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**Title: Which species distribution models are more (or less) likely to project broad-scale, climate-induced shifts in species ranges?**

**Running title: Biases in species distribution models**

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## **Abstract**

Species distribution models (SDMs) frequently project substantial declines in the spatial extent of climatically suitable habitat in response to scenarios of future climate change. Such projections are highly disconcerting. Yet, considerable variation can occur in the direction and magnitude of range changes projected by different SDM methods, even when predictive performance is similar. In this study, we assessed whether particular methods have a tendency to predict substantial loss or gain of suitable habitat. In particular, we asked, “are 14 SDM methods equally likely to predict extreme changes to the future extent of suitable habitat for 220 Australian mammal species?”. We defined five non-mutually exclusive categories of ‘extreme’ change, based on stability or loss of current habitat, or the dislocation of current and future habitat; a) no future habitat (range extinction), b) low stability of current habitat ( $\leq 10\%$  remains), c) no gain of habitat in new locations, d) all future habitat is in new locations (i.e. completely displaced from current habitat) and e) substantial increase in size of habitat (future habitat is  $\geq 100\%$  larger than current). We found that some SDM methods were significantly more likely than others to predict extreme changes. In particular, distance-based models were significantly less likely than other methods to predict substantial increases in habitat size; Random Forest models and Surface Range Envelopes were significantly more likely to predict a complete loss of current habitat, and future range extinction. Generalised Additive Models and Generalised Linear Models rarely predicted range extinction; future

habitat completely disjunct from current habitat was predicted more frequently than expected by Classification Tree Analysis and less frequently by Maxent. Random Forest generally predicted extreme range changes more frequently than other SDM methods. Our results identify trends among different methods with respect to tendency to predict extreme range changes. These are of significance for climate-impact assessments, with implications for transferability of models to novel environments. Our findings emphasise the need to explore and justify the use of different models and their parameterisations, and to develop approaches to assist with optimisation of models.

## **Highlights**

- Species distribution models frequently predict substantial range shifts due to climate change
- Frequency of predictions of extreme range changes varies across SDM methods
- Random Forest was more likely than other methods to predict extreme range changes
- Distance-based models never predicted substantial increases in range size

## **Introduction**

Contemporary anthropogenic climate change has already resulted in shifts in the range margins of numerous, diverse taxa (Melles *et al.*, 2010; Chen *et al.*, 2011; Garroway *et al.*, 2011; Rubidge *et al.*, 2011), and paleoecological data suggest that range shifts were the norm during previous episodes of climate change (Birks, 1989; Huntley, 1990; Graham, 1992; Willis & MacDonald, 2011). Understanding the extent to which species distributions may shift in response to climate change over the course of this century and beyond may assist in identifying species vulnerable to climate change, prioritising conservation efforts, and developing optimal adaptation and land management plans.

Correlative species distribution models (SDMs) are frequently used to examine the potential for changes to the distribution and quality of habitat under scenarios of future climate (Franklin, 2010). This approach is based on the assumption that the location of populations reflects the environmental preferences and tolerances of a species (Guisan & Thuiller, 2005). Models of this relationship, based on the observed subset of those populations, can then be used to assess the suitability of a region, for a particular species, under current, past or future climate. Such climate change experiments assume that relationships inferred from historical data remain consistent under a changing climate (Maraun, 2015).

Predictions of broad-scale changes in species' distributions in response to climate change have contributed to substantial concern regarding the fate of

biodiversity in a warming world (e.g. Thomas *et al.*, 2004; Ben Ras Lasram *et al.*, 2010; Sauer *et al.*, 2011; Ihlow *et al.*, 2012; Ochoa-Ochoa *et al.*, 2012; Warren *et al.*, 2013). Numerous studies indicate that, under climate change, a sizeable proportion of species will be faced with an absence of suitable habitat (range extinction), or future habitat that is completely disjunct from current habitat. For example, ~24% of Mexican amphibian species are at risk of losing all habitat by 2080 (Ochoa-Ochoa *et al.*, 2012), while habitat for nearly 12% of the world's Chelonian species may lie in areas currently unsuitable (Ihlow *et al.*, 2012). Depending upon the scenario of greenhouse gas emissions, 11–19% of 1,541 seed plants in Alberta, Canada, were predicted to lose > 90% of current habitat by 2080 (Zhang *et al.*, 2015). Similarly, an analysis of 2,954 species across North and South America predicted the loss of all existing habitat for at least 10% of species by 2071–2100 (Lawler *et al.*, 2009). In one of the largest studies to date, Warren *et al.* (2013) estimated that after accounting for dispersal, 2 –6% of 5,382 animal species would lose  $\geq$  90% of current habitat. Hence, predictions by SDMs indicate that the rate of extinction in the 21<sup>st</sup> century may exceed estimates of historical extinction events from the fossil record. However, there is on-going debate as to the accuracy of SDMs and whether they will over- or under-estimate range changes and extinction (see review by Bellard *et al.*, 2012).

In recent years the number of methods for fitting SDMs has increased considerably (Elith & Graham, 2009; Elith & Leathwick, 2009; Franklin, 2010), and

now includes distance-based or profile models (e.g. BIOCLIM, Nix, 1986; Booth *et al.*, 2014), statistical models that can fit complex non-linear relationships (such as Generalised Additive Models) and machine-learning techniques (Elith *et al.*, 2008; Elith & Leathwick, 2009). These methods differ in complexity, data requirements and ease of use, and their characteristics have been well-described elsewhere (e.g. Syphard & Franklin, 2009; Franklin, 2010; Xinhai & Wang, 2013).

SDM method choice is influenced by access to software, data availability (e.g. presence only or presence/absence records), user ability and the particular goals of the study (Segurado & Araújo, 2004; Elith & Leathwick, 2009; Ahmed *et al.*, 2015), although different disciplines and geographic regions have traditionally utilized different techniques (Elith & Leathwick, 2009). However, identification of the most appropriate SDM method is complicated by a number of factors, and multi-model comparisons have repeatedly concluded that there is no single 'best' method (Elith *et al.*, 2006; Diniz Filho *et al.*, 2010).

Several studies have suggested that SDM methods with high flexibility in modelling complex species-environment relationships may outperform simpler methods (Elith *et al.*, 2006; Tsoar *et al.*, 2007; Li & Wang, 2013). For instance, machine-learning algorithms (such as Random Forests, Maxent and Boosted Regression Trees) frequently outperform regression-based approaches (e.g. Multivariate Adaptive Regression Splines and Generalised Linear Models) (Bucklin *et al.*, 2015, but see Guillera-Aroita *et al.*, 2015). Others caution the use of complex methods: while these may predict observed occurrence patterns well, they may



result in overfitting, and hence may not necessarily predict an acceptable representation of the species' potential distribution (Diniz-Filho *et al.*, 2010; Li & Wang, 2013). Models with high predictive accuracy for the data used in their calibration may also demonstrate poor transferability (or generality); that is, their performance may decline when projected onto different geographic regions (Heikkinen *et al.*, 2012) or time periods (Fronzek *et al.*, 2011). The Random Forest model, for example, generally has high predictive capacity as determined by AUC (the area under the receiver-operating characteristic curve) calculated on data similar to those used for model-fitting (Coetzee *et al.*, 2009; Virkkala *et al.*, 2010; Yen *et al.*, 2011). However, this model has been shown to have lower transferability than other SDM methods, such as Generalised Linear Models (Heikkinen *et al.*, 2012; Crimmins *et al.*, 2013).

Different SDM methods may also have similar predictive performance (e.g. as quantified by the AUC or True Skill Statistic [TSS]; Allouche *et al.*, 2006) yet generate very different predictions of suitable habitat (Beaumont *et al.*, 2009; Parviainen *et al.*, 2009; Syphard & Franklin, 2009). These differences can be magnified when models are projected onto alternative climate scenarios (for which there is also no 'best' choice), with different SDMs varying in the magnitude and direction of predicted changes (Pearson *et al.*, 2006; Beaumont *et al.*, 2007; Beaumont *et al.*, 2008). Model performance can also be influenced by a species' characteristics and its distribution data (Syphard & Franklin, 2009; Dobrowski *et al.*, 2011; García-Callejas & Araújo, 2015), selection of predictor variables (Barbet-

Massin & Jetz, 2014) and geographic location (Engler *et al.*, 2011). As such, the choice of SDM method constitutes the primary source of variation in predictions of species' future distributions (Buisson *et al.*, 2010; Garcia *et al.*, 2012).

These factors have led to the current trend to combine multiple models into 'ensembles' (Araújo & New, 2007, Beaumont *et al.*, 2009, Marmion *et al.*, 2009b; Grenouillet *et al.*, 2011; Garcia *et al.*, 2012; Xinhai & Wang, 2013). Ensemble modelling experiments can then be used to distinguish regions of model consensus (i.e. where most models agree that the environment is suitable/unsuitable) from regions where there is disagreement. A number of studies have found that ensembles outperform individual models (Marmion *et al.*, 2009a; Yen *et al.*, 2011; Grenouillet *et al.*, 2011; Crossman *et al.*, 2012), however, the use of ensembles can still be problematic (Elith *et al.*, 2010; Rapacciuolo *et al.*, 2012; Crimmins *et al.*, 2013) as errors/biases in individual SDMs may lead to incorrect conclusions being drawn from the ensemble.

