

This file is part of the following work:

Munroe, Samantha Elizabeth Mitchell (2015) *Habitat and dietary selectivity of nearshore shark populations*. PhD Thesis, James Cook University.

Access to this file is available from:

<https://doi.org/10.25903/rscz%2D4q09>

Copyright © 2015 Samantha Elizabeth Mitchell Munroe

The author has certified to JCU that they have made a reasonable effort to gain permission and acknowledge the owners of any third party copyright material included in this document. If you believe that this is not the case, please email

researchonline@jcu.edu.au

Habitat and Dietary Selectivity of Nearshore Shark Populations

Thesis submitted by
Samantha Elizabeth Mitchell Munroe
BSc Honours (Acadia University)

For the degree of
Doctor of Philosophy
Centre for Sustainable Tropical Fisheries and Aquaculture
College of Marine and Environmental Science
James Cook University
Townsville, Queensland
January 2015

Statement of the Contribution of Others

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34

Financial Support

- James Cook University international postgraduate scholarship co-funded the with AIMS@JCU program
- School of Earth and Environmental Sciences, James Cook University
- National Environmental Research Program
- Great Barrier Reef Marine Park Authority
- Linnaean Society of New South Wales

In-Kind Support

- Centre for Sustainable Tropical Fisheries and Aquaculture, School of Earth of Environmental Sciences, James Cook University
- GLIER Laboratory, University of Windsor

Statistical and Analytical Support

- Prof. Colin Simpfendorfer
- Dr. Michelle Heupel
- Prof. Aaron Fisk
- Dr. Murray Logan
- Dr. Jenny Elliman

Supervision

- Prof. Colin Simpfendorfer
- Dr. Michelle Heupel
- Dr. James Moloney

Editorial Support

- Prof. Colin Simpfendorfer
- Dr. Michelle Heupel
- Dr. James Moloney
- Prof. Aaron Fisk

Acknowledgements

35

36

37 I would like to thank the staff and students at James Cook University, the Centre for
38 Sustainable Tropical Fisheries and Aquaculture, and the College of Marine and Environmental
39 Science. Thank you to Colin, Michelle, and James for giving me the chance to see a new part
40 of the world and follow my dreams. My life has been forever changed by the opportunities you
41 have given me. Your expertise, advice, PATIENCE, and empathy have made all of this
42 possible. I truly appreciate your doors always being open. Thank you for helping me "turn the
43 corner" to become the researcher I am today. Thanks to Glenn, Bec, Jo, Beth, and Melissa for
44 helping me navigate the choppy waters of university paper work. To my fellow students, I am
45 forever in your debt. Your creative ideas helped me solve my most confounding problems.
46 More importantly, your friendship and company on and off the water have helped make this
47 experience a happy and memorable one. I would like to acknowledge Andrew Chin, Audrey
48 Schlaff, Fernanda de Faria, and Steve Moore, who were my earliest teachers. Thank you to
49 Leanne Currey for tolerating such a chatty officemate and helping me to see the bright side of
50 every situation. Thanks to Vinay Udyawer, your programming skills and generosity were
51 invaluable. Thank you to Mario Espinoza for his great sense of humour and encouragement.
52 And of course, Peter Yates, for letting me tag along on his research trips and always supplying
53 ample food for the long days in the field. I want to acknowledge my amazing volunteers,
54 including Madi Green, Lauren Meyer, and David Moreno. I could always count on you to get
55 up early, stay out late, and go home exhausted, all in the name of science!

56 I also want to thank the staff and students at the Great Lakes Institute for Environmental
57 Research including Aaron Fisk, Anna and Nigel Hussey, Brittney Charron and Steve Kessel. I
58 greatly appreciate all your advice and the warm welcome I received at your lab. I would like to
59 thank Jenny Elliman at the JCU School of Veterinary and Biomedical Sciences for assistance

60 in preparing tissues for isotope analysis. I also want to extend my gratitude to Murray Logan at
61 AIMS for all his statistical advice.

62 I would like to acknowledge the financial support from the Australian Government's National
63 Environmental Research Program, James Cook University, AIMS@JCU, the Great Barrier
64 Reef Marine Park Authority, the School of Earth and Environmental Science, and the Linnaean
65 Society of New South Wales. Without their support, my work would not have been possible.

66 To Ceccha, Kate, and Sarah, I know I have not been around very much, but despite the
67 distance, there hasn't been a day where I didn't know I had three lifelong friends I could always
68 count on. I want to give a great big hug to Chloe and Liz, who were more than roommates, they
69 were my guides. Thank you for making a stranger feel so welcome in a strange land. To Elle,
70 Chris, the Phils, and Tuna, thanks for welcoming me into the fold. And of course, James White,
71 what can I say except you would be on my zombie apocalypse team any day. I know I can
72 always count on you to do what is necessary.

73 To my astounding and caring family, there are no words to describe the depth of my gratitude
74 for all of your love and support. To my beautiful mother Patti, thank you for giving me such an
75 amazing example of what it is to be a strong woman and always supporting my dreams in any
76 way you could. Thanks to my dad Craig, for always believing in my abilities and being brave
77 enough to try on a wetsuit! To Rene, thank you for completing our family, I cannot imagine a
78 home without you. Jack and Ben, no matter where I am a piece of my heart is always with you.
79 To Zoey, I want you to know I really appreciated all the unconditional love, as long I fed you
80 treats, played with you, and was available for cuddles at 2:00 AM. Finally, to my indestructible
81 Jon, thank you for your patience, support, cooking, bar-tending, and friendship throughout this
82 process. After nearly four years of blood, sweat, tears, and analysis, all I truly know is I would
83 not have made it without you.

84
85
86
87
88
89
90
91
92
93
94
95
96
97
98
99
100

List of Publications from this Research

Munroe, S.E.M., Simpfendorfer, C.A., and Heupel, M.R. (2013). Defining shark ecological specialisation: concepts, context, and examples. *Reviews in Fish Biology and Fisheries* **24**, 317-331. DOI 10.1007/s11160-013-9333-7.

