calculated and compared against thresholds of 0.7–0.8, levels recommended for conducting regressions without multicollinearity posing an issue (*Green, 1979; Menard, 2002*). Subsequently, the variance inflation factor (VIF) was calculated using the equation: $VIF_j = 1/(1-R_j^2)$. The VIF was then compared against thresholds of 4–10, below which multicollinearity would not be a concern (*Harrell, 2001; Guisan, Edwards & Hastie, 2002; Hair et al., 2006*).

Effect of rank 1 on social structure

The possible effects of the size of the rank 1 individual on group size and structure were analyzed using separate univariate regressions, with length of rank 1 as the independent variable, and the social group size, length of rank 2, and length of rank 3 as the dependent variable in each regression. The length of rank 1 was used as the social control factor, as previous studies have shown that female size (always the rank 1 individual) is a significant predictor of social group size and the size of subordinate individuals (*Buston, 2003a; Mitchell & Dill, 2005*).

Path analysis

A path analysis was conducted using structural equation modelling to determine the likely pathways that link anemone size to social group size and structure. Path analysis imposes directional causation from one variable to the next and includes the magnitude of such effects. Relationships in nature are often not solely unidirectional and, as in the case of the relationship between anemone and anemonefish, may involve feedback loops where both members yield influence on each other. Nevertheless, path analysis allows exploration of the strength of interactions between many interacting factors in a potentially complex model (*Streiner, 2005*). Therefore, use of the path analysis in this study provides a useful

platform for assessing the magnitude of both environmental and social influences on the group structure of anemonefish.

The path analysis in this study was conducted in R using the Lavaan package and devised using the results from the individual regressions as a starting point. The causation directions were informed both from previous literature on the hierarchy of *A. percula* group structure (*Buston, 2003a*) as well as a basic assumption that anemone size may be influenced by both depth and distance offshore. The fit of the model was evaluated by examining the R^2 of the variables and comparing fit indices to accepted thresholds (e.g., a Comparative Fit Index of greater than 0.95 and a standardized root mean square residual below 0.08) (*Browne & Cudeck, 1993; Hu & Bentler, 1999*). Direct effects were determined by examining the path coefficients. Indirect effects were assessed by first determining the compound path and subsequently multiplying the coefficients of the component paths. The use of path analysis in this study is an a posteriori approach, fitting a model to a known data set for explanatory analysis, as opposed to creating a model prior to examining the data available.

RESULTS

General patterns

A total of 98 *H. magnifica* with resident fish were mapped and sampled on the 19 inshore reefs. Six of these anemones were not included in statistical analyses of anemone size as they were contracted when photographed, and therefore measurements of surface area would have been underestimated. The distances from shore of the 92 anemones included in statistical analyses ranged from 185 to 1,400 m, and the depths occupied ranged from 3.3 to 19 m. The size of *A. percula* social groups ranged from two to five individuals per anemone.

Are group size and structure positively related to anemone size?

Social group size, body length of rank 1, body length of rank 2, and body length of rank 3 all increased significantly with an increase in the log of anemone surface area (Fig. 2). Anemone size explained 28.3% of the variation in group size, 72.4% of the variation in rank 1 length, 65.5% of the variation in rank 2 length, and 48.25% of the variation in rank 3 length. All linear regressions were statistically significant ($F_{1,90} = 35.7$, p < 0.001; $F_{1,90} = 236.1$, p < 0.001; $F_{1,88} = 167.7$, p < 0.001; $F_{1,72} = 67.14$, p < 0.001).

Does anemone depth or distance offshore explain variation in group size and structure, and can this be attributed to spatial variation in anemone size?

There were significant, weak positive relationships between depth of the anemone and the body lengths of rank 1 rank 2, and rank 3 ($F_{1,90} = 17.3$, p < 0.001; $F_{1,88} = 6.591$, p = 0.012; $F_{1,72} = 5.054$, p = 0.028; Fig. 3). These relationships explained 16.1% of the variation in rank 1 body length, 7.0% of the variation in rank 2 body length, and 6.6% of the variation in rank 3 body length, respectively. There was no significant





relationship between social group size and depth of the anemone (although there was a positive trend).

Distance from shore had no significant effect on group size, the body length of rank 2, or the body length of rank 3 (Fig. 4). However, there was a weak positive relationship between body length of rank 1 and the log of the distance from shore, which explained 8.2% of the variation in the length of rank 1 ($F_{1,90} = 7.991$, p = 0.006; Fig. 4).