While previous studies have assessed sources of variation in SDM output (e.g., Dormann *et al.*, 2008; Nenzen & Araujo, 2011; Watling *et al.*, 2015), less attention has been given to whether particular SDM methods are biased towards predicting substantial loss or gain of suitable habitat, and if so, why. Therefore, in this study, we explored 'extreme' range changes predicted by 14 SDM methods incorporated into the R packages *biomod2* (Thuiller *et al.*, 2012) and *dismo* (Hijmans *et al.*, 2011). In particular, we asked whether these SDM methods predicted 'extreme' range changes with equal frequency. We defined five non-mutually

exclusive categories of extreme change, based on stability or loss of current habitat or the dislocation of current and future habitat: a) no future habitat (range extinction), b) low stability of current habitat (i.e.  $\leq 10\%$  of current habitat remains, although future habitat may be predicted in new locations), c) no gain of habitat in new locations (future habitat is identical to, or a subset of, current habitat), d) all future habitat is in new locations (future habitat does not overlap with current habitat), and e) substantial increase in size of habitat (i.e. future habitat is at least twice the size of current habitat). We modelled future habitat suitability for 220 mammal species across Australia and compared the proportion of predictions from each SDM method that calculated these extreme range changes.

## **Methods**

### *Species records*

For this study, we focused on native terrestrial mammal species in Australia, as their distributions are generally well known and their taxonomy has been resolved. Occurrence records for the 338 extant mammal species recorded on the National Species List (NSL) were downloaded from the Atlas of Living Australia (ALA, [www.ala.org.au](http://www.ala.org.au)). The ALA aggregates species records from a wide array of providers including the major museums across the country and maintains the NSL. We applied a number of filters before downloading data, requiring that records be resolved to species-level, observations were made after January 1950, and were

georeferenced. Observations were discarded if the referenced location was classified by ALA as an environmental outlier. After downloading, records were aggregated to a spatial resolution of  $5 \times 5$  km, and reduced to a maximum of one record per species, per cell. We excluded species for which there were fewer than ten occurrence records, since models fit with so few data may be unreliable (Stockwell & Peterson 2002; Hernandez *et al.*, 2006; van Proosdij *et al.* 2016). This left 220 species with between 10 and 7,137 records that, when combined, spanned the breadth of the continent (Supplementary Information Table S1). These species belong to seven orders: Chiroptera (bats,  $n = 54$  species), Dasyuromorphia (carnivorous marsupials,  $n = 46$ ), Diprotodontia (e.g. kangaroos, wallabies, possums,  $n = 66$ ), Monotremata (platypus, echidna,  $n = 2$ ), Notoryctemorphia (marsupial mole,  $n = 1$ ), Peramelemorphia (omnivorous marsupials such as bandicoots and bilbies,  $n = 5$ ) and Rodentia (native rats,  $n = 46$ ) (Supplementary Information Table S1).

#### *Current climate data*

Eight bioclimatic variables for the current period (1950-2000) were derived using ANUCLIM 5.1 (Houlder *et al.*, 2000), at a resolution of  $5 \times 5$  km. These variables were selected due to their influence on the physiological and distributional limits of vertebrates, and included annual mean temperature (T), temperature seasonality (TS), maximum temperature of the warmest month (Tmax), minimum temperature of coldest month (Tmin), annual precipitation (Pr), precipitation seasonality (PS), precipitation of the driest quarter (Pdry) and precipitation of wettest quarter (Pwet).

We recognise that there are correlations among these data. We have elected to utilise a common set of variables to provide consistency between models, thereby aiding direct comparisons. For multi-species studies such as ours where, combined, the target species cover a broad geographic region it is unlikely that a common set of predictor variables that are uncorrelated across all species can be identified. Generating distinct subsets for individual species would be worthwhile but is computationally challenging.

#### *Future climate data*

We utilised future climate data developed by Reside *et al.* (2013), for the 30-year period centred on 2055 and based on the RCP8.5 greenhouse gas concentration trajectory (the highest of the four Representative Concentration Pathways presented in the Intergovernmental Panel on Climate Change's Fifth Assessment Report, with radiative forcing exceeding  $8.5 \text{ Wm}^{-2}$  by 2100, see Moss *et al.*, 2010). We chose two global climate models (GCM) for comparison, which represented contrasting futures based on similarity to Australia's current climate in terms of average temperature and precipitation. In doing so, our aim was to take account of the existing variability in future climates and test the SDM methods with the output from two GCMs that represent the range of this variability. Of the 18 GCMs we examined, the National Centre for Atmospheric Research's Community Climate System Model v 3.0 (NCAR-CCSM3; Collins *et al.*, 2004) was the most similar to current temperature and

precipitation (averaged across the continent), whereas the Max Planck Institute for Meteorology's atmospheric general circulation model (MPI-ECHAM5; Roeckner *et al.*, 2006) extrapolated the most for temperature and was among the most different for precipitation.

### *Species distribution models*

To explore variability across alternate SDM methods, we used an ensemble of 14 algorithms included in the R packages *dismo* v1.0-5 (Hijmans *et al.*, 2011) and *biomod2* v3.1-64 (Thuiller *et al.*, 2012). These represent the gamut of approaches used by ecologists and conservation practitioners, and include (i) three regression-based approaches (Generalised Linear Models [GLM, McCullagh and Nelder, 1989], Generalised Additive Models [GAM, Hastie *et al.*, 1990], Multivariate Adaptive Regression Splines [MARS]), (ii) four machine-learning methods (Artificial Neural Networks [ANN, Lek & Guégan, 1999], Random Forests [RF, Breiman, 2001], Boosted Regression Trees [BRT, included in both packages, Friedman, 2001; Elith *et al.*, 2008], and Maxent [Phillips *et al.*, 2006]), (iii) two classification schemes (Classification Tree Analysis [CTA, Brieman *et al.*, 1984], Flexible Discriminant Analysis [FDA, Hastie *et al.*, 1994]), and (iv) four distance-based or profile methods (Surface Range Envelope [SRE], BIOCLIM [Nix, 1986; Busby, 1991], Domain [Carpenter *et al.*, 1993], Mahalanobis [Mahal, Farber & Kadmon, 2003]). Of the above, *biomod2* was used to fit models GLM, GAM, MARS, ANN, RF, CTA, FDA and SRE and the first implementation of BRT (denoted from now as BRT<sub>bio</sub>);

BIOCLIM, Domain, Mahal and the second implementation of BRT (denoted as BRT<sub>dis</sub>) were derived from *dismo*. Maxent was implemented separately via the command line. Characteristics of each algorithm, in the context of species distribution modelling, have been described in detail elsewhere (Elith *et al.*, 2006; Phillips *et al.*, 2006; Phillips & Dudik, 2008; Elith *et al.*, 2011; Hijmans & Elith, 2013; Xinhai & Wang, 2013), and default settings are listed in Supplementary Information Table S2.

We ran most models with their default settings (Supplementary Information Table S2). Maxent was run with threshold features disabled, while “fadebyclamping” was enabled for its predictions. Note also that the two BRTs differed in their implementation, with the *dismo* function using a cross-validation procedure to estimate the optimal number of boosting trees. Further, BRT<sub>dis</sub> has a default tree complexity of 1 (meaning that it fits only additive terms) while in BRT<sub>bio</sub> (where complexity is referred to as *interaction.depth*) this value is 7. As such, BRT<sub>bio</sub> allows up to seven-way interactions, and can thus fit models of far greater complexity. Similarly, SRE (*biomod2*) and BIOCLIM (*dismo*) are different modifications of Nix’s (1986) original BIOCLIM. This model is a straight-forward boxcar or parallelepiped approach, whereby the species envelope is bounded by the minimum and maximum values for all presences (or adjusted to exclude outliers) (Booth *et al.*, 2014). The default setting for this model (SRE) in *biomod2* excludes values above the 97.5th percentile, and below the 2.5th percentile for each variable, preventing these from setting the tolerance boundaries for the species (Thuiller *et al.*, 2012). As such,

predictions are binary (suitable/unsuitable). In contrast, in *dismo*'s BIOCLIM a greater range of percentiles are calculated, with the assumption that areas closer to the median are more suitable (Hijmans *et al.*, 2011), and allowing for output that is continuous.

*Background data:* Absence data were unavailable for this study, and it is highly likely that occurrence records sourced from natural history collections contain sampling biases. One approach to reducing the impact of these biases is to utilise background data that have the same sampling distribution as the occurrence records (i.e. target-group background) (Elith & Leathwick, 2007; Phillips & Dudik, 2008). Hence, background data comprised the occurrence records for all mammals. We note, however, that this approach may also inflate accuracy measures for species with narrow distributions, due to high specificity (see Lobo *et al.*, 2008).

*Model calibration and performance:* The predictive performance of models was assessed by the True Skill Statistic, TSS. In the context of SDMs, TSS is a threshold-dependent measure based on sensitivity (Se) and specificity (Sp), or the probability that the model correctly predicts true presences and true absences, respectively (Allouche *et al.*, 2006; Liu *et al.*, 2009). Specifically,  $TSS = Se + Sp - 1$ . Values therefore range from -1 to 1, with values below 0.4 indicating poor model discrimination between occurrence and background locations (i.e., Se and Sp are, on average, less than 0.7) and values close to 1 indicating excellent discrimination.



We used ten-fold cross-validation to estimate out-of-sample predictive performance. For each species, this entailed splitting model-fitting data into ten subsets of approximately equal size (i.e., folds), fitting each model to nine of the ten folds, and predicting to the tenth. This process was repeated until each fold had been used nine times for fitting and once for testing. TSS was calculated for the predictions to each test fold, with the average of the ten TSSs values used to estimate the performance of the final model (see below).

*Final models:* Models were calibrated with all available occurrence data and projected onto current climate and two climate futures for 2055 (described above). Continuous predictions were converted to binary layers indicating suitable/unsuitable habitat, with threshold values chosen to maximise the sum of sensitivity and specificity (equivalent to maximising TSS) as this threshold tends to well reflect the modelled species' prevalence (Jimenez-Valverde & Lobo, 2007). Further, this threshold is less sensitive to modelling method, and has been demonstrated to produce less variation across the output of multiple models than most other thresholds (Nenzen & Araujo, 2011). Models with an average cross-validated TSS below 0.4 were removed from further analyses, since these are considered to have low accuracy (see BIOMOD manual from Thuiller *et al.* 2012).