Munroe, S.E.M., Simpfendorfer, C.A., and Heupel, M.R. (2014). Habitat and space use of an abundant nearshore shark, *Rhizoprionodon taylori*. *Marine & Freshwater Research*. DOI <http://dx.doi.org/10.1071/MF13272>.

Munroe S.E.M., Heupel, M.R., Fisk, A.T., and Simpfendorfer, C.A. (accepted). Geographic and temporal variation in the trophic ecology of a small bodied-shark: Evidence of resilience to environmental change. *Canadian Journal of Fisheries and Aquatic Sciences*.

Munroe, S.E.M., Heupel, M.R., Fisk, A.T., Logan, M., and Simpfendorfer, C.A. (accepted). Regional movement patterns of a small-bodied shark revealed by stable isotope analysis. *Journal of Fish Biology*.

Munroe, S.E.M., Simpfendorfer, C.A., Moloney, J. and Heupel, M.R. (accepted). Nearshore movement ecology of a medium-bodied shark, the creek whaler *Carcharhinus fitzroyensis*.

Conference and Meeting Presentations

101

102

103 Munroe, S.E.M., Simpfendorfer, C.A., and Heupel M.R. (2013) Space use and habitat selection
104 of the Australian sharpnose shark *Rhizoprionodon taylori* in a nearshore environment. Oral
105 presentation. AIMS @JCU Student Conference, Townsville, Queensland (2nd best student oral
106 presentation winner).

107

108 Munroe, S.E.M., Simpfendorfer, C.A., and Heupel M.R. (2013) Space use and habitat selection
109 of the Australian sharpnose shark *Rhizoprionodon taylori* in a nearshore environment. Oral
110 presentation. School of Earth and Environmental Science Conference, Townsville, Queensland.

111

112 Munroe, S.E.M., Simpfendorfer, C.A., and Heupel M.R. (2013). Space use and habitat selection
113 of the Australian sharpnose shark *Rhizoprionodon taylori* in a nearshore environment. Oral
114 presentation. Oceania Chondrichthyan Society Conference. Brisbane, Queensland (Best student
115 oral presentation winner).

116

117 Munroe, S.E.M., Simpfendorfer, C.A., and Heupel M.R. (2013). Shark habitat selection in
118 nearshore ecosystems. Three Minute Thesis Conference. James Cook University, Townsville,
119 Queensland (School and Department presentation winner).

120

121 Munroe, S.E.M., Heupel, M.R., Fisk, A.T., Logan, M., and Simpfendorfer, C.A. (2014). Coastal
122 habitat use of an abundant mesopredator revealed through telemetry and stable isotope analysis.
123 Oral presentation. Sharks International. Durban, South Africa.

124

125 Munroe, S.E.M., Heupel, M.R., Fisk, A.T., Logan, M., and Simpfendorfer, C.A. (2014). Stable
126 isotope analysis reveals dietary selection patterns of a nearshore shark population. Poster
127 presentation, Sharks International. Durban, South Africa.

128

129 Munroe, S.E.M., Heupel, M.R., Fisk, A.T., Logan, M., and Simpfendorfer, C.A. (2014). You are
130 where you eat! Regional residency patterns of a small-bodied shark revealed by stable isotope
131 analysis. Oral presentation AIMS @JCU Student Conference, Townsville, Queensland.

132

133 Munroe, S.E.M., Heupel, M.R., Fisk, A.T., Logan, M., and Simpfendorfer, C.A. (2014). You are
134 where you eat! Regional residency patterns of a small-bodied shark revealed by stable isotope
135 analysis . Oral presentation. School of Earth and Environmental Science Conference,
136 Townsville, Queensland (Best student oral presentation winner).

137

138

139

140

Abstract

141
142
143 The resource use strategies species exhibit affects their role within communities and how
144 they respond to environmental change. Species that adopt generalist strategies are typically
145 less vulnerable to environmental fluctuations than specialists. However, specialists often
146 have lower energy costs and are more efficient at extracting and processing preferred
147 resources. This dissertation defines shark ecological specialisation in order to evaluate the
148 resource use patterns of nearshore sharks and discusses how shark resource use patterns can
149 affect their vulnerability to environmental change.

150
151 Sharks are traditionally classified as generalists that use a variety of habitats and prey.
152 While this is an accurate description of some species, sharks exhibit a range of resource use
153 strategies that includes highly selective or specialised behaviours. However, discussion on
154 how to define the ecological specialisation of sharks has been limited. This dissertation
155 presents a conceptual framework within which to define the specialisation of sharks that
156 can be applied to different environmental scales. Shark species with varying degrees of
157 specialisation are presented within the proposed context.

158
159 Passive acoustic telemetry was used to examine the residency, space use, and habitat
160 selection and specialisation patterns of the small-bodied Australian sharpnose shark,
161 *Rhizoprionodon taylori*, and the medium-bodied creek whaler *Carcharhinus fitzroyensis*, in
162 Cleveland Bay, Queensland, Australia. Stable isotope analysis of $\delta^{13}\text{C}$ ($^{13}\text{C}/^{12}\text{C}$) and $\delta^{15}\text{N}$
163 ($^{15}\text{N}/^{14}\text{N}$) was used to define the regional nearshore residency, movements, trophic level,
164 and benthic and pelagic contributions to the diet of *R. taylori*. $\delta^{13}\text{C}$ values vary at the base

165 of the food chain but are conserved up the food chain. $\delta^{15}\text{N}$ values increase in predictable
166 quantities between trophic levels. Therefore, $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values can be used to indicate
167 the foraging location and diet of populations. Plasma and muscle $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of *R.*
168 *taylori* were collected from five embayments, including Cleveland Bay, on the northeast
169 coast of Queensland.