There were significant positive relationships between the log of the anemone surface area and the log of the distance from shore, and the log of the depth ($F_{1,90} = 7.515$, p = 0.007; $F_{1,90} = 20.35$, p < 0.001 values; Fig. 5). Depth and distance from shore were weakly correlated with each other (r = 0.22, $t_{2,90} = 2.155$, p = 0.034). A multiple regression with depth and distance from shore as independent variables explained 21.9% of the variation in anemone surface area ($F_{2,89} = 12.51$, p < 0.001). The VIF factor was 1.28, indicating that the collinearity among the environmental factors did not limit the ability to distinguish among variables.

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Figure 3 Relationships between log depth and (A) social group size, (B) rank 1 length, (C) rank 2 length, (D) rank 3 length. Lines indicate significant relationships. Full-size 🖾 DOI: 10.7717/peerj.5841/fig-3

Are the group size, and the size of the second and third largest individuals related to the size of the largest individual?

Social group size, body length of rank 2, and body length of rank 3 each increased significantly with an increase in body length of rank 1 ($F_{1,90} = 36.74$, p < 0.001; $F_{1,88} = 315.4$, p < 0.001; $F_{1,72} = 152.2$, p < 0.001; Fig. 6). Rank 1 body length explained 29.0% of the variation in group size, 78.2% of the variation in rank 2 length, and 67.9% of the variation in the length of rank 3.

Does anemone size directly determine group size or is this mediated by the size of the largest individual?

The model produced by path analysis explained between 58% and 78% of the variation in the continuous group structure variables. The model fit the data well, with a Comparative Fit Index of 0.992, a Tucker-Lewis Index of 0.986, a root mean square error of approximation of 0.052, and a standardized root mean square residual of 0.051 (Fig. 7). Depth and distance from shore maintained weak, but direct, effects on anemone surface







Figure 5 Relationships between log anemone surface area and (A) log distance from shore, and (B) log depth. Lines indicate significant relationships. Full-size DOI: 10.7717/peerj.5841/fig-5







Figure 7 Path analysis showing the influence of environmental and social variables on social group size and structure. Direct effects are indicated by solid lines, with line thicknesses indicating the strength of the effect. Path coefficients, standardized versions of linear regression weights, are indicated below each solid line. Covariances are indicated by dotted lines.

Full-size 🖾 DOI: 10.7717/peerj.5841/fig-7

area. Anemone surface area had a strong direct effect on rank 1 body length, and rank 1 body length had strong, direct effects on the rank 2 body length and rank 3 body length. Through its effects on subordinate lengths, rank 1 maintained an indirect, yet important effect on social group size. Within this model, the direct effect of the anemone surface area on rank 1 body length was the strongest, direct interaction. However, anemone surface area also had positive, indirect effects on social group size and structure, via its effect on rank 1 body length (Fig. 7).

DISCUSSION

Overall, the results indicate that both environmental and social factors play important roles in determining group size and structure in *A. percula* associated with *H. magnifica* anemones in Papua New Guinea. Anemone surface area was clearly the most influential environmental variable, with larger social groups and larger individuals inhabiting larger anemones. Distance from shore and depth also had positive, indirect effects on group structure, most likely through their positive effects on anemone surface area, whereby anemones were larger at greater depths and distances from shore. Social factors also played an important role in regulating social group structure, with rank 1 positively affecting social group size, the body length of rank 2, and the body length of rank 3. Examination of these combined effects through path analysis demonstrated that anemone surface area directly affected the structure of rank 1 and rank 2 body lengths, and influenced rank 3 length and group size through the direct effects on the two highest ranking individuals.

Critical importance of anemone size

Our results confirm those of previous studies showing that anemone size has a major influence on the size and structure of social groups of anemonefishes (*Fricke, 1980*; *Fautin, 1992; Mitchell & Dill, 2005; Dixon, McVay & Chadwick, 2017*). Anemone size was a good predictor of group size, and the lengths of rank 1, rank 2, and rank 3 individuals

present on the anemone. The strong relationship between anemone size and rank 1 length is consistent with previous work (*Elliott & Mariscal, 2001; Mitchell & Dill, 2005*). *Buston (2003a)* did not find this relationship for *A. percula* in Madang, Papua New Guinea, suggesting some spatial variation in this phenomenon. It is important to note that the strength of this interaction is likely to be reinforced by reciprocal benefits that anemonefish bestow on their hosts, including defense (*Porat & Chadwick-Furman, 2004; Godwin & Fautin, 1992*) and provision of nutrients (*Porat & Chadwick-Furman, 2005; Cleveland, Verde & Lee, 2010; Roopin & Chadwick, 2009; Roopin et al., 2011*).