### *Statistical Analyses*

Changes in habitat suitability predicted by SDM methods can be summarised as the proportion of current habitat ( $cH$ ) that remains suitable in the future (i.e. is 'stable'), and the proportion of future habitat ( $fH$ ) that is located in areas currently unsuitable (i.e., 'gain' of new habitat). Based on these definitions of stability and gain, we assessed five non-mutually exclusive categories of extreme changes to the spatial configuration of suitable habitat. When graphed along these two axes models may predict a number of extreme outcomes (Fig. 1). For the purposes of this study, we categorised extreme to include the following, although we note that 'extreme' may be context dependent:

- a) No  $fH$  (i.e. range extinction);
- b) Low stability of  $cH$  (< 10% of  $cH$  remains stable, although gains may occur elsewhere);
- c) No gain of new habitat (i.e.  $fH$  is entirely contained within  $cH$ ; this excludes predictions that meet criterion (a) of no  $fH$ );
- d) All  $fH$  is gain (i.e. all  $fH$  is in previously unsuitable areas); and
- e)  $fH$  is at least double the size of  $cH$ .

For each of the above, we tested the null hypothesis that all 14 SDM methods used in this study predict that extreme scenario with equal frequency. Under the null hypothesis, the expected frequency of extreme <sub>$i$</sub> , predicted by SDM <sub>$j$</sub> , was calculated as the number of predictions made by SDM <sub>$j$</sub>  as a fraction of the total number of predictions (i.e. made by all 14 SDMs), multiplied by the total number of instances of

extreme<sub>i</sub>. Chi-squared Goodness-of-fit tests were used to assess the null hypothesis. When  $p < 0.05$ , we calculated the standardised residuals ( $\hat{r}_i$ ) of the contingency table to identify for which SDM(s) the observed frequency ( $f_o$ ) deviated significantly from the expected ( $f_e$ ). A larger  $\hat{r}_i$  means a greater difference between  $f_o$  and  $f_e$ , and provides an indication of the contribution of the SDM method to the significance of the Chi-squared test. For any SDM method with  $|\hat{r}_i| > 2$ ,  $f_o$  differs from  $f_e$  at a significance level of  $p < 0.05$ . Negative and positive  $\hat{r}_i$  indicate that  $f_o$  is less than or greater than  $f_e$ , respectively (Sheskin, 2004).

Scatterplots of stable habitat versus habitat gain were created to reveal any patterns amongst this relationship across either SDM method or taxonomic order. All statistics were calculated in R v. 3.1.2 (R Development Core Team, 2014).

## Results

### *SDM predictive power*

We calibrated 3,080 models (220 species  $\times$  14 SDM methods), of which 160 returned errors and were excluded from further analysis. These were primarily CTA (133 models) and ANN (28). In each case, the model failed due to a single value being predicted (i.e. all presences or all absences), resulting from the model being unable to discriminate between these. For the remaining 2,920 models, predictive performance measured using TSS averaged 0.784 (SD = 0.158), although this varied considerably across the 14 algorithms (Fig. 2a). Classification methods

generally had poorer performance, in terms of TSS, than machine learning techniques (ranging from mean TSS for CTA  $0.537 \pm \text{S.D. } 0.240$  to mean TSS for RF  $0.997 \pm \text{S.D. } 0.008$ ).

We excluded from further analyses 226 models (7.7%) that did not meet the TSS threshold of 0.4. Of these, 68 were modelled by Mahal and 24 by CTA. In contrast, all RF models, and all but one GBA and Maxent model exceeded the threshold. Hence, the number of projections in our final analysis was 2,694 (Table 2).

Among the five mammal taxonomic orders, mean TSS values were lowest for Notoryctemorphia, which is represented here by a single species ( $0.634 \pm 0.183$ ), and highest for Diprotodontia ( $0.822 \pm 0.137$ ) and Peramelemorphia ( $0.829 \pm 0.107$ ) (Supplementary Information Fig. S1).

In general, distance-based methods tended to predict a larger area of current habitat than other SDM methods, with projections by Mahal being, on average, two orders of magnitude greater ( $3.03 \times 10^6 \pm 2.27 \times 10^6 \text{ km}^2$ ), than those from RF ( $3.55 \times 10^4 \pm 7.16 \times 10^4 \text{ km}^2$ ) (Fig. 2b).

### *Predictions of future distributions - overview*

SDM methods differed in how frequently they projected extreme range changes. Across all species, SDM methods and climate futures, changes to the overall area of suitable habitat were right skewed, with 66.3% of models predicting declines to the area of *fH* relative to *cH* (median change = -25.3% [10:90<sup>th</sup> percentiles: -90 to 121.8%]). Only 6.3% of models predicted that the area of *fH* would be similar to that

of *cH* (i.e. within 5%) while 4.2% predicted there to be no suitable habitat in the future (extreme *a*). At the other end of our spectrum of extremes, 11.5% of models predicted that the area of *fH* would be at least double that of *cH* (extreme *e*), and more than half of these (7.7%) predicted the area of suitable habitat to triple.

Of course, range changes comprise a mix of contraction of *cH* and expansion into new areas (i.e. 'gain'). Total loss of *cH* was projected for ~9% of models, with an additional 10% predicting that  $\leq 10\%$  of *cH* would remain by 2050 (extreme *b*). Approximately 5.6% of projections reported no gain in *fH* (extreme *c*) while 5.4% predicted that all *fH* would lie in currently unsuitable areas (extreme *d*).

Projections onto ECHAM5 (which was the least similar to current conditions) predicted greater and more variable changes in range size than projections onto the CCSM3 climate (ECHAM5: median -38.7% [10-90th percentile; -94.8%, 167.4%]; CCSM3: median -11.6% [10-90th percentile; -80.5%, 84.1%]).

Four mammal orders were represented by > 5 species (Chiroptera, Dasyuromorphia, Diprotodontia, Rodentia). For Chiroptera, < 1% and 2% of projections predicted no *fH* (CCSM3 and ECHAM5) while 12.8% and 21.6% predicted range area to more than double. Across the models for Diprotodontia, 6% predicted no *fH*, while 7.1% predicted *fH* to double (ECHAM5). Current suitable habitat was predicted to decline substantially (> 90%) for 27% of Rodentia models while 17% predicted *fH* to double (ECHAM5).

*Do SDM methods project extreme spatial changes with equal frequency?*

We assessed the hypothesis that the 14 SDM methods were equally likely to project particular range changes, i.e.  $f_e$  (expected frequency) of a given extreme equals the number of projections of that extreme by  $SDM_i$  as a fraction of all SDM projections. In total, 5,388 projections of future distributions were analysed (2,694 for each climate future). The total number of models with sufficient predictive capacity ( $TSS \geq 0.4$ ) varied across algorithms, resulting in  $f_e$  ranging from 0.023 (CTA) to 0.082 (RF) (Table 1).

Chi-squared test statistics ranged from 49.3 to 297.6 for the five categories of extreme change and were significant to at least  $p = 0.0001$  (Table 3). This demonstrates that the 14 SDM methods are unequally represented in each category (Supplementary Information Table S2). Across the five categories, RF was frequently the most under- or over-represented method.

*a) No future habitat (fH)*

Of the 2,694 models projected onto both climate futures (ECHAM5 and CCSM3), 144 (5.3%) and 80 (3.0%) predicted total loss of  $fH$ , respectively. RF accounted for approximately half of these, making it significantly more likely than the other methods to return this result ( $\hat{\chi}^2 = 12.85, 16.13$  for ECHAM5 and CCSM3, respectively), although SRE was also significantly over-represented in the ECHAM5 climate future ( $\hat{\chi}^2 = 5.04$ ). In contrast, MARS, Mahal and  $BRT_{dis}$  were significantly under-represented ( $\hat{\chi}^2 -2.187$  to  $-3.13$ ) with 0-3 projections each within this category.

Observed frequencies for ANN, BIOCLIM, CTA, Domain and BRT<sub>bio</sub> did not differ significantly from expected ( $|\hat{\eta}| < 2$ ) (Fig. 3).

*b) Low stability of current habitat (cH)*

Current suitable habitat was projected to decline in size by at least 90% for 575 (21.3%) and 318 (11.8%) models under the ECHAM5 and CCSM3 future climates, respectively. Again, RF was significantly over-represented in both future climates, comprising 17.7 and 30.5% of these projections ( $\hat{\eta} = 8.38$ ; 14.54). Indeed, over 40% of RF projections fell into this category. CTA was also over-represented, with 33.3% ( $\hat{\eta} = 3.46$ ; CCSM3) and 41.3% ( $\hat{\eta} = 5.03$ ; ECHAM5) of the 63 CTA models predicting extensive loss of *cH*. In comparison, BRT<sub>dis</sub>, GLM, MARS and Maxent were significantly under-represented, with between 2.9-13.4% of their projections falling into this category (with  $\hat{\eta}$  ranging from -2.52 to -4.051). Observed frequencies of both FDA and GAM did not differ significantly from expected for either climate future.