170

171 Results of the acoustic tracking showed the majority of *R. taylori* were present in Cleveland
172 Bay for short periods of time, ranging from 1 to 112 days (mean \pm SE = 17 ± 5). The
173 majority of individuals were present in Cleveland Bay for less than two weeks. Low
174 residency suggests that *R. taylori* home ranges likely span multiple bays. Individual
175 monthly activity space ranged from 4.3 and 21.4 km² (mean \pm SE = $11.3 \text{ km}^2 \pm 0.90$) for
176 50% kernel utilisation distributions (KUDs) and 21.5 and 80.4 km² (mean \pm SE = 51.0 km^2
177 ± 3.9) for 95% KUDs. Space use analysis indicated *R. taylori* roamed widely throughout
178 the bay, but monthly activity space size was consistent among individuals and over time.
179 Sex and size had no influence on *R. taylori* residency or activity space size. Both the
180 population and individuals occupied wide habitat niches which included seagrass, outer bay
181 mud substrate, and sandy inshore habitat. However, both resident and transitory *R. taylori*
182 consistently selected for seagrass over other habitats, potentially for feeding. Mudflat and
183 reef habitats were generally avoided. Habitat selection appeared to be influenced by
184 changes in freshwater input into Cleveland Bay. Selection for seagrass habitat, which is
185 adjacent to large river mouths, decreased during periods of high river flow, suggesting *R.*
186 *taylori* may have limited tolerance to low salinity.

187

188 Stable isotope analysis showed that there was a positive correlation between *R. taylori*
189 tissue and environmental (seagrass and plankton) $\delta^{13}\text{C}$ values based on location. Shark
190 populations with the highest tissue $\delta^{15}\text{N}$ were collected from areas with the highest baseline
191 $\delta^{15}\text{N}$ values. Moreover, populations from bays that were > 100 km apart had distinct
192 isotopic values. These results indicate *R. taylori* were not foraging more than 100 km from
193 their capture location within 6 to 12 months. However, $\delta^{13}\text{C}$ values of individuals in nearby
194 bays (30-70 km apart) were indistinguishable, suggesting individuals foraged and moved
195 between bays that were within 100 km of each other during a 6 to 12 month period.
196 Therefore, isotope results were consistent with the low residency exhibited by *R. taylori* in
197 Cleveland Bay. Isotope analysis also revealed *R. taylori* had a wide trophic range and
198 consumed prey from benthic and pelagic sources. In all areas, benthic sources were
199 important to the diet, suggesting benthic habitats (e.g. seagrass) may be important to *R.*
200 *taylori*. However, there was geographic and temporal variation in *R. taylori* diet. These
201 results indicate *R. taylori* has a broad dietary niche, but different populations may have
202 unique effects on distinct areas. Variation in diet also suggests *R. taylori* may be adaptive
203 to changes in prey availability.

204

205 In contrast to *R. taylori*, most *C. fitzroyensis* were highly resident and present in Cleveland
206 Bay for long periods of time, ranging 1 to 452 days (mean \pm SE = 205 \pm 53). However, a
207 few individuals spent less than two weeks in the bay, suggesting broader movements occur
208 in a portion of the population. Size and sex had no effect on presence. Individual monthly
209 activity space ranged from 2.6 to 19.8 km² (mean \pm SE = 10.6 km² \pm 0.3) for 50% KUDs
210 and 9.1 to 81.9 km² (mean \pm SE = 47.9 km² \pm 1.0) for 95% KUDs. Activity space size
211 varied between months and diel period but was not affected by animal size. Activity spaces

212 in August were significantly smaller and concentrated closer to the shore than in other
213 months. This simultaneous shift in space use by all individuals may have been due to biotic
214 changes in the bay, such as changes in prey availability. Larger day time activity spaces
215 suggest *C. fitzroyensis* may be primarily diurnal feeders. All resident *C. fitzroyensis* spent
216 the majority of time in seagrass and to a lesser extent outer bay mud substrate habitat.
217 Seagrass was consistently selected for throughout the monitoring period while use of outer
218 bay mud substrate was highly irregular. Shallow mudflat, sandy inshore, and reef habitats
219 were rarely used. There was no difference in space or habitat use between immature and
220 mature individuals, indicating different age classes shared space and habitats.

221
222 The results of acoustic and isotope analyses indicated that seagrass habitat is the preferred
223 habitat of *R. taylori* and *C. fitzroyensis*. Seagrass is typically highly productive and may be
224 an important foraging habitat for these species. For that reason, seagrass conservation will
225 be an important consideration for the future spatial management of these species. However,
226 *R. taylori* used different habitats and embayments and had a broad diet. Therefore, results
227 indicate *R. taylori* has a low degree of resource specialisation and is probably adaptive to
228 local environmental change. In contrast, the movement patterns exhibited by *C. fitzroyensis*
229 suggest this species has a moderately high degree of habitat specialisation and is highly
230 resident. Although diet information is not available for *C. fitzroyensis*, the movement
231 patterns of *C. fitzroyensis* indicate this species will likely be more vulnerable to local
232 environmental change, specifically a decline in seagrass abundance. By evaluating the
233 resource use patterns of nearshore sharks, this dissertation has provided valuable
234 information on the potential vulnerabilities of poorly understood shark species while also
235 developing a conceptual framework for future resource specialisation investigations.

Table of Contents

236		
237	Chapter 1	
238	General Introduction.....	1
239		
240	Chapter 2	
241	Defining Shark Ecological Specialisation: Concepts, Context, and Examples	6
242	2.1 Introduction.....	6
243	2.2 Defining Shark Specialisation: Terminology and the Continuum Concept.....	8
244	2.3 Defining Shark Specialisation: The Ecological Niche of a Shark	10
245	2.4 Shark Habitat Specialisation	16
246	2.5 Shark Dietary Specialisation.....	21
247	2.6 Implications of Resource Use Strategies	26
248	2.7 Conclusion	28
249		
250	Chapter 3	
251	General Methods.....	30
252	3.1 Acoustic Analysis	30
253	3.1.1 Study Site	30
254	3.1.2 Study Species	32
255	3.1.3 Field Methods.....	36
256	3.2 Stable Isotope Analysis.....	39
257	3.2.1 Study Site	39
258	3.2.2 Study Species	41
259	3.2.3 Field Methods.....	41
260	3.2.4 Sample Preparation and Isotope Analysis	44
261	3.3 Permits and Ethics.....	45
262		
263	Chapter 4	
264	Habitat and Space Use of an Abundant Nearshore Shark, <i>Rhizoprionodon taylori</i>	46
265	4.1 Introduction.....	46
266	4.2 Data Analysis	47
267	4.2.1 Residency	47
268	4.2.2 Space Use	48