Role of depth and distance offshore

Although the regression analysis suggested depth did not have a direct effect on social group size, deeper anemones certainly supported larger rank 1 and rank 2 individuals. Because the highest rank individuals tend to be the breeding pair, and fecundity increases with body size (Saenz-Agudelo et al., 2015), this relationship may have important consequences for reproductive potential. The depth effect is most likely related to the reduction in light with increasing depth. It is known that anemones have diel contraction patterns in response to the availability of light, with tentacles that have high densities of zooxanthellae responding to levels of light intensity (Sebens & DeRiemer, 1977). Anemones in shallow areas may retain retracted tentacles due to the presence of potentially high levels of photosynthetically active radiation, which can adversely affect the anemone and its Symbiodinium by inhibiting photosynthesis (Mass et al., 2010). Anemones at greater depths may conversely expand their oral disc to capture more light for photosynthesis, as light intensity is reduced at greater depths. However, anemones have also been shown to increase the concentration of their Symbiodinium at depth as a way to increase their photosynthetic capability (Dixon et al., 2014), and thus an alternative explanation is that anemones might actually be larger at greater depths, due to reduced wave action, increased nutrient availability, or other environmental parameters (Sebens, 1982). Either way, the outcome is a greater surface area available for A. percula on deeper anemones.

Distance from shore also had a weak positive effect on rank 1 length, again, most likely as a result of the effect of distance offshore on anemone surface area. Anemones that were further offshore tended to be slightly larger. The presence of smaller anemones closer to shore may be due to the likely higher levels of sedimentation near the fringing reefs. Increased sediment has previously been linked to a reduction in photosynthetic ability and an increase in tissue bleaching in anemones (*Goreau, 1964; Rogers, 1990*). Anemones produce mucus to remove sediment (*Patton, 1979*), which may be energetically costly and result in reduced growth in waters with higher sediment loads. Higher levels of sediment. Conditions further offshore may also foster more favorable environments in terms of nutrient availability, current patterns, and, more broadly, a reduction in anthropogenic stressors (*Smith et al., 2008*). One or both of these mechanisms would lead to a decrease in anemone surface area with proximity to the shore and a smaller area for *A. percula* to inhabit.

Role of the largest individual

The size of the rank 1 individual (in most cases-the female) exhibited strong relationships with rank 2 body size, rank 3 body size, and the group size. This is likely the outcome of the strong social hierarchy in anemonefish groups, and our results are consistent with other work on anemonefishes (Fricke & Fricke, 1977; Fricke, 1979, 1983; Godwin, 1994; Buston, 2003a; Iwata et al., 2008). Mitchell & Dill (2005) concluded that social group size in the sister species, A. ocellaris, is predominantly controlled by the dominant female (rank 1), based on the effect of female size on the size structure of the group. Buston (2003a) has previously established the importance of social control on individual body lengths in A. percula, with a fixed size difference between fishes that are sequentially ranked in the social hierarchy. With such fixed size ratios, the larger the rank 1 individual, the more individuals can be accommodated in the group. The mechanism controlling group size in anemonefish is likely forcible eviction or competitive exclusion (Buston, 2003b), leading to excluded fish finding refuge in potentially less preferable hosts (Huebner et al., 2012). Lower social rank has been associated with lower survivorship in A. percula, likely due to competitive eviction by higher ranked individuals (Buston, 2003c). Subordinate individuals may in turn be reducing their food intake to avoid exceeding size thresholds that could lead to eviction, as observed in other space-limited fishes (*Wong* et al., 2008). Hence, the size of the group and its structure may be determined by complex top-down and bottom-up interactions between competing individuals in the social hierarchy, including the species of host anemone.

Distinguishing direct and indirect effects

It is very difficult to tease apart the relative importance of environmental factors (e.g., anemone size) and social factors (e.g., size of individuals by rank) in determining social structure variables, such as group size or the size of the subordinate fishes, since all the variables are correlated. Anemone size may directly influence group size through strict space limitation and associated density-dependent processes, or indirectly via the effect of anemone size on rank 1 size (Mitchell & Dill, 2005). Buston (2003a) concluded that the dominant female's body length (rank 1 length) controlled the body lengths of the subordinates, because anemone diameter did not have a significant effect. Mitchell & Dill (2005) reached similar conclusions on the basis of stronger relationships with female size, compared with anemone size. Our path analysis strongly suggested that both direct and indirect effects are involved. Anemone surface area had a strong direct effect on rank 1 size and a slightly weaker effect on rank 2 size. The size of rank 1, in turn had a strong direct effect on the sizes of rank 2 and rank 3, while maintaining a substantial indirect effect on social group size through its effects on subordinates. Hence, anemone area had considerable indirect effects on all of the variables governing group size and structure. Further, the total effect (direct + indirect) of anemone size on rank 2 size was comparable to the direct effect of rank 1. The path analysis also demonstrated that the environmental variables (distance from shore, depth) have indirect effects on all four group structure variables through their influence on anemone surface area, although the effect of depth is arguably greater than that of distance from the coastline.