*c) No gain of new habitat*

This category, in which ~2% of projections fell, denotes models for which *fH* is entirely contained within areas currently classified as suitable. Significantly under-represented in this category were projections from BIOCLIM ( $\hat{\eta} = -2.32$ ; -2.13, ECHAM5 and CCSM3 respectively) and SRE, ( $\hat{\eta}$  ranging from -2.30 to -2.11), while GLM was over-represented ( $\hat{\eta} = 3.44$ , 3.61, ECHAM5 and CCSM3, respectively). Under the ECHAM5 climate future, ANN and Maxent were also over-represented, while FDA was for CCSM3. BRT<sub>dis</sub>, CTA, GAM, BRT<sub>bio</sub> and Mahal did not differ from expected for either climate future.

d) *All fH is gain*

For ECHAM5 and CCSM3, respectively, 231 (8.6%) and 105 (3.9%) models projected that all *fH* would lie in areas currently classified as unsuitable, that is, that suitable habitat by 2055 would be completely disjunct from current habitat. Again, RF was significantly over-represented in both climate futures ( $\hat{\eta} = 5.56, 9.06$ ), as was CTA ( $\hat{\eta} = 6.36, 6.16$ ) and, to a lesser extent, BRT<sub>bio</sub> ( $\hat{\eta} = 2.20, 3.74$ ). These three algorithms combined accounted for 38% (CCSM3) and 62% (ECHAM5) of the projections that fell into this category. Mahal, MARS and Maxent were significantly under-represented (with  $\hat{\eta}$  ranging from -2.29 to -3.80), while the frequencies for ANN, BIOCLIM, Domain, and GAM did not differ from expected.

e) *fH is, at least, double the size of cH*

This category consisted of 13.9% (ECHAM5) and 9.1% (CCSM3) of projections. There was substantial variation across SDMs in the frequency with which their projections fell within this category. For instance, none of the models fitted using the four distance-based methods (BIOCLIM, Domain, Mahal, SRE) were included in this category for either climate future ( $\hat{\eta}$  between -3.82 and -5.61). However, BRT<sub>bio</sub> ( $\hat{\eta} = 6.36, 4.96$ ) and RF ( $\hat{\eta} = 6.13, 6.10$ ), and to a lesser extent FDA and GAM, were significantly over-represented. Indeed, for both BRT<sub>bio</sub> and RF, 29% of their projections under the ECHAM5 climate future predicted *cH* to at least double in area. BRT<sub>dis</sub>, CTA and Maxent were the only models for which  $f_o$  did not differ significantly from  $f_e$ . These patterns are clear in scatterplots of the proportion of *cH* remaining stable versus the proportion of *fH* in new habitat: distance-based methods have few



points in the upper-right quadrant in contrast to RF (Supplementary Information Fig. S2).

#### *Consistency of patterns across taxonomic orders*

For each SDM, we visually assessed consistency in projections of stability and gain of new habitat across the various mammal orders. Within SDMs, the relationship between stability and gain was generally consistent across taxonomic orders (Supplementary Information Fig. S2). For some SDMs, there was an apparent negative relationship between these variables, which was most pronounced for distance-based methods (BIOCLIM, Domain, Mahal, SRE). When comparing GCMs, there were no clearly discernible differences in these relationships, though retention of existing suitable habitat in some cases appeared to be greater for CCSM3 than for ECHAM5 (which presented a climate future more different to current conditions than CCSM3).

#### **Discussion**

Species distribution models differ significantly from one another in the frequency with which they project extreme changes to the future size and configuration of suitable habitat. These changes (such as the total loss of current suitable habitat, future habitat that is completely disjunct from current habitat, and substantial increases in the future size of habitat) are extreme from a spatial perspective only – this does not

mean that such outcomes are rarely projected. To the contrary, approximately one-quarter of our model predictions fell into at least one category of extreme.

*Which SDM methods projected extreme range changes more (or less) frequently than others, and why?*

Such dramatic changes to the size of species' climatically suitable habitat, due to climate change projected to occur in coming decades, is disconcerting. Yet, this outcome has been projected for numerous species across a multitude of studies (e.g. Ihlow *et al.*, 2012; Ochoa-Ochoa *et al.*, 2012; Warren *et al.*, 2013; Krause *et al.*, 2015). However, our results demonstrate that the frequency with which such extreme range changes are projected may be related to the choice of SDM. For instance, we found that RF projected substantial range loss for almost half of the 220 species studied - a frequency significantly at odds with other SDM methods. Similarly, CTA and SRE were significantly more likely to project loss of current habitat (*cH*), while CTA and RF frequently projected future habitat (*fH*) to be located in areas currently unsuitable. In stark contrast to the other methods, the four distance-based approaches (BIOCLIM, Domain, Mahal, SRE) never projected extreme increases in the area of suitable habitat.

These results are a function of how a model fits the species-environment relationship and selects from competing, and potentially correlated, predictors. It may also be a function of model parameterisation and the details of extrapolation to environments beyond those represented by the training data.

The goal of SDM studies is to describe accurately and precisely the species-environment relationship and, frequently, to project this relationship onto data for other time periods or geographic regions. Given the same input data, the outputs of competing methods will differ because their fundamental approaches to describing the species-environment relationship and capturing interactions among variables, as well as their parameterisation and ability to interpolate/extrapolate, also differ (Merow *et al.*, 2014). For example, BIOCLIM, a simple boxcar approach, generally sets a species' environmental tolerance to the most extreme values in the training records, making this method very sensitive to outliers or insufficient sampling of the environment. Interactions between variables are not considered; all variables are weighted equally and extrapolation beyond the environmental envelope is not possible. Further, all areas within the envelope are deemed equally suitable, although note that the *dismo* version of BIOCLIM calculates a greater range of percentiles from the training data, with the assumption that grid cells closer to the 50<sup>th</sup> percentile are more suitable (Hijmans *et al.*, 2011).

In contrast, machine learning methods, such as Maxent and RF, can identify non-linear species-environment relationships and may be more efficient at modelling complex patterns, possibly leading to more accurate predictions than GLMs or GAMs (Thibaud *et al.*, 2014). Unless model complexity is managed, however (e.g., with regularisation; Warren *et al.* 2011), machine learning techniques are prone to describing noise in the training data, leading to poor generality.

Therefore, the extent to which a model can estimate realistic response curves is critical, and errors may result in models that are under- or over-fitted (for a description of simple and complex response curves from different methods, see Table 1 in Merow *et al.*, 2014). Simplistic models that fail to capture the underlying relationship are said to under-fit. These models have low variance, in that they can reproduce the result consistently, but are highly biased, i.e. model-fitting data are poorly predicted. Conversely, over-fitting occurs when noise or random error within the data is described by the model (Radosavljevic & Anderson, 2014), generating unrealistically complex response curves. In this instance, the model may describe the occurrence pattern accurately (i.e., it has low bias, and hence high predictive performance, with respect to model-fitting data), but transferability to other locations or time periods may not be equally high. Both under- and over-fitting may lead to poor predictions across space and time (Merow *et al.*, 2014).

Of interest is the propensity for RF to report very high TSS or AUC values while also projecting extreme range changes (also see Guo *et al.*, 2015). Previous studies have found this model to interpolate accurately but extrapolate or transfer poorly (e.g., see Wenger & Olden, 2012). This model produces very jagged response curves that may fit data well but are biologically unrealistic, and are unlikely to transfer (Wenger & Olden, 2012).

Typically, the predictive performance of a model is measured by splitting data into training and test sets. However, the two subsets are rarely independent (Veloz 2009; Radosavljevic & Anderson, 2014). Hence, performance measures derived

from them may be optimistic, and fail to reflect the performance of a model when it is projected onto environmental data from different regions or time periods. Crimmins *et al.* (2013) used five SDM methods to model the distributions of 145 plant species. Models calibrated and tested using cross-validation procedures, where 25% of data were randomly selected and withheld from calibration, were compared to models tested with temporally independent data. They found that, as with other studies (e.g. Grenouillet *et al.*, 2011; Bucklin *et al.*, 2015; Guo *et al.*, 2015), RF returned the highest AUC values when tested with non-independent data (i.e. via cross-validation) (mean ~0.98). However, for all models, AUC values were lower when tested with the independent data, with RF the second lowest (mean ~0.77) indicating poor transferability. This was primarily the result of high rates of false positives (median ~33%), with one-quarter of RF models having an FP rate of 1.0. Similarly, Heikkinen *et al.*, (2012) assessed the interpolation and extrapolation ability of 10 SDM methods against spatially independent data. Interpolation accuracy was highest for RF, Maxent and BRT<sub>bio</sub>. Extrapolation proved more difficult for all methods, particularly RF and MARS.

The way in which the fitted functions of a model are constrained during extrapolation will also influence its performance when doing so (Elith & Graham, 2009). For instance, Maxent by default utilises a 'clamping' approach whereby the response to environmental variables beyond their calibration range is kept at a constant value (Elith & Graham, 2009). Similarly, CTA, BRT<sub>dis</sub> and RF extrapolate at a constant value from the most extreme environmental value (Elith & Graham, 2009).

GLMs with cubic and quadratic functions continue the fitted trend beyond the most extreme environmental value, which can cause unrealistic results, although options do exist to control extrapolation (Elith & Graham, 2009). Stohlgren *et al.* (2011) note that restricting extrapolation by CART and Maxent can greatly reduce gain of habitat in new locations, and suggest that bounding techniques be included in best practices for SDMs. In contrast to the above models, distance-based measures tend not to extrapolate to conditions outside their training data, instead assuming these conditions are unsuitable (e.g. see Pearson *et al.*, 2006).

An additional consideration is multicollinearity. We used the same eight variables for each SDM but the direction and magnitude of collinearity among these varies spatially and temporally. For instance, across northern Australia, the driest months are generally in winter while to the south they are in summer. The consequences of collinearity depend on the method, but in general, the accuracy and precision of estimated coefficients (and thus partial response curves) are lower for correlated predictors than for uncorrelated predictors. Further, small changes in values of the input data can result in large changes in the model. This can lead to unreliable projections onto alternate climate scenarios, particularly if correlations change in direction (Elith *et al.*, 2010).