269	4.2.3 Habitat Selection	49
270	4.2.4 Habitat Niche Breadth	50
271	4.3 Results.....	51
272	4.3.1 Residency	52
273	4.3.2 Space Use	54
274	4.3.3 Habitat Selection and Niche Breadth	56
275	4.4 Discussion	63
276		
277	Chapter 5	
278	Regional Movement Patterns of a Small-bodied Shark Revealed by Stable Isotope Analysis ..	69
279	5.1 Introduction.....	69
280	5.2 Data Analysis	72
281	5.3 Results.....	76
282	5.3.1 Study Site $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$	76
283	5.3.2 Shark tissue $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$	80
284	5.4 Discussion	86
285		
286	Chapter 6	
287	Geographic and Temporal Variation in the Trophic Ecology of a Small-bodied Shark	92
288	6.1 Introduction.....	92
289	6.2 Data Analysis	95
290	6.3 Results.....	98
291	6.3.1 Study Site $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$	98
292	6.3.2 Shark $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$	100
293	Discussion	106
294		
295	Chapter 7	
296	Movement Ecology of a Nearshore, Medium-bodied Shark, the Creek Whaler <i>Carcharhinus</i>	
297	<i>fitzroyensis</i>	112
298	7.1 Introduction.....	112
299	7.2 Data Analysis	114
300	7.2.1 Residency	114
301	7.2.2. Space use	114
302	7.2.3 Habitat selection	115

303	7.2.4 Habitat niche breadth	116
304	7.3 Results.....	117
305	7.3.1 Residency	117
306	7.3.2 Space Use	119
307	7.3.3 Habitat Selection and Specialisation	124
308	7.4 Discussion	128
309		
310	Chapter 8	
311	General Discussion	133
312	8.1 Conclusions and Implications	133
313	8.2 Future Research	137
314		
315	References	140
316		
317		

318

319

List of Tables

320	Table 4.1. Strauss selectivity index values of low residency <i>Rhizoprionodon taylori</i> (< 2	
321	weeks detected), for each primary habitat in the bay, indicating sex as M (male) or F (female)	
322	and size as stretch total length in mm for each individual.....	57
323		
324	Table 4.2. Mean and standard deviation of of time spent in each habitat by high residency (> 2	
325	weeks) <i>Rhizoprionodon taylori</i> (n=7) released in year two (September 2012 to April 2013) in	
326	Cleveland Bay, measured as a percent with standard error (SE).....	59
327		
328	Table 5.1 Sample size (n) and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ range (mean \pm SD) of benthic and pelagic	
329	sources from each location, Repulse Bay (R), Upstart Bay (UP), Bowling Green Bay (BG),	
330	Cleveland Bay (CB), and Rockingham Bay (RO).....	77
331		
332	Table 5.2. <i>Rhizoprionodon taylori</i> male (M) and female (F) sample size, combined stretch total	
333	length in mm (STL mm), and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ tissue range (mean \pm SD) from Repulse Bay (R),	
334	Upstart Bay (UP), Bowling Green Bay (BG), Cleveland Bay (CB), and Rockingham Bay (RO).	
335	81
336		
337	Table 5.3. Results of linear Bayesian correlation analysis between the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of	
338	<i>Rhizoprionodon taylori</i> and the benthic and pelagic producers across sample bays. ρ defined	
339	the correlation parameter. Correlations were considered significant if the 95% credibility	
340	intervals (CIs) of the posterior draws did not overlap with 0.	85
341		
342	Table 6.1. The $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ range (mean \pm SD) of combined pelagic and benthic samples	
343	from each sample area, Repulse Bay (RE), Cleveland Bay Unit (CBU), and Rockingham Bay	
344	(RO).	99
345		
346	Table 6. 2. Total catch, $\delta^{13}\text{C}$ range, $\delta^{15}\text{N}$ range, trophic position (TP) range and mean with	
347	standard error (SE) of female <i>Rhizoprionodon taylori</i> in each sample area (Area), Repulse Bay	
348	(RE), Cleveland Bay Unit (CBU), and Rockingham Bay (RO).....	102
349		
350	Table 7.1. Effect of animal size, month and diel period on creek whaler <i>Carcharhinus</i>	
351	<i>fitzroyensis</i> 50% and 95% kernel utilisation distribution (KUD) size (km ²). Degrees of	
352	Freedom (df), Akaike's information criterion correction (AICc), ΔAICc , and Akaike weights	
353	(W) values are given for each model. AICc values are marked with an * if models were	
354	significantly different from the null model using a likelihood ratio test. The best fit models are	
355	bolded	120
356		
357	Table 7.2. Mean and standard error of time spent in each habitat by mature and immature	
358	<i>Carcharhinus fitzroyensis</i> in Cleveland Bay, measured as a percent (%) with standard error	
359	(SE).....	125
360		