Future directions

While the path analysis helps resolve the likely cause-effect links and direct vs indirect relationships, ultimately this complexity will need to be resolved by experimentation. Experiments have previously demonstrated that anemonefish provide important benefits to their hosts through a multitude of mechanisms. While *Godwin & Fautin (1992)* demonstrated the importance of *A. percula* for protection of anemones, other potential benefits, such as nutrient transfer and aeration, have yet to be tested for this species and need be resolved. Observation of the behavior of different individuals in such experimental designs may also aid in revealing the exact mechanisms responsible for control of group size and structure.

Further investigation is needed to understand the reasons underlying the effects of depth and distance offshore on social group structure. Variations in depth are coupled with variations in light intensity, temperature, and pressure (*Levinton*, 1982). Similarly, distance from the coastline often varies with other factors such as nutrient concentration, sediment runoff, and anthropogenic impacts (*Fabricius*, 2005; *Wenger et al.*, 2015). It would be tractable to experimentally manipulate other environmental variables, such as light intensity and temperature, to fully understand how each variable influences the host anemones, and consequently, the structure of *A. percula* social groups.

There is increasing evidence that shallow water anemones experience severe bleaching after warm water events (*Saenz-Agudelo et al., 2011*; *Hobbs et al., 2013*). Bleaching not only reduced size and abundance of anemones (*Hobbs et al., 2013*), but further resulted in lower egg production and recruitment of anemone fish (*Saenz-Agudelo et al., 2011*). Higher temperatures have also been linked to breeding disruption, egg degradation, and possible connectivity among anemonefish populations (*Munday et al., 2008, 2009*). These effects are likely to have flow-on effects on the structure of clownfish social groups. Further attention should be directed to evaluating ongoing effects of climate change on the ecology and behavior of fishes that are so closely linked to the quality of their habitat.

CONCLUSIONS

This study suggests that the surface area of the host anemone and the length of the largest individual are two critical factors defining the social group size and structure of *A. percula.* Variation in group structure along spatial gradients such as depth or distance from the coast may also be explained by the effects of these gradients on host anemone size. These results highlight the critical roles of habitat patch size and social hierarchy in governing breeding population size and access to limited resources in these highly specialized fishes. Additional experimental work is needed to fully resolve the direct and indirect effects of anemone size and the size of the largest individual on group size and structure, and how the behavior of this species will be shaped by increasing stresses on the anemone habitat.

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The authors declare that they have no competing interests.

Author Contributions

- Juliette Chausson conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Maya Srinivasan conceived and designed the experiments, performed the experiments, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
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Data Availability

The following information was supplied regarding data availability: Raw data is provided in the Supplemental Files.

Supplemental Information

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REFERENCES

Alcock J. 2001. Animal Behavior: An Evolutionary Approach. Sunderland: Sinauer, Associates.