#### *Implications for projections of potential range shifts*

How sensitive are projections of range changes to model selection and parameterisation? Our results demonstrate that the frequency with which extreme

range changes are predicted can vary significantly across SDM methods. To an (unknown) extent, this may be due to the choice of model settings as can be seen by comparing the results for  $BRT_{bio}$  and  $BRT_{dis}$ . The frequency with which these methods projected extreme range changes differed substantially. For instance, while  $BRT_{bio}$  was significantly more likely than expected to project substantial declines to  $cH$  and that  $fH$  would be mostly disjunct from  $cH$ ,  $BRT_{dis}$  had the opposite response, being significantly less likely than other methods to project these extremes. Further,  $BRT_{bio}$  frequently projected  $fH$  to be at least double the size of  $cH$ , while  $BRT_{dis}$  was unlikely to project no  $fH$ .

To date, there is little guidance on approaches to parameterising SDMs (although see Merow *et al.*, 2014), and for some studies parameterisation of each model for each species may be prohibitively time-consuming or, in the case of small or biased datasets, may be unreliable (Phillips & Dudik, 2008). Hence, default settings for numerical model parameters are typically relied upon, with the assumption that they are appropriate as they were previously tested on large datasets and were found to perform well (e.g., see Elith *et al.*, 2006; Phillips & Dudik, 2008).

Many of the settings used in the R packages for SDMs are the program's default values (e.g., as with FDA within *biomod2*; pers.comm. W. Thuiller), while others may be the result of trial and error by package authors. For example, within *biomod2*, the default value for the maximum number of iterations used by ANN was 200. This value was derived experimentally and is the result of a trade-off between

computing time and efficiency (pers.comm. W. Thuiller). Similarly, default settings for Maxent were selected based on assessments across diverse datasets (Phillips & Dudik, 2008). Default settings, however, should be used with caution, since they do not always result in optimal models (e.g., Moreno-Amat *et al.*, 2015). Further, experimentally derived settings are usually optimised according to their ability to accurately predict independent test data rather than their accuracy when projecting to other locations or time periods. However, the purpose that model settings are tuned for is not necessarily the purpose for which the models are being used. That is, a model with high predictive performance based on current distributional data may not be particularly effective at predicting the effects of climate change, and so default values, or “best practices”, may not be applicable depending on the application of the model.

Systematic evaluations of alternative values of settings for individual algorithms are rarely undertaken. In their review of the SDM literature, Hallgren and Mackey (2014) found that the majority of SDM studies do not provide values for algorithm configuration options, and when available the justifications for these values were usually minimal. The overall lack of documentation of the rationale for values for model configuration options has the potential to be of concern. Without knowing how sensitive a model - and model output - is to these values, then arbitrarily assigned values and the lack of a tested range of suitable values could lead, in an unpredictable manner, to erroneous and/or unrealistic model results (Hallgren and Mackey, 2014). Optimisation of most models is possible, however, comparisons



within and between models undergoing optimisation may be complex (Merow *et al.*, 2014). Further, optimisation has generally not been automated within SDM packages (but see BRT<sub>dis</sub>) hence, large multi-species studies may be limited to default settings.

## **Conclusion**

Crucially, our study shows that model selection and parameterisation will influence inferences of the likelihood of species facing extreme changes to the distribution of suitable habitat as climate changes. To summarise, we found that compared to other models, distance-based measures are significantly less likely to project massive increases to the size of suitable habitat, while SRE is highly likely to project no future habitat; CTA projects substantial loss of current habitat and that future habitat will be disjunct from current habitat more frequently than expected, while Maxent predicts these outcomes less frequently than expected; RF frequently projects extreme range changes – indeed this model is the least likely to project stability of current habitat under alternative climate scenarios.

For some models, these patterns may be influenced by parameterisation, the impact of predictor collinearity on the model, and the way in which models extrapolate beyond the training data. A comprehensive sensitivity analysis of the predictions of SDM methods to their settings would provide valuable guidance on setting sensible parameter values for the models. Future developments in coding to automate optimisation procedures (similar to BRT<sub>dis</sub>) will greatly assist with fine-

tuning models used for multi-species studies rather than relying on a 'one-size-fits-all' approach.

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**Table 1. Number of models per SDM method.** 220 mammal species were modelled using 14 SDM methods. Projections with TSS values < 0.4 were excluded from further analyses, leaving from 63 to 220 species modelled by each method. For a given method, the expected frequency of each type of extreme change in habitat area (see Table 2) is equal to the proportion of all models (2,694) that were fit using that method.

<b>SDM</b>	<b>No. of models</b>	<b>Expected Frequency</b>
ANN	178	0.066
BIOCLIM	209	0.078
BRT <sub>bio</sub>	219	0.081
BRT <sub>dis</sub>	214	0.079
CTA	63	0.023
Domain	208	0.077
FDA	186	0.069
GAM	215	0.080
BRT <sub>bio</sub>	219	0.081
GLM	201	0.075
Mahal	152	0.056
MARS	205	0.076
Maxent	219	0.081
RF	220	0.082
SRE	205	0.076
<b>Total</b>	<b>2,694</b>	<b>1.000</b>

Table S3 is going here in final version.

**Table 3. Chi-squared statistic for each of the ‘extreme’ range change categories assessed.** The statistic is reported for two climate futures (ECHAM5 and CCSM3). *cH* = current habitat; *fH* = future habitat. In all instances,  $p < 0.0001$ .  $df = 13$ .

Extreme range change	$\chi^2$ statistic	
	ECHAM5	CCSM3
No <i>fH</i>	230.9	282.8
Low stability of <i>cH</i>	154.8	297.6
No gain of new <i>fH</i>	72.3	49.3
All <i>fH</i> is gain	147.4	163.0
<i>fH</i> at least double size of <i>cH</i>	217.1	164.9

## Figures

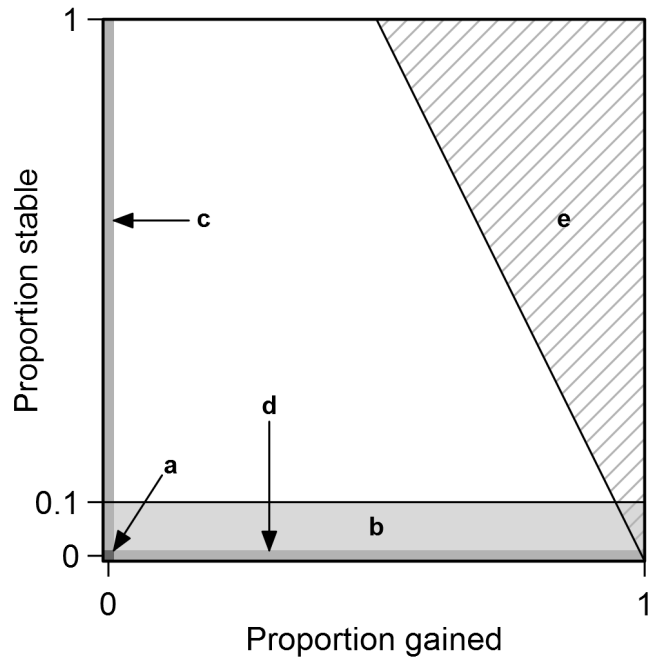
**Figure 1.** Changes in habitat suitability projected by species distribution models can be summarised along two axes: ‘Proportion stable’ (i.e. proportion of the area of current habitat [ $cH$ ] projected to remain suitable in the future) and ‘Proportion gained’ (i.e. proportion of the area of future habitat [ $fH$ ] projected to occur in currently unsuitable regions). Based on these two axes we calculated the number of predictions from 14 SDM methods that fell into the following categories: a) no  $fH$  (Prop. stable = 0, Prop. gained = 0); b) low stability of  $cH$  (Prop. stable  $\leq 0.1$ , Prop. gained  $> 0$ ); c) no gain of new habitat (Prop. stable  $> 0$ ; Prop. gained = 0); d) all  $fH$  is currently unsuitable (Prop. stable = 0, Prop. gained  $> 0$ ); and e) substantial increase in area of  $fH$  compared to  $cH$  ( $fH \geq 2cH$ ).

**Figure 2.** Box-and-whisker plot of AUC and size of current habitat projected for 220 Australian mammal species by 14 SDM methods. SDM methods can be broadly placed into one of four categories: distance-based, classification, regression, and machine learning. The dotted line in 2A indicates the TSS threshold (0.4) below which models were excluded due to poor predictive performance.

**Figure 3.** Proportional change in size of suitable habitat for Australian mammals under two climate futures. 14 SDM methods were calibrated for 220 species. Models with poor performance (TSS  $< 0.4$ ) were excluded from these analyses, resulting in 63–220 species being modelled per SDM method (see Table 1). Data are presented on a cube-root scale.

**Figure 4.** Standardised residuals from Chi-squared goodness-of-fit tests. Chi-squared tests were used to assess whether 14 SDM methods vary from each other in the frequency with which they project five categories of extreme changes to the size of habitat for 220 Australian mammal species, under two climate futures. Standardised residuals can be used to identify which SDM method contributed the most to the significance of the Chi-squared test. Here, residuals more extreme than  $\pm 2$  indicate that the observed frequency of the SDM methods differed significantly from expected ( $p < 0.05$ ). This threshold is represented by the two dotted lines. Figure 1 describes the categories of extreme range changes.