361

List of Figures

362	
363	
364	Fig. 2.1. A hypothetical example of Hutchinson’s niche space with two commonly examined
365	shark habitat use variables, depth (y) and temperature (x). x1 and x2 denote a species’
366	temperature range, y1 and y2 denote a species’ depth range. The dark grey space created by
367	these overlapping ranges is the niche space of an organism, any point within which it can
368	survive. Any point outside it is not considered a part of the species niche. 13
369	
370	Fig. 2.2. (a) Hypothetical depiction of Hutchinson’s (1957) niche hyper-volume of a species
371	with a large niche breadth, indicating a lesser degree of specialisation (b); Hypothetical
372	depiction of Hutchinson’s (1957) niche hyper-volume of a species with a narrow niche breadth,
373	indicating a greater degree of specialisation..... 14
374	
375	Fig. 3.1. Cleveland Bay, Queensland, Australia, locations of receivers in intertidal mudflat (○),
376	seagrass (■), outer bay mud substrate (●), inshore sand (Δ), and reef habitat (▲)..... 31
377	
378	Fig. 3.2. a) <i>Rhizoprionodon taylori</i> from Last and Stevens (2009) and b) photo of <i>R. taylori</i>
379	(photo credit Centre for sustainable) 33
380	
381	Fig. 3.3. a) <i>Carcharhinus fitzroyensis</i> from Last and Stevens (2009) b) photo of <i>C. fitzroyensis</i>
382	(photo credit Vinay Uduywer)..... 34
383	
384	Fig. 3.4. a) V13 and b) V16 acoustic transmitters (photo credit Vemco Ltd., Canada)..... 37
385	
386	Fig. 3.5. Surgery and processing procedures for sharks, a) surgical implantation of V13
387	acoustic transmitter b) measuring and tagging of a captured shark 38
388	
389	Fig. 3.6. Sampling region for <i>Rhizoprionodon taylori</i> indicating the five sample bays. Inset
390	indicates location along the north Queensland coast, Australia. 40
391	
392	Fig. 3.7. a) Blood extraction from <i>Rhizoprionodon taylori</i> and b) onboard centrifuge
393	equipment. 42
394	
395	Fig. 4.1 Daily presence of <i>Rhizoprionodon taylori</i> released with acoustic transmitters in
396	Cleveland Bay in 2011- 2013. Individuals are identified by sex and stretch total length (mm).53
397	
398	Fig. 4.2. <i>Rhizoprionodon taylori</i> monthly activity spaces of three individuals (a, b, c) in
399	December 2012, January 2013 and February 2013. Each panel shows the 95% (blue fill) and
400	50% (yellow fill) kernel utilisation distributions..... Error! Bookmark not defined.
401	
402	Fig. 4.3 Strauss linear selection index values of resident (> 2 weeks) <i>Rhizoprionodon taylori</i>
403	released in Cleveland Bay in year one (September 2011 to September 2012) a) female 574 mm
404	stretch total length (STL) and b) female 713 mm STL. 58
405	
406	Fig. 4.4. Mean Strauss linear selection index values of resident (> 2 weeks) <i>Rhizoprionodon</i>
407	<i>taylori</i> in Cleveland Bay ($n = 7$) between September 2012 to April 2013. Bars indicate standard
408	error. 61
409	

410 **Fig. 4.5.** *Rhizoprionodon taylori* mean fortnightly individual Strauss linear selection values for
411 resident individuals from year two for seagrass (green line), sand (yellow line), outer bay mud
412 substrate (blue line), intertidal mudflat (brown line), and reef (pink line). Mean fortnightly river
413 discharge was measured in m^3s^{-1} (thin black dashed line).62
414

415 **Fig. 5.1.** Median $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values from Bayesian ANOVA of (a) benthic (white), pelagic
416 (dark grey), (b) *Rhizoprionodon taylori* muscle (red), and plasma (green) samples in Repulse
417 Bay (●), Upstart Bay (■), Bowling Green Bay (◆), Cleveland Bay (▲) and Rockingham Bay
418 (▼), black lines show 50% and 95% credibility intervals of posterior draws78
419

420 **Fig. 5.2.** Median $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of Bayesian ANOVA of female (white) and male
421 (dark grey) *Rhizoprionodon taylori* for muscle (a) and plasma(b) tissue in Bowling Green Bay
422 (●), Cleveland Bay (▲) and Rockingham Bay (■). Black lines show 50% and 95% credibility
423 intervals of posterior draws83
424

425 **Fig. 6.1.** Map of stable isotope sampling region indicating the five sampling locations and three
426 designated feeding areas, Rockingham Bay, Cleveland Bay Unit (CBU), and Repulse Bay for
427 *Rhizoprionodon taylori*. Inset indicates location along the north Queensland coast, Australia.. 96
428

429 **Fig. 6.2.** (a) Median $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ results of Bayesian ANOVA of benthic (black) and pelagic
430 baselines (white) in Repulse Bay (■), the Cleveland Bay Unit (●) and Rockingham Bay (▲);
431 (b) median $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ results of Bayesian ANOVA of *Rhizoprionodon taylori* for muscle
432 (red) and plasma (green) in Repulse Bay (■), the Cleveland Bay Unit (●) and Rockingham Bay
433 (▲). Black lines show 95% credibility intervals of posterior draws..... 101
434

435 **Fig. 6.3.** Isotopic niche breadth of *Rhizoprionodon taylori*. Convex hulls of total niche width of
436 muscle (a) and plasma (b) are dotted lines. Bayesian Standard Ellipses (SEA_b) isotope niches
437 are shown for Repulse Bay (RE; ■/black), Cleveland Bay Unit (CBU; ●/red), and Rockingham
438 Bay (RO; ▲/green). SEA_b area calculations are also given as 50, 75, 95 credibility intervals
439 (dark to light grey) of posterior draws for muscle (c) and plasma (d), black dots indicate
440 median values. 103
441

442 **Fig. 6.4.** Proportional contributions of benthic and pelagic food web sources to *Rhizoprionodon*
443 *taylori* diet using a two-source Bayesian mixing model for plasma and muscle tissue in a)
444 Repulse Bay, b) Cleveland Bay Unit, and c) Rockingham Bay. Shaded boxes are 50, 75, 95
445 (from dark to light grey) credibility intervals of posterior draws of SEA_b 105
446

447 **Fig. 7.1.** Daily presence of *Carcharhinus fitzroyensis* released with acoustic transmitters in
448 Cleveland Bay in 2012- 2014. Individuals are identified by maturity (mature=MAT,
449 immature=IMMAT) and sex (male=M, female=F). Detections in Cleveland Bay are indicated
450 by black circles. Additional detections in Bowling Green Bay are indicated by white triangles
451 118
452

453 **Fig.7.2.** Effect of Diel period on (a) 50% and (b) 95% kernel utilisation size (km^2) of
454 *Carcharhinus fitzroyensis*. Blue bars are mean predicted value of linear mixed effect model
455 output, grey bands are 95% confidence intervals of model output 121
456

457 **Fig.7.3.** Effect of Month on 50% (a) and 95% (b) kernel utilisation size (km^2) of *Carcharhinus*
458 *fitzroyensis*. Blue bars are mean predicted value of linear mixed effect model output, grey
459 bands are 95% confidence intervals of model output..... 122