- Almany GR, Berumen ML, Thorrold SR, Planes S, Jones GP. 2007. Local replenishment of coral reef fish populations in a marine reserve. *Science* 316(5825):742–744 DOI 10.1126/science.1140597.
- Browne MW, Cudeck R. 1993. Alternative ways of assessing model fit. In: Bollen K, Long J, eds. *Testing Structural Equation Models*. Newbury Park: Sage, 136–162.
- Buston P. 2003a. Social hierarchies, size and growth modification in clownfish. *Nature* 424(6945):145–146 DOI 10.1038/424145a.
- Buston P. 2003b. Forcible eviction and prevention of recruitment in the clown anemonefish. *Behavioral Ecology* 14(4):576–582 DOI 10.1093/beheco/arg036.
- **Buston PM. 2003c.** Mortality is associated with social rank in the clown anemonefish (*Amphiprion percula*). *Marine Biology* **143(4)**:811–815 DOI 10.1007/s00227-003-1106-8.
- **Buston PM, Elith J. 2011.** Determinants of reproductive success in dominant pairs of clownfish: a boosted regression tree analysis. *Journal of Animal Ecology* **80(3)**:528–538 DOI 10.1111/j.1365-2656.2011.01803.x.
- Caraco T, Martindale S, Whitlam TS. 1980. An empirical demonstration of risk-sensitive foraging preferences. *Animal Behaviour* 28(3):820–830 DOI 10.1016/S0003-3472(80)80142-4.
- Caraco T, Wolf LL. 1975. Ecological determinants of group sizes of foraging lions. American Naturalist 109(967):343–352 DOI 10.1086/283001.
- **Chapman CA, Chapman LJ. 2000.** Determinants of group sizes in primates: the importance of travel costs. In: Boinski S, Garber PA, eds. *On the Move: How and Why Animals Travel in Groups*. Chicago: University of Chicago Press, 24–41.
- Choat JH, Bellwood DR. 1985. Interactions amongst herbivorous fishes on a coral reef: influence of spatial variation. *Marine Biology* 89(3):221–234 DOI 10.1007/BF00393655.
- Cleveland A, Verde EA, Lee RW. 2010. Nutritional exchange in a tropical tripartite symbiosis: direct evidence for the transfer of nutrients from anemonefish to host anemone and zooxanthellae. *Marine Biology* **158**(3):589–602 DOI 10.1007/s00227-010-1583-5.
- Cody ML. 1971. Finch flocks in the Mohave desert. *Theoretical Population Biology* 2(2):141–158 DOI 10.1016/0040-5809(71)90012-8.
- **Crook JH. 1970.** The socio-ecology of primates. In: Crook JH, ed. *Social Behaviour in Birds and Mammals: Essays on the Social Ethology of Animals and Man.* New York: Academic Press, 103–166.
- D'Aloia CC, Majoris JE, Buston PM. 2011. Predictors of the distribution and abundance of a tube sponge and its resident goby. *Coral Reefs* 30(3):777–786 DOI 10.1007/s00338-011-0755-1.
- **Dixon AK, McVay MJ, Chadwick NE. 2017.** Demographic modelling of giant sea anemones: population stability and effects of mutualistic anemonefish in the Jordanian Red Sea. *Marine and Freshwater Research* **68(11)**:2145–2155 DOI 10.1071/MF16361.
- Dixon A, Needham D, Al-Horani AF, Chadwick NE. 2014. Microhabitat use and photoacclimation in the clownfish sea anemone *Entacmaea quadricolor*. *Journal of the Marine Biological Association of the United Kingdom* **94(3)**:473–480 DOI 10.1017/S0025315413001719.

- Elliott JK, Elliott JM, Mariscal RN. 1995. Host selection, location, and association behaviors of anemonefishes in field settlement experiments. *Marine Biology* 122(3):377–389 DOI 10.1007/BF00350870.
- Elliott JK, Mariscal RN. 2001. Coexistence of nine anemonefish species: differential host and habitat utilization, size and recruitment. *Marine Biology* **138**(1):23–36 DOI 10.1007/S002270000441.
- **Fabricius KE. 2005.** Effects of terrestrial runoff on the ecology of corals and coral reefs: review and synthesis. *Marine Pollution Bulletin* **50(2)**:125–146 DOI 10.1016/j.marpolbul.2004.11.028.
- Fautin DG. 1992. Anemonefish recruitment: the roles of order and chance. Symbiosis 14:143–160.
- **Fautin DG, Allen GR. 1992.** *Field Guide to Anemonefishes and their Host Sea Anemones.* Perth: Western Australian Museum.
- Fitzpatrick JL, Desjardins JK, Milligan N, Stiver KA, Montgomerie R, Balshine S. 2008. Female-mediated causes and consequences of status change in a social fish. *Proceedings of the Royal Society B: Biological Sciences* 275(1637):929–936 DOI 10.1098/rspb.2007.1449.
- Fricke HW. 1979. Mating system, resource defence and sex change in the anemonefish Amphiprion akallopisos. Zeitschrift für Tierpsychologie 50(3):313–326 DOI 10.1111/j.1439-0310.1979.tb01034.x.
- Fricke HW. 1980. Control of different mating systems in a coral reef fish by one environmental factor. *Animal Behavior* 28(2):561–569 DOI 10.1016/S0003-3472(80)80065-0.
- Fricke HW. 1983. Social control of sex: field experiments with the anemonefish *Amphiprion* bicinctus. Zeitschrift für Tierpsychologie 61(1):71–77 DOI 10.1111/j.1439-0310.1983.tb01327.x.
- Fricke HW, Fricke S. 1977. Monogamy and sex change by aggressive dominance in coral reef fish. *Nature* 266(5605):830–832 DOI 10.1038/266830a0.
- Frisch AJ, Rizzari JR, Munkres KP, Hobbs JA. 2016. Anemonefish depletion reduces survival, growth, reproduction and fishery productivity of mutualistic anemone–anemonefish colonies. *Coral Reefs* 35(2):375–386 DOI 10.1007/S00338-016-1401-8.
- Gilmour KM, DiBattista JD, Thomas JB. 2005. Physiological causes and consequences of social status in salmonid fish. *Integrative and Comparative Biology* **45(2)**:263–273 DOI 10.1093/icb/45.2.263.
- **Godwin J. 1994.** Behavioural aspects of protandrous sex change in the anemonefish, *Amphiprion melanopus*, and endocrine correlates. *Animal Behaviour* **48(3)**:551–567 DOI 10.1006/anbe.1994.1275.
- Godwin J, Fautin DG. 1992. Defense of host actinians by anemonefishes. *Copeia* 1992(3):902–908 DOI 10.2307/1446171.
- Goreau TF. 1964. Mass expulsion of zooxanthellae from Jamaican reef communities after Hurricane Flora. *Science* 145(3630):383–386 DOI 10.1126/science.145.3630.383.
- **Green RH. 1979.** Sampling Design and Statistical Methods for Environmental Biologists. New York: Wiley.
- **Guisan A, Edwards TC, Hastie T. 2002.** Generalized linear and generalized additive models in studies of species distributions: setting the scene. *Ecological Modelling* **157(2–3)**:89–100 DOI 10.1016/S0304-3800(02)00204-1.
- **Gust N, Choat JH, McCormick MI. 2001.** Spatial variability in reef fish distribution, abundance, size and biomass: a multi-scale analysis. *Marine Ecology Progress Series* **214**:237–251 DOI 10.3354/meps214237.
- Hair JF, Black WC, Babin BJ, Anderson RE, Tatham RL. 2006. *Multivariate Data Analysis*. Vol. 6. Upper Saddle River: Prentice Hall.