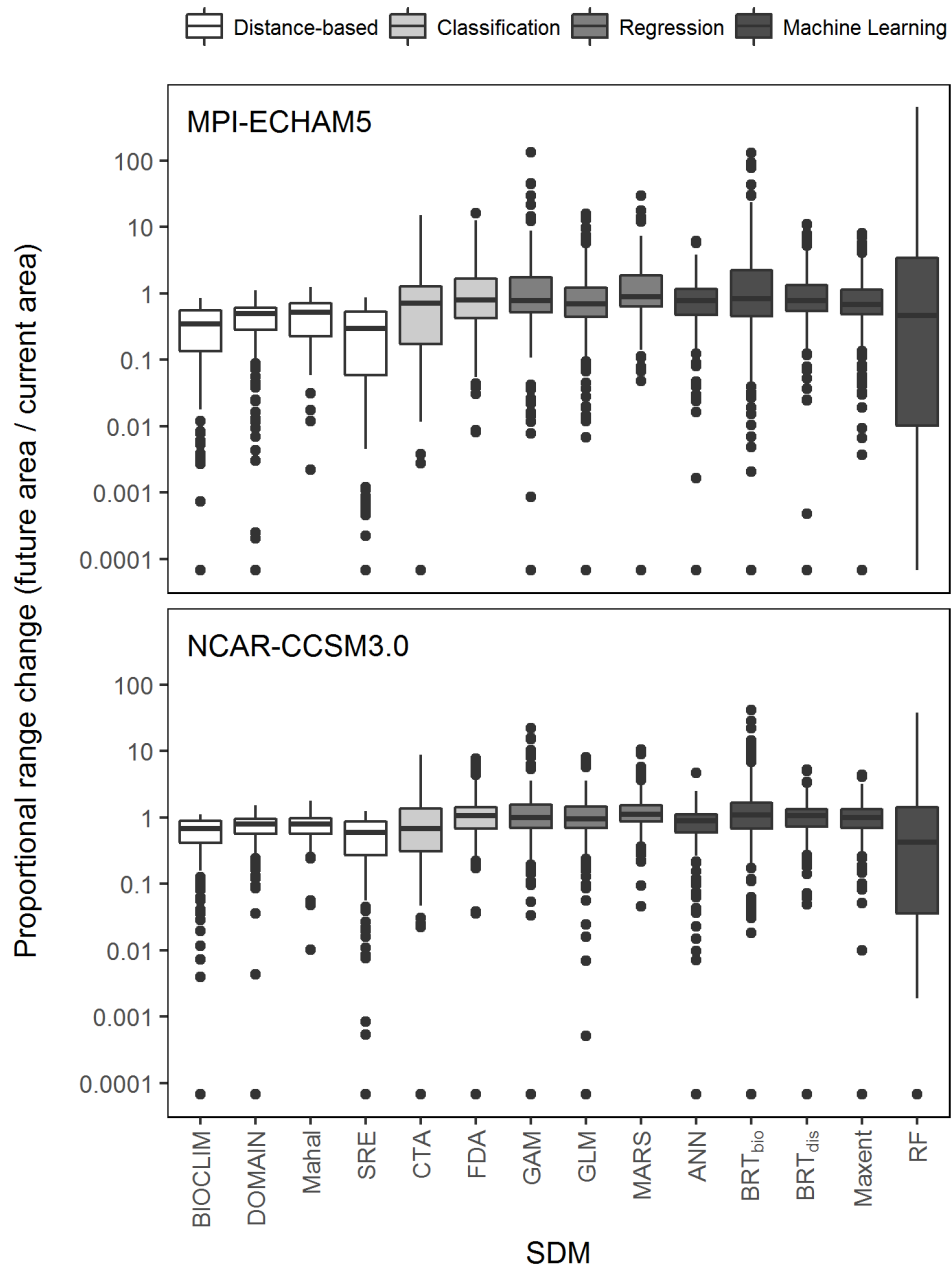
Figure 1.



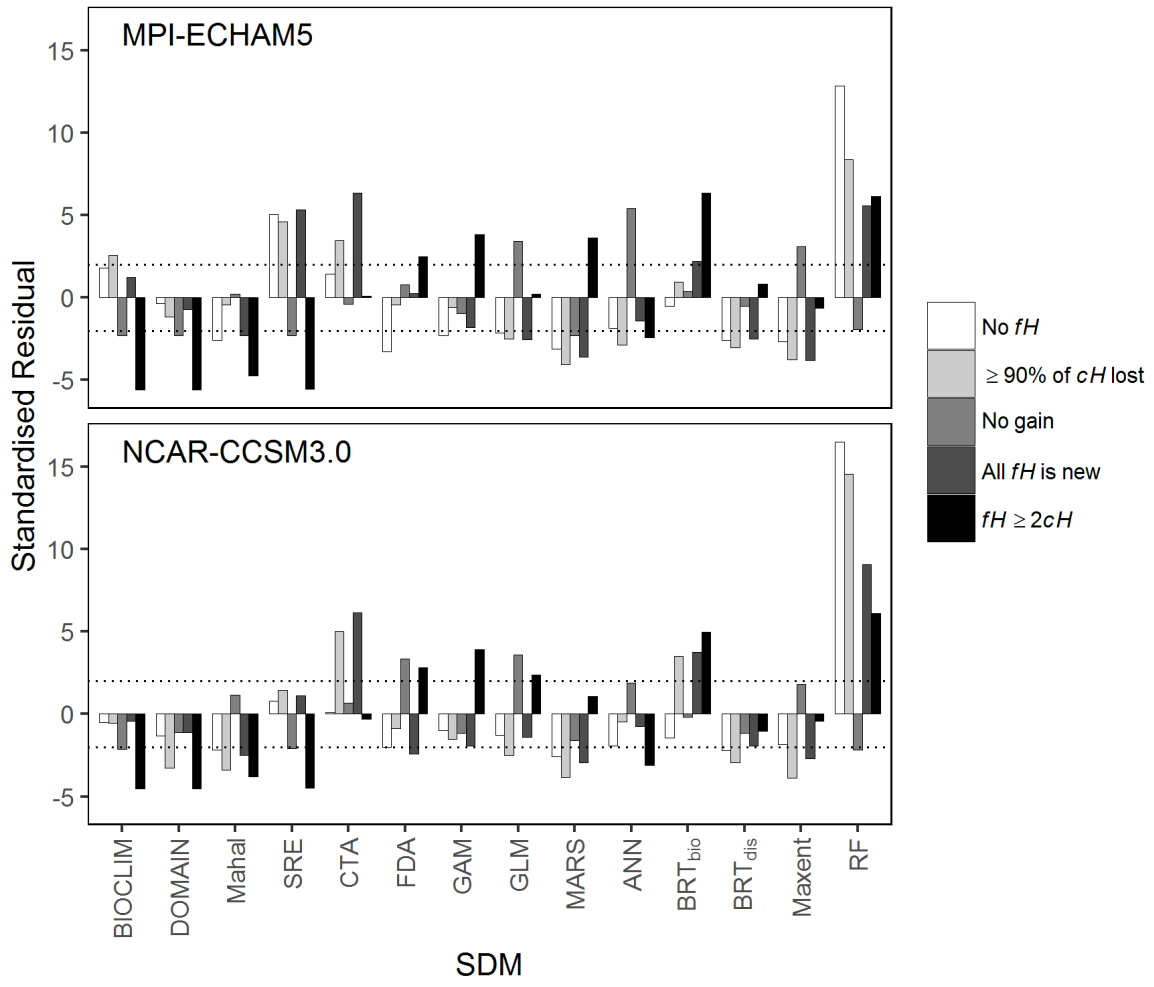
**Figure 2.**

**Figure 3.**





**Figure 4.**



## Which species distribution models are more (or less) likely to project broad-scale, climate-induced shifts in species ranges?

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## Supplementary Information

**Table S1.** 220 native Australian mammal species, their taxonomic order, and number of occurrence records used to model habitat suitability.

Order	Species	No. Records
Chiroptera		
	<i>Chaerephon jobensis</i>	211
	<i>Chalinolobus dwyeri</i>	308
	<i>Chalinolobus gouldii</i>	3,532
		50

<i>Chalinolobus morio</i>	2,326
<i>Chalinolobus nigrogriseus</i>	373
<i>Chalinolobus picatus</i>	303
<i>Falsistrellus tasmaniensis</i>	642
<i>Hipposideros ater</i>	135
<i>Hipposideros diadema</i>	27
<i>Hipposideros stenotis</i>	35
<i>Kerivoula papuensis</i>	264
<i>Macroderma gigas</i>	167
<i>Macroglossus minimus</i>	68
<i>Miniopterus australis</i>	739
<i>Miniopterus schreibersii</i>	1,411
<i>Mormopterus beccarii</i>	125
<i>Murina florium</i>	11
<i>Myotis macropus</i>	596
<i>Nyctimene robinsoni</i>	102
<i>Nyctophilus arnhemensis</i>	130
<i>Nyctophilus bifax</i>	186
<i>Nyctophilus geoffroyi</i>	3,301
<i>Nyctophilus gouldi</i>	1,724
<i>Nyctophilus timoriensis</i>	281
<i>Nyctophilus walkeri</i>	85
<i>Pipistrellus adamsi</i>	86
<i>Pipistrellus westralis</i>	39
<i>Pteropus alecto</i>	431
<i>Pteropus conspicillatus</i>	89
<i>Pteropus poliocephalus</i>	999
<i>Pteropus scapulatus</i>	643
<i>Rhinolophus megaphyllus</i>	906
<i>Rhinonictes aurantia</i>	105
<i>Saccolaimus flaviventris</i>	724
<i>Scoteanax rueppellii</i>	611
<i>Scotorepens balstoni</i>	849
<i>Scotorepens greyii</i>	829
<i>Scotorepens orion</i>	703
<i>Scotorepens sanborni</i>	43
<i>Syconycteris australis</i>	125
<i>Tadarida australis</i>	2,401
<i>Taphozous australis</i>	24
<i>Taphozous georgianus</i>	375
<i>Taphozous hilli</i>	98
<i>Taphozous kapalgensis</i>	18
<i>Vespadelus baverstocki</i>	302
<i>Vespadelus caurinus</i>	180
<i>Vespadelus darlingtoni</i>	1,221