460		
461	Fig.7.4. Monthly activity spaces of (a) one immature and (b, c) two mature <i>Carcharhinus</i>	
462	<i>fitzroyensis</i> in Cleveland Bay in May 2013, August 2013 and December 2013. Each panel	
463	shows the 95% (blue fill) and 50% (yellow fill) kernel utilisation distributions.	123
464		
465	Fig.7.5. Mean Strauss linear selection index values of potential seagrass (green), outer bay mud	
466	substrate (blue), reef (red), sandy inshore (yellow) and intertidal mudflat (black) habitats by	
467	<i>Carcharhinus fitzroyensis</i> in Cleveland Bay between September 2012 to May 2014. Bars	
468	indicate standard error.	126
469		
470	Fig. 7.6. Mean fortnightly individual Strauss linear selection values of <i>Carcharhinus</i>	
471	<i>fitzroyensis</i> in Cleveland Bay for seagrass (green), outer bay mud substrate (blue), reef (red),	
472	sandy inshore (yellow) and intertidal mudflat (black).....	127
473		

Chapter 1

General Introduction

474

475

476

477 Tropical nearshore areas are highly productive environments that contain a diverse range of
478 habitats including mangroves, rivers, and reefs (Nixon *et al.* 1986; Spalding *et al.* 2007). In
479 addition to diverse habitats, nearshore areas often experience large-scale changes in
480 environmental factors such as salinity, water temperature, oxygen content and nutrient
481 availability (Breitburg 1990; Clarke and Leakey 1996; Meynecke and Lee 2011). This
482 diversity and productivity sustains a high abundance of species over a broad range of taxa
483 (Beck *et al.* 2001). However, as a result of its productivity, diversity, and proximity to human
484 settlements, nearshore environments are also heavily exploited (Suchanek 1994). A high level
485 of historically unmonitored inshore fishing has resulted in the decline of numerous nearshore
486 species (Jackson *et al.* 2001). Human development in coastal areas, such as dredging and
487 construction (e.g. seawalls), often result in habitat destruction or decline (Lotze *et al.* 2006;
488 Bulleri and Chapman 2010). Pollution is also a major contributor to nearshore environmental
489 deterioration (Shahidul Islam and Tanaka 2004). Chemical contaminants such as herbicides
490 and heavy metals have been linked to nearshore habitat damage and disease in a variety of
491 marine taxa (teleosts, marine mammals, etc.) (Kennish 1998; Haynes *et al.* 2000). Therefore,
492 nearshore species may be highly susceptible to population decline as a result of decreasing
493 habitat quality due to human-induced change.

494

495 A diverse range of shark species use nearshore areas (Compagno 2001). Body types range
496 from large-bodied requiem sharks, such as the sandbar shark *Carcharhinus plumbeus*

497 (Compagno 1984), to dorsoventrally flattened species, such as the Pacific angel shark
498 *Squatina californica* (Gaida 1997). A variety of reproductive strategies, from viviparity (e.g.
499 the bonnethead shark; Parsons 1993) to oviparity (e.g. The Port Jackson shark, *Heterodontus*
500 *portusjacksoni*; McLaughlin and O'Gower 1971), are found among sharks in nearshore areas.
501 Species also exhibit a broad range of behaviours and nearshore resource use strategies. Sharks
502 have been found in a wide variety of nearshore habitats, including rivers (Heupel *et al.* 2010;
503 Pillans *et al.* 2010), mudflats and soft substrate (Espinoza *et al.* 2011), seagrass (Heithaus *et*
504 *al.* 2006), and rocky substrate or coastal reefs (Carraro and Gladstone 2006). Different
505 species also spend variable periods of time in nearshore areas. For example, leopard sharks
506 *Triakis semifasciata* were found to be highly resident in estuarine environments (Carlisle and
507 Starr 2009), while the sevengill shark *Notorynchus cepedianus* exhibited seasonal patterns of
508 presence in Pacific estuarine embayments (Williams *et al.* 2012). Other species, such as the
509 spottail shark *Carcharhinus sorrah*, have demonstrated high individual variability (i.e.
510 different patterns) in presence in nearshore areas (Knip *et al.* 2012b). Although most shark
511 species have been reported to use a wide range of nearshore habitats and prey (Hanchet 1991;
512 Gelsleichter *et al.* 1999; Carlson *et al.* 2008), some species exhibit strong preferences for
513 specific nearshore habitats, potentially due to high prey availability (Heithaus *et al.* 2002;
514 Carlisle and Starr 2009) or the protection these areas provide from predators (Branstetter
515 1990; Morrissey and Gruber 1993; Heupel and Hueter 2002). As a result, some species
516 consume a relatively narrow range of spatial and/or dietary resources in coastal areas (Cortes
517 *et al.* 1996; Simpfendorfer *et al.* 2001; Chin *et al.* 2013).

518

519 The diversity of resource use patterns observed in nearshore sharks is the result of a variety of
520 factors, including physiology (Pillans *et al.* 2005), morphology (Edmonds *et al.* 2001), and
521 resource needs (Carlisle and Starr 2010). However, the life history of a species can also have

522 a substantial influence on its presence in nearshore habitats and its resource use patterns
523 (Cortés 2000). Juveniles of large-bodied, slow growing, late maturing species, such as the
524 blacktip shark *Carcharhinus limbatus* (Heupel, *et al.* 2010) or the bull shark *Carcharhinus*
525 *leucas* (Ortega *et al.* 2009; Werry *et al.* 2011), commonly use nearshore environments as
526 nursery areas (Springer 1967; Grubbs 2010). Shallow nearshore areas can provide juvenile
527 sharks with protection from predators and function as highly productive foraging grounds
528 (Branstetter 1990; Heithaus 2007). Therefore, high residency to these areas helps to ensure
529 juveniles reach sexually maturity (Heupel *et al.* 2007). As a result, juveniles typically spend
530 long periods of time, sometimes several years, in a single nearshore embayment (Conrath and
531 Musick 2010; Knip *et al.* 2011). Large-bodied adults have significantly larger home ranges
532 that include offshore habitats (Dicken *et al.* 2008; Carlson *et al.* 2010). Consequently,
533 juvenile and adult populations are often spatially segregated (Grubbs 2010).