- Hamilton WD. 1971. Geometry for the selfish herd. *Journal of Theoretical Biology* 31(2):295–331 DOI 10.1016/0022-5193(71)90189-5.
- Harrell FE. 2001. Regression Modeling Strategies: With Applications to Linear Models, Logistic Regression and Survival Analysis. New York: Springer-Verlag.
- Hobbs JPA, Frisch AJ, Ford BM, Thums M, Saenz-Agudelo P, Furby KA,
 Berumen ML. 2013. Taxonomic, spatial and temporal patterns of bleaching in anemones inhabited by anemonefishes. *PLOS ONE* 8(8):e70966
 DOI 10.1371/journal.pone.0070966.
- Hobbs JPA, Munday PL, Jones GP. 2004. Social induction of maturation and sex determination in a coral reef fish. *Proceedings of the Royal Society B: Biological Sciences* 271(1553):2109–2114 DOI 10.1098/rspb.2004.2845.
- Holbrook SJ, Schmitt RJ. 2005. Growth, reproduction and survival of a tropical sea anemone (Actiniaria): benefits of hosting anemonefish. *Coral Reefs* 24(1):67–73 DOI 10.1007/s00338-004-0432-8.
- Hoogenboom MO, Connolly SR, Anthony KRN. 2008. Interactions between morphological and physiological plasticity optimize energy acquisition in corals. *Ecology* **89(4)**:1144–1154 DOI 10.1890/07-1272.1.
- Hu LT, Bentler PM. 1999. Cutoff criteria for fit indexes in covariance structure analysis: conventional criteria versus new alternatives. *Structural Equation Modeling: A Multidisciplinary Journal* 6(1):1–55 DOI 10.1080/10705519909540118.
- Huebner LK, Dailey B, Titus BM, Khalaf M, Chadwick NE. 2012. Host preference and habitat segregation among Red Sea anemonefish: effects of sea anemone traits and fish life stages. *Marine Ecology Progress Series* 464:1–15 DOI 10.3354/meps09964.
- Iwata E, Nagai Y, Hyoudou M, Sasaki H. 2008. Social environment and sex differentiation in the false clown anemonefish, *Amphiprion ocellaris*. Zoological Science 25(2):123–128 DOI 10.2108/zsj.25.123.
- Jankowski MW, Gardiner NR, Jones GP. 2015. Depth and reef profile: effects on the distribution and abundance of coral reef fishes. *Environmental Biology of Fishes* **98(5)**:1373–1386 DOI 10.1007/s10641-014-0365-1.
- Jankowski MW, Graham NAJ, Jones GP. 2015. Depth gradients in diversity, distribution and habitat specialisation in coral reef fishes: implications for the depth-refuge hypothesis. *Marine Ecology Progress Series* 540:203–215 DOI 10.3354/meps11523.
- Krebs CJ. 1972. *Ecology: The Experimental Analysis of Distribution and Abundance*. New York: Harper Collins.
- Krebs JR, Davies NB. 1981. An Introduction to Behavioural Ecology. Oxford: Blackwell.
- Levinton JS. 1982. Marine Ecology. Englewood Cliffs: Prentice Hall.
- MacDonald C, Bridge TCL, Jones GP. 2016. Depth, bay position and habitat structure as determinants of coral reef fish distributions: are deep reefs a potential refuge? *Marine Ecology Progress Series* 561:217–231 DOI 10.3354/meps11953.
- Markham AC, Gesquiere LR, Alberts SC, Altmann G. 2015. Optimal group size in a highly social mammal. Proceedings of the National Academy of Sciences of the United States of America 112(48):14882–14887 DOI 10.1073/pnas.1517794112.
- Mass T, Kline DI, Roopin M, Veal CJ, Cohen S, Iluz D, Levy O. 2010. The spectral quality of light is a key driver of photosynthesis and photoadaptation in *Stylophora pistillata* colonies from different depths in the Red Sea. *Journal of Experimental Biology* **213**(23):4084–4091 DOI 10.1242/jeb.039891.