<i>Vespadelus douglasorum</i>	34
<i>Vespadelus finlaysoni</i>	327
<i>Vespadelus pumilus</i>	827
<i>Vespadelus regulus</i>	1,386
<i>Vespadelus troughtoni</i>	129
<i>Vespadelus vulturnus</i>	2,422
Dasyuromorphia	
<i>Antechinomys laniger</i>	285
<i>Antechinus adustus</i>	21
<i>Antechinus agilis</i>	893
<i>Antechinus bellus</i>	114
<i>Antechinus flavipes</i>	1,301
<i>Antechinus godmani</i>	25
<i>Antechinus leo</i>	15
<i>Antechinus minimus</i>	175
<i>Antechinus stuartii</i>	1,296
<i>Antechinus subtropicus</i>	13
<i>Antechinus swainsonii</i>	736
<i>Dasyercus cristicauda</i>	62
<i>Dasyuroides byrnei</i>	20
<i>Dasyurus geoffroii</i>	140
<i>Dasyurus hallucatus</i>	446
<i>Dasyurus maculatus</i>	2,724
<i>Dasyurus viverrinus</i>	629
<i>Ningau ridei</i>	373
<i>Ningau yvonneae</i>	264
<i>Phascogale calura</i>	64
<i>Phascogale pirata</i>	34
<i>Phascogale tapoatafa</i>	650
<i>Planigale gilesi</i>	197
<i>Planigale ingrami</i>	221
<i>Planigale maculata</i>	366
<i>Planigale tenuirostris</i>	210
<i>Pseudantechinus bilarni</i>	73
<i>Pseudantechinus macdonnellensis</i>	201
<i>Pseudantechinus ningbing</i>	40
<i>Sarcophilus harrisii</i>	1,336
<i>Sminthopsis aitkeni</i>	10
<i>Sminthopsis bindi</i>	37
<i>Sminthopsis butleri</i>	21
<i>Sminthopsis crassicaudata</i>	1,628
<i>Sminthopsis dolichura</i>	350
<i>Sminthopsis gilberti</i>	85
<i>Sminthopsis griseoventer</i>	127
<i>Sminthopsis hirtipes</i>	125

<i>Sminthopsis leucopus</i>	174
<i>Sminthopsis longicaudata</i>	28
<i>Sminthopsis macroura</i>	1,234
<i>Sminthopsis murina</i>	823
<i>Sminthopsis ooldea</i>	364
<i>Sminthopsis psammophila</i>	46
<i>Sminthopsis virginiae</i>	168
<i>Sminthopsis youngsoni</i>	303
Diprotodontia	
<i>Acrobates pygmaeus</i>	1,041
<i>Aepyprymnus rufescens</i>	418
<i>Bettongia gaimardi</i>	421
<i>Bettongia penicillata</i>	65
<i>Bettongia tropica</i>	26
<i>Burramys parvus</i>	21
<i>Cercartetus caudatus</i>	21
<i>Cercartetus lepidus</i>	157
<i>Cercartetus nanus</i>	398
<i>Dactylopsila trivirgata</i>	88
<i>Dendrolagus bennettianus</i>	25
<i>Dendrolagus lumholtzi</i>	60
<i>Gymnobelideus leadbeateri</i>	28
<i>Hemibelideus lemuroides</i>	53
<i>Hypsiprymnodon moschatus</i>	71
<i>Lasiorhinus latifrons</i>	467
<i>Macropus agilis</i>	729
<i>Macropus antilopinus</i>	239
<i>Macropus bernardus</i>	56
<i>Macropus dorsalis</i>	84
<i>Macropus eugenii</i>	112
<i>Macropus fuliginosus</i>	3,412
<i>Macropus giganteus</i>	3,139
<i>Macropus parma</i>	218
<i>Macropus parryi</i>	182
<i>Macropus robustus</i>	3,117
<i>Macropus rufogriseus</i>	2,503
<i>Macropus rufus</i>	5,081
<i>Onychogalea fraenata</i>	18
<i>Onychogalea lunata</i>	27
<i>Onychogalea unguifera</i>	441
<i>Petauroides volans</i>	1,713
<i>Petaurus australis</i>	1,356
<i>Petaurus breviceps</i>	3,031
<i>Petaurus gracilis</i>	63
<i>Petaurus norfolcensis</i>	786

<i>Petrogale assimilis</i>	59
<i>Petrogale brachyotis</i>	230
<i>Petrogale concinna</i>	38
<i>Petrogale godmani</i>	28
<i>Petrogale herberti</i>	24
<i>Petrogale inornata</i>	38
<i>Petrogale lateralis</i>	308
<i>Petrogale mareeba</i>	33
<i>Petrogale penicillata</i>	367
<i>Petrogale persephone</i>	11
<i>Petrogale purpureicollis</i>	21
<i>Petrogale rothschildi</i>	34
<i>Petrogale sharmani</i>	11
<i>Petrogale xanthopus</i>	207
<i>Petropseudes dahli</i>	93
<i>Phascolarctos cinereus</i>	3,258
<i>Potorous longipes</i>	19
<i>Potorous tridactylus</i>	562
<i>Pseudocheirus peregrinus</i>	2,377
<i>Pseudochiroptis archeri</i>	79
<i>Pseudochirulus cinereus</i>	15
<i>Setonix brachyurus</i>	68
<i>Spilocuscus maculatus</i>	12
<i>Thylogale billardieri</i>	821
<i>Thylogale stigmatica</i>	242
<i>Thylogale thetis</i>	330
<i>Trichosurus caninus</i>	601
<i>Trichosurus vulpecula</i>	5,153
<i>Vombatus ursinus</i>	3,994
<i>Wallabia bicolor</i>	3,025
Monotremata	
<i>Ornithorhynchus anatinus</i>	2,389
<i>Tachyglossus aculeatus</i>	7,135
Notoryctemorphia	
<i>Notoryctes typhlops</i>	105
Peramelemorphia	
<i>Isoodon macrourus</i>	1,085
<i>Isoodon obesulus</i>	1,106
<i>Macrotis lagotis</i>	632
<i>Perameles gunnii</i>	498
<i>Perameles nasuta</i>	1,410
Rodentia	
<i>Conilurus penicillatus</i>	107
<i>Hydromys chrysogaster</i>	960
<i>Leggadina forresti</i>	392

<i>Leggadina lakedownensis</i>	157
<i>Mastacomys fuscus</i>	126
<i>Melomys burtoni</i>	592
<i>Melomys capensis</i>	21
<i>Melomys cervinipes</i>	431
<i>Mesembriomys gouldii</i>	215
<i>Mesembriomys macrurus</i>	12
<i>Notomys alexis</i>	1,020
<i>Notomys amplus</i>	14
<i>Notomys aquilo</i>	16
<i>Notomys cervinus</i>	64
<i>Notomys fuscus</i>	134
<i>Notomys mitchellii</i>	348
<i>Pseudomys apodemoides</i>	169
<i>Pseudomys australis</i>	91
<i>Pseudomys bolami</i>	262
<i>Pseudomys calabyi</i>	33
<i>Pseudomys delicatulus</i>	453
<i>Pseudomys desertor</i>	505
<i>Pseudomys fumeus</i>	54
<i>Pseudomys gracilicaudatus</i>	106
<i>Pseudomys hermannsburgensis</i>	1,858
<i>Pseudomys higginsii</i>	104
<i>Pseudomys johnsoni</i>	43
<i>Pseudomys nanus</i>	479
<i>Pseudomys novaehollandiae</i>	208
<i>Pseudomys occidentalis</i>	28
<i>Pseudomys oralis</i>	84
<i>Pseudomys pilligaensis</i>	46
<i>Pseudomys shortridgei</i>	43
<i>Rattus colletti</i>	152
<i>Rattus fuscipes</i>	2,657
<i>Rattus leucopus</i>	71
<i>Rattus lutreolus</i>	1,186
<i>Rattus sordidus</i>	57
<i>Rattus tunneyi</i>	518
<i>Rattus villosissimus</i>	308
<i>Uromys caudimaculatus</i>	131
<i>Xeromys myoides</i>	14
<i>Zyzomys argurus</i>	582
<i>Zyzomys maini</i>	35
<i>Zyzomys pedunculatus</i>	26
<i>Zyzomys woodwardi</i>	44



**Table S2.** Settings used in *biomod2*, *dismo* and Maxent.

Algorithm	Description of modelling options adopted in BIOMOD	Function arguments
Artificial neural networks (ANN)	5-fold cross-validation to find best size and decay parameters. Initial random weights = 0.1, maximum number of iterations = 200.	NbCV = 5 size = NULL decay = NULL rang = 0.1 maxit = 200
BIOCLIM	—	—
Boosted Regression Tree ( <i>biomod2</i> implementation; BRT <sub>bio</sub> )	Bernoulli error distribution, interaction depth of 7, learning rate of 0.001, bag.fraction of 0.5, 5-fold cross-validations to select optimal number of trees with a maximum of 2500 trees fitted to the data.	distribution = 'bernoulli' n.trees = 2500 interaction.depth = 7 n.minobsinnode = 5 shrinkage = 0.001 bag.fraction = 0.5 train.fraction = 1 cv.folds = 3 keep.data = FALSE verbose = FALSE perf.method = 'cv'
Boosted Regression Tree ( <i>dismo</i> implementation; BRT <sub>dis</sub> )	Bernoulli error distribution, interaction depth of 1, learning rate of 0.01, bag fraction of 0.75, 10-fold cross-validations to select optimal number of trees with a maximum of 10,000 trees fitted to the data.	offset = NULL fold.vector = NULL tree.complexity = 1 learning.rate = 0.01 bag.fraction = 0.75 site.weights = rep(1, nrow(data)) var.monotone = rep(0, length(BRT.x)) n.folds = 10 prev.stratify = TRUE family = "bernoulli" n.trees = 50 step.size = n.trees max.trees = 10000