534

535 The nearshore movement patterns of small-bodied, highly productive, fast growing sharks,
536 such as the milk shark *Rhizoprionodon acutus* (Henderson *et al.* 2006; Schroeder 2011),
537 strongly contrast with those of large-bodied species. Small-bodied sharks are typically found
538 in nearshore areas throughout their lives, where juveniles and adults simultaneously use the
539 same nearshore habitats (Simpfendorfer and Milward 1993; Knip *et al.* 2010). There is also
540 evidence to indicate that, despite their small size, individuals regularly move between
541 different nearshore areas (Kohler *et al.* 1998; Carlson *et al.* 2008), in contrast to the juveniles
542 of large-bodied species. However, compared to large-bodied species, little information is
543 available on how small coastal species use nearshore areas.

544

545 The nearshore resource use patterns of medium-bodied species, such as the whiskery shark
546 *Furgaleus macki* (1500 mm total length), or the nervous shark *Carcharhinus cautus* (1200-
547 1400 mm total length), are also poorly understood (Last and Stevens 2009). The life history
548 traits of these species are sometimes an intermediate of those exhibited by large- and small-
549 bodied sharks (Lyle 1987; Simpfendorfer *et al.* 2000). Therefore, medium-bodied sharks may
550 demonstrate unique coastal movement patterns compared to large- and small-bodied species.

551

552 Given their variability in use of nearshore regions, sharks play a variety of ecological roles in
553 nearshore ecosystems. Sharks can control prey populations via direct predation (Stevens *et al.*
554 2000; Heithaus *et al.* 2008), and have also been shown to alter prey behaviour via risk
555 avoidance (Heithaus and Dill 2002; Heithaus *et al.* 2012). However, biological factors such as
556 morphology will influence the effect sharks have on nearshore environments. For example,
557 large-bodied species often occupy high trophic positions within nearshore food chains and
558 prey on larger coastal fauna, whereas small-bodied mesopredators exert top-down control
559 over smaller-bodied nearshore species and may also be prey for larger sharks (Cortés 1999).
560 Mobility will also affect the influence sharks have on nearshore ecosystems. Highly mobile or
561 migratory species may connect separated food webs by moving between them and consuming
562 local resources (Lundberg and Moberg 2003). Less mobile or more resident populations will
563 likely have a more localized on nearshore regions.

564

565 The behaviour and resource use patterns exhibited by nearshore sharks will also affect their
566 vulnerability to environmental change. Species that are highly mobile and use a wide range of
567 resources (i.e. habitats or prey) will be less vulnerable to environmental change than species
568 that are highly specialised and use a narrow range of resources (Futuyma and Moreno 1988;

569 Wilson *et al.* 2008). This is because highly specialised species may not be able to use new or
570 additional resources if preferred resources decline in health or availability (Colles *et al.* 2009;
571 Clavel *et al.* 2010). In contrast, during periods of environmental change or decline, species
572 that use large resource niches should be able to use supplemental resources and thus maintain
573 a relatively high level of fitness (Marvier *et al.* 2004). Therefore, defining resource use
574 patterns of sharks, most notably the degree of specialisation, is critical to understanding their
575 role in nearshore ecosystems and their vulnerability to change.

576

577 Given current knowledge gaps and research needs, the primary aims of this dissertation were
578 to: 1) develop a definition of shark resource specialisation, 2) use this definition to evaluate
579 the resource use patterns (i.e. habitat and diet) of poorly understood nearshore shark species,
580 and 3) discuss how the observed resource use patterns affect vulnerability to nearshore
581 environmental change. To accomplish these aims the ecological literature on niche theory,
582 specialisation, and shark resource use was reviewed to create a definition for shark ecological
583 specialisation (Chapter 2). The application and interpretation of this definition was explored
584 using past shark resource use studies. Acoustic telemetry (Chapters 4 and 7) and stable
585 isotope analysis (Chapter 5 and 6) were used to investigate the movement and resource use
586 patterns of a small-bodied and a medium-bodied nearshore shark species. The accumulated
587 data were then used to evaluate and compare these species' vulnerability to nearshore
588 environmental change (Chapter 8). This dissertation provides new and important information
589 on the resource use patterns of poorly understood nearshore shark species and will contribute
590 to marine coastal management as well as the study of other shark species.

591

592

Chapter 2

593

Defining Shark Ecological Specialisation: Concepts, Context, and Examples

594

595

2.1 Introduction

597

598

Sharks are found in the majority of aquatic environments and as higher trophic level

599

consumers they have direct and indirect influences on community structure and function

600

(Stevens *et al.* 2000; Heithaus *et al.* 2008). To quantify sharks relationship with the

601

environment, a growing body of literature has evaluated shark habitat use, distribution, and

602

diet (e.g. Carlisle and Starr 2009; Cabrera-Chávez-Costa *et al.* 2010; Cartamil *et al.* 2010;

603

Clarke *et al.* 2011). These data are critical to the creation of successful management solutions

604

for declining and endangered populations as well as ecosystems as a whole. Recognition of

605

differing strategies leads to a better understanding of the strengths and vulnerabilities of a

606

species in their environment and aids conservation programs (Dulvy *et al.* 2008; Chin *et al.*

607

2010; Simpfendorfer *et al.* 2011).

608

609

Two broad resource use strategies exist among organisms. There are generalists that have

610

large ecological niches and use a wide range of resources; and specialists that have relatively

611

small ecological niches and use a comparatively narrow range of resources (Futuyma and

612

Moreno 1988; Irschick *et al.* 2005). In the past sharks have been depicted as mobile

613

generalists with wide, sometimes global, ranges that feed opportunistically rather than

614

foraging for specific prey items (Wetherbee *et al.* 1990). While this is an accurate description

615

for some species, research has revealed others specialise on a narrow range of habitat and

616

prey. In reality, species exhibit strategies across the continuum between these two extremes

617 (Compagno 1990). The use of generalist or specialist strategies has distinct implications for
618 the populations that adopt them, with costs and benefits resulting from each (DeWitt 1998;
619 Richmond *et al.* 2005; Colles *et al.* 2009). Therefore, knowledge of which species uses each
620 strategy is essential to not only understanding ecological interactions but also to the
621 development of appropriate management schemes. For these reasons, as shark research
622 advances, a greater number of studies will aim to define shark resource use as selective,
623 opportunistic, specialised or generalised. The application of ecological theory to analyse
624 variability in resource use is also likely to increase.