- McGehee MA. 1994. Correspondence between assemblages of coral reef fishes and gradients of water motion, depth, and substrate size off Puerto Rico. *Marine Ecology Progress Series* 105:243–255 DOI 10.3354/meps105243.
- Menard SW. 2002. Applied Logistic Regression Analysis. Thousand Oaks: Sage Publications.
- Mitchell JS, Dill LM. 2005. Why is group size correlated with the size of the host sea anemone in the false clown anemonefish? *Canadian Journal of Zoology* 83(2):372–376 DOI 10.1139/z05-014.
- Munday PL, Jones GP, Caley MJ. 1997. Habitat specialisation and the distribution and abundance of coral-dwelling gobies. *Marine Ecology Progress Series* 152:227–239 DOI 10.3354/meps152227.
- Munday PL, Jones GP, Pratchett MS, Williams AJ. 2008. Climate change and the future for coral reef fishes. *Fish and Fisheries* 9(3):261–285 DOI 10.1111/j.1467-2979.2008.00281.x.
- Munday PL, Leis JM, Lough JM, Paris CB, Kingsford MJ, Berumen ML, Lambrechts J. 2009. Climate change and coral reef connectivity. *Coral Reefs* 28(2):379–395 DOI 10.1007/s00338-008-0461-9.
- Noonan SHC, Jones GP, Pratchett MS. 2012. Coral size, health and structural complexity: effects on the ecology of a coral reef damselfish. *Marine Ecology Progress Series* **456**:127–137 DOI 10.3354/meps09687.
- Patton WK. 1979. On the association of the spider crab, *Mithrax* (Mithraculus) *cinctimanus* (Stimpson) with Jamaican sea anemones. *Crustaceana* Suppl. 5(Studies on Decapoda):55–61.
- Pittman SJ, Brown KA. 2011. Multi-scale approach for predicting fish species distributions across coral reef seascapes. *PLOS ONE* 6(5):e20583 DOI 10.1371/journal.pone.0020583.
- Planes S, Jones GP, Thorrold SR. 2009. Larval dispersal connects fish populations in a network of marine protected areas. *Proceedings of the National Academy of Sciences of the United States of America* 106(14):5693–5697 DOI 10.1073/pnas.0808007106.
- Porat D, Chadwick-Furman NE. 2004. Effects of anemonefish on giant sea anemones: expansion behavior, growth, and survival. *Hydrobiologia* 530–531(1–3):513–520 DOI 10.1007/s10750-004-2688-y.
- **Porat D, Chadwick-Furman NE. 2005.** Effects of anemonefish on giant sea anemones: ammonium uptake, zooxanthella content and tissue regeneration. *Marine and Freshwater Behaviour and Physiology* **38(1)**:43–51 DOI 10.1080/10236240500057929.
- Ricklefs RE, Miller GL. 1999. Ecology. San Francisco: Freeman.
- **Rogers CS. 1990.** Responses of coral reefs and reef organisms to sedimentation. *Marine Ecology Progress Series* 62:185–202 DOI 10.3354/meps062185.
- Roopin M, Chadwick NE. 2009. Benefits to host sea anemones from ammonia contributions of resident anemonefish. *Journal of Experimental Marine Biology and Ecology* **370(1–2)**:27–34 DOI 10.1016/j.jembe.2008.11.006.
- **Roopin M, Thornhill DJ, Santos SR, Chadwick NE. 2011.** Ammonia flux, physiological parameters, and Symbiodinium diversity in the anemonefish symbiosis on Red Sea coral reefs. *Symbiosis* **53(2)**:63–74 DOI 10.1007/s13199-011-0110-x.
- Saenz-Agudelo P, Jones GP, Thorrold SR, Planes S. 2011. Detrimental effects of host anemone bleaching on anemonefish populations. *Coral Reefs* 30(2):497–506 DOI 10.1007/s00338-010-0716-0.
- Saenz-Agudelo P, Jones GP, Thorrold SR, Planes S. 2015. Mothers matter: contribution to local replenishment is linked to female size, mate replacement and fecundity in a fish metapopulation. *Marine Biology* 162(1):3–14 DOI 10.1007/s00227-014-2556-x.