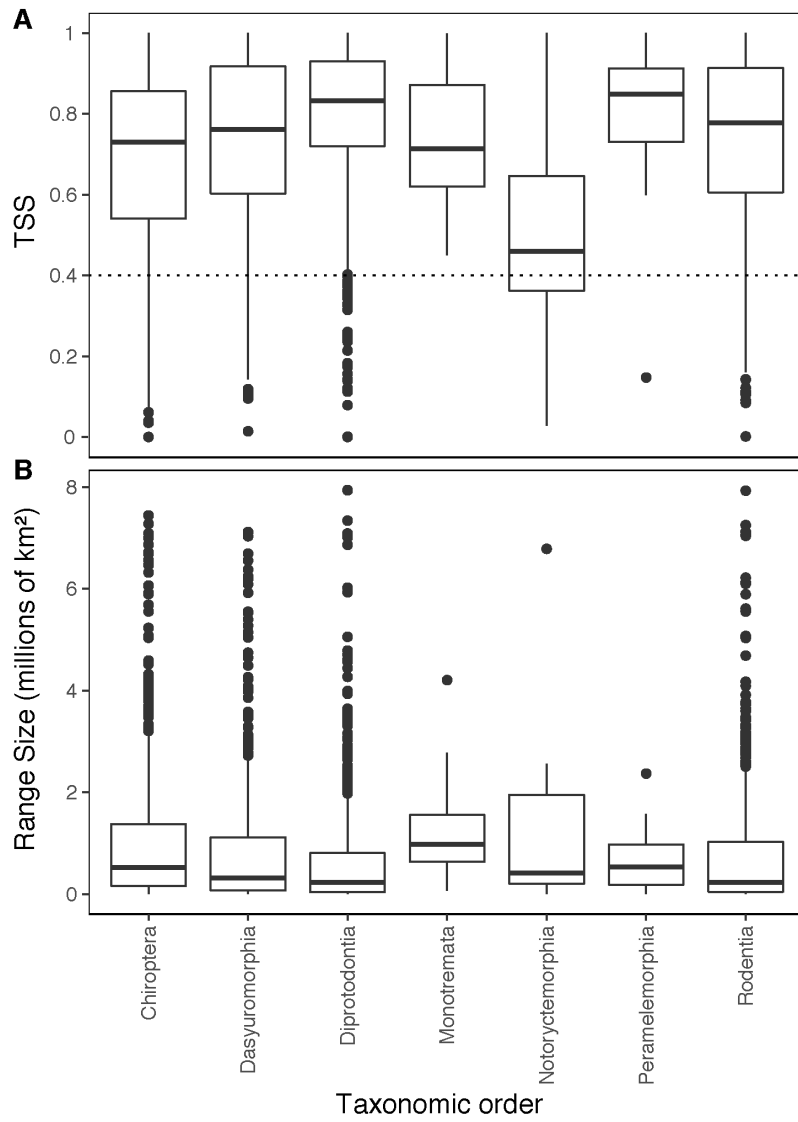
		<pre> tolerance.method = "auto" tolerance = 0.00 keep.data = FALSE plot.main = TRUE plot.folds = FALSE verbose = TRUE silent = FALSE keep.fold.models = FALSE keep.fold.vector = FALSE keep.fold.fit = FALSE </pre>
Classification tree analysis (CTA)	5-fold cross-validation.	<pre> method = 'class' parms = 'default' cost = NULL control = list(xval = 5, minbucket = 5, minsplit = 5, cp = 0.001, maxdepth = 25) </pre>
DOMAIN	—	—
Flexible discriminant analysis (FDA)	Linear regression method used for optimal scaling, via function 'polyreg'.	<pre> method = 'mars' </pre>
Generalised Additive Model (GAM)	Algorithm = 'GAM_mgcv' with cubic spline smoother, with 4 degrees of freedom. Logistic link function and a binomial error distribution of the response. Note that GVC (Generalised Cross Validation) will automatically choose the number of knots to balance simplicity against explanatory power.	<pre> algo = 'GAM_mgcv' type = 's_smoother' k = -1 interaction.level = 0 myFormula = NULL family = binomial(link = 'logit') method = 'GCV.Cp' optimizer = c('outer', 'newton') select = FALSE knots = NULL paraPen = NULL control = list(nthreads = 1, irls.reg = 0, epsilon = 1e-07, maxit = 200, trace = FALSE, mgcv.tol = 1e-07, mgcv.half = 15, rank.tol = 1.49011611938477e-08, nlm = list(ndigit=7, gradtol=1e-06, stepmax=2, steptol=1e-04, iterlim=200, check.analyticals=0), optim = list(factr=1e+07), newton = list(conv.tol=1e-06, maxNstep=5, maxSstep=2, maxHalf=30, </pre>

		<code>use.svd=0), outerPIsteps = 0, idLinksBases = TRUE, scalePenalty = TRUE, keepData = FALSE, scale.est = fletcher)</code>
Generalised Linear Model (GLM)	Logistic link function and a binomial error distribution of the response. Linear and quadratic terms with no interaction level. Stepwise procedure using the AIC criterion to select the most significant terms.	<code>type = 'quadratic' interaction.level = 0 myFormula = NULL test = 'AIC' family = binomial(link = 'logit') mustart = 0.5 control = glm.control(epsilon = 1e-08, maxit = 50, trace = FALSE)</code>
Mahalanobis (Mahal)	—	—
Multivariate Adaptive Regression Spline (MARS)	Two-way interactions, cost per degree of freedom change = 2, forward stepwise stopping threshold = 0.001.	<code>degree = 2 nk = NULL penalty = 2 thresh = 0.001 prune = TRUE</code>
Maxent	Linear, Product, Quadratic and Hinge features used; 10-fold cross-validations.	<code>cat('java -mx2048m -jar ,maxent.jar, ' -e ',bkgd.data, ' -s ',occur.data, ' -o ,maxent.dir, ' nothreshold nowarnings novisible nowriteclampgrid nowritemess writeplotdata -P -J -r -a \n', sep=" ", file=shell.file))</code>
Random Forest (RF)	Classification RF, number of predictors to be chosen randomly at each node = $\sqrt{p}$ where $p$ is number of variables. Number of trees grown = 500.	<code>do.classif = TRUE ntree = 500 mtry = 'default' nodesize = 5 maxnodes = NULL</code>
Surface Range Envelope (SRE)	Quantile of extreme environmental variable to be removed from envelope.	<code>quant = 0.025</code>



SDM	No. of species	Expected frequency	a	b	c	d	e	a	b	c
ANN	178	0.066	4 (9.5)	21 (38)	15 (4.2)	10 (15.3)	13 (24.7)	1 (5.3)	19 (21)	7 (3.6)
BIOCLIM	209	0.078	17 (11.2)	61 (44.6)	0 (5)	23 (17.9)	0 (29)	5 (6.2)	22 (24.7)	0 (4.2)
BRT <sub>bio</sub>	219	0.081	10 (11.7)	53 (46.7)	6 (5.2)	28 (18.8)	64 (30.4)	3 (6.5)	43 (25.9)	4 (4.4)
BRT <sub>dis</sub>	214	0.079	3 (11.4)	26 (45.7)	4 (5.1)	8 (18.3)	34 (29.7)	1 (6.4)	11 (25.3)	2 (4.3)
CTA	63	0.023	6 (3.4)	26 (13.4)	1 (1.5)	20 (5.4)	9 (8.7)	2 (1.9)	21 (7.4)	2 (1.3)
Domain	208	0.077	10 (11.1)	37 (44.4)	0 (4.9)	15 (17.8)	0 (28.9)	3 (6.2)	9 (24.6)	2 (4.2)
FDA	186	0.069	0 (9.9)	37 (39.7)	6 (4.4)	17 (15.9)	38 (25.8)	1 (5.5)	18 (22)	10 (3.7)
GAM	215	0.080	4 (11.5)	42 (45.9)	3 (5.1)	11 (18.4)	50 (29.8)	4 (6.4)	18 (25.4)	2 (4.3)
GLM	201	0.075	4 (10.7)	27 (42.9)	12 (4.8)	12 (17.2)	29 (27.9)	3 (6)	12 (23.7)	11 (4)
Mahal	152	0.056	1 (8.1)	30 (32.4)	4 (3.6)	5 (13)	0 (21.1)	0 (4.5)	4 (17.9)	5 (3)
MARS	205	0.076	1 (11)	18 (43.8)	0 (4.9)	3 (17.6)	47 (28.5)	0 (6.1)	6 (24.2)	1 (4.1)
Maxent	219	0.081	3 (11.7)	22 (46.7)	12 (5.2)	3 (18.8)	27 (30.4)	2 (6.5)	7 (25.9)	8 (4.4)
RF	220	0.082	54 (11.8)	102 (47)	1 (5.2)	42 (18.9)	63 (30.5)	47 (6.5)	97 (26)	0 (4.4)
SRE	205	0.076	27 (11)	73 (43.8)	0 (4.9)	39 (17.6)	0 (28.5)	8 (6.1)	31 (24.2)	0 (4.1)
<b>Sum</b>	<b>2,694</b>		<b>144</b>	<b>575</b>	<b>64</b>	<b>231</b>	<b>374</b>	<b>80</b>	<b>318</b>	<b>54</b>

**Figure S1.** Box-and-whisker plot of AUC and size of current habitat projected for 220 Australian mammal species belonging to seven taxonomic orders. Projections were made using 14 SDM methods. The dotted line indicates the TSS threshold (0.4) below which models were excluded due to poor predictive performance.



**Figure S2.**