- Sebens KP. 1982. The limits to indeterminate growth: an optimal size model applied to passive suspension feeders. *Ecology* 63(1):209–222 DOI 10.2307/1937045.
- Sebens K, DeRiemer K. 1977. Diel cycles of expansion and contraction in coral reef anthozoans. *Marine Biology* 43(3):247–256 DOI 10.1007/BF00402317.
- Shapiro DY. 1981. Size, maturation and the social control of sex reversal in the coral reef fish Anthais squamipinnis. Journal of Zoology 193(1):105–128 DOI 10.1111/j.1469-7998.1981.tb01494.x.
- Smallhorn-West PF, Bridge TCL, Munday PL, Jones GP. 2017. Depth distribution and abundance of a coral-associated reef fish: roles of recruitment and post-recruitment processes. *Coral Reefs* 36(1):157–166 DOI 10.1007/s00338-016-1509-x.
- Smith TV, Chapman CA. 2007. Primate group size and interpreting socioecological models: do folivores really play by different rules? *Evolutionary Anthropology: Issues, News, and Reviews* 16(3):94–106 DOI 10.1002/evan.20132.
- Smith TB, Nemeth RS, Blondeau J, Calnan JM, Kadison E, Herzlieb S. 2008. Assessing coral reef health across onshore to offshore stress gradients in the US Virgin Islands. *Marine Pollution Bulletin* 56(12):1983–1991 DOI 10.1016/j.marpolbul.2008.08.015.
- Srinivasan M. 2003. Depth distributions of coral reef fishes: the influence of microhabitat structure, settlement, and post-settlement processes. *Oecologia* 137(1):76–84 DOI 10.1007/s00442-003-1320-6.
- Streiner DL. 2005. Finding our way: an introduction to path analysis. *Canadian Journal of Psychiatry* 50(2):115–122 DOI 10.1177/070674370505000207.
- Szczebak JT, Henry RP, Al-Horani FA, Chadwick NE. 2013. Anemonefish oxygenate their anemone hosts at night. *Journal of Experimental Biology* 216(6):970–976 DOI 10.1242/jeb.075648.
- Thompson VJ, Munday PL, Jones GP. 2007. Habitat patch size and mating system as determinants of social group size in coral-dwelling fishes. *Coral Reefs* 26(1):165–174 DOI 10.1007/s00338-006-0181-y.
- Wenger AS, Fabricius KE, Jones GP, Brodie JE. 2015. Sedimentation, eutrophication and pollution: effects on coral reef fishes. In: Mora C, ed. *Ecology of Fishes on Coral Reefs: The Functioning of an Ecosystem in a Changing World*. Cambridge: Cambridge University Press, 145–153.
- Williams DMcB. 1991. Patterns and processes in the distribution and abundance of coral reef fishes. In: Sale PF, ed. *The Ecology of Fishes on Coral Reefs*. San Diego: Academic Press, 437–474.
- Wilson EO. 1975. Sociobiology: The New Synthesis. Cambridge: Harvard University Press.
- Wong MYL, Buston PM, Munday PL, Jones GP. 2007. The threat of punishment enforces peaceful cooperation and stabilizes queues in a coral-reef fish. *Proceedings of the Royal Society B: Biological Sciences* 274(1613):1093–1099 DOI 10.1098/rspb.2006.0284.
- Wong MYL, Munday PL, Buston P, Jones GP. 2008. Fasting or feasting in a fish social hierarchy. *Current Biology* 18(9):R372–R373 DOI 10.1016/j.cub.2008.02.063.
- Wong MYL, Munday PL, Jones GP. 2005. Habitat patch size, facultative monogamy and sex change in a coral-dwelling fish, *Caracanthus unipinna*. *Environmental Biology of Fishes* 74(2):141–150 DOI 10.1007/s10641-005-6715-2.