625

626 Unfortunately, similar to other fields of ecological research, definitions for what is meant by
627 “selection”, “specialist” or “generalist” are rarely stated in literature. Lack of definition has
628 led to confusion in shark ecology over the appropriate application of these important
629 concepts. As a consequence of this confusion, specialisation is either poorly articulated in
630 publications or inferred using inappropriate methodologies. Incorrect classifications inhibit
631 discussion and make it difficult to accurately compare one species to another. There are two
632 primary reasons for the absence of definition in shark literature. First, there is inherent
633 difficulty associated with applying ecological theory to an analytical framework (Chase and
634 Leibold 2009). Second, it has only recently become possible to develop accurate theoretical
635 guidelines for defining and measuring shark resource use due to advancements in animal
636 tracking and dietary analysis. As a result, there has been limited scientific discussion on how
637 to define specialisation and classify sharks. However, as the aforementioned terms are used
638 more often (ex. Taylor and Bennett 2008; Wilga *et al.* 2012), it is important to link applied
639 science with the theoretical concepts of resource use. Lack of definition will lead to the
640 continued misuse of concepts, the incorrect classification of species, hinder research progress,
641 and impede successful management efforts.

642 Definitions of each concept and how best to measure them can vary based on context and the
643 authors analytical intent (Berenbaum 1996; Poisot *et al.* 2012). Therefore, a well-structured
644 definition and conceptual framework for specialisation should be tailored for specific types of
645 research. How specialisation should be defined is dependant not only on broader ecological
646 theory, but also on the organism, the scale of the study (species to individual), and the type of
647 specialisation being evaluated (e.g. behavioural, dietary, habitat) (Irschick *et al.* 2005;
648 Devictor *et al.* 2010). Moreover, selection and specialisation have similar quantitative and
649 qualitative characteristics. Therefore, it is important to define each term in context to prevent
650 interchangeable and inconsistent use. To alleviate the current confusion in shark literature,
651 highlight the importance of contextual definitions in shark ecology, and showcase the
652 diversity of strategies among sharks I will (1) discuss the theoretical differences between the
653 related but distinct ideas of selection and specialisation; (2) propose an ecological definition
654 and conceptual framework for resource specialisation widely applicable to shark species; (3)
655 review a range of ecological adaptations of sharks within the proposed context in two
656 important foci: diet and habitat; and (4) briefly discuss the potential implications of these
657 strategies on species resilience.

658

659 **2.2 Defining Shark Specialisation: Terminology and the Continuum** 660 **Concept**

661

662 Multiple forms of specialisation exist among sharks, including morphological specialisations
663 such as the long upper caudal lobe of the common thresher shark *Alopias vulpinus* (Aalbers *et*
664 *al.* 2010); behavioural specialisations such as the ectoparasitic feeding strategy of the cookie
665 cutter shark *Isistius brasiliensis* (Papastamatiou *et al.* 2010); and ecological specialisations
666 such as the specialised cephalopod-based diet of the whiskery shark *Furgaleus macki*

667 (Simpfendorfer *et al.* 2001b). Therefore differentiation between unique forms of
668 specialisation requires definitions designed to suit the goals and scale of the research
669 (Futuyma and Moreno 1988; Irschick *et al.* 2005). I am focused on defining shark ecological
670 specialisation. Therefore, all terminology has been tailored to fit within that context.

671

672 Four terms are primarily used to describe shark resource use. Selection is defined as the use
673 of resources out of proportion or in different proportions to their availability. Opportunism,
674 selection's conceptual opposite, is defined as the use of resources in the same proportion as
675 their availability (Johnson 1980; Buskirk and Millspaugh 2006). Selective animals bypass
676 certain resources in favour of others; whereas opportunistic animals use whatever is available.
677 It is typically quantified for one resource at a time. Measurements of several resources are
678 compared to each other to determine relative levels of selection for any given resource
679 (Manly *et al.* 2002). The term specialist describes species, populations, or individuals that
680 have a narrow or restricted niche breadth. Generalists are species, populations, or individuals
681 that have a large niche breadth (Vandermeer 1972; Devictor *et al.* 2010). Niche breadth is a
682 collective measurement of all resources used relative to the resources available within the
683 environment as a whole (Colwell and Futuyma 1971). Although selection and specialisation
684 may seem quite similar (i.e. a highly selective species is likely to have small niche breadth),
685 the terms are not interchangeable. The main difference being one is measured in reference to
686 an animal's niche (specialisation and generalisation) and the other measures the proportion of
687 an animal's resource use (selection and opportunism). Therefore analytical methods that can
688 determine an animal's selectivity may not be equally capable of measuring specialisation.

689

690 To build a definition for specialisation that is more specific and better suited to shark resource
691 use, an ecologically appropriate conceptual framework must be developed. Although animals
692 are often defined using the opposing categories of specialist or generalist, in nature,
693 specialisation is more accurately depicted as a continuum, where species are ranked along a
694 specialisation gradient ranging from the largest to smallest possible niche. In this framework,
695 sharks become less specialised with increasing niche breadth. Highly specialised species are
696 those with very small niche breadths. A continuum context for defining shark specialisation is
697 beneficial because while some sharks may be good examples of specialists or generalists,
698 most sharks will not meet the strict requirements of either definition. Such species cannot be
699 easily categorized. For example, a hypothetical species that selects for specific resources in a
700 given environment, while avoiding others, cannot be defined as a generalist without exception
701 because it has a somewhat restricted niche. However, if the same species has a large niche
702 compared to other species in the environment, it is also not a specialist. Such strategies and
703 resulting niche breadth values exist somewhere between a generalist and a specialist.
704 Therefore, when studying shark resource use patterns and defining behaviours, it is more
705 appropriate to measure and refer to a species' degree of specialisation, rather than
706 categorizing a species as one extreme or the other. This concept is also quantitatively
707 appropriate as most traditional measurements of specialisation use a sliding scale or index to
708 measure specialisation (e.g. Levins 1968; Feinsinger *et al.* 1981; Smith 1982). Modern
709 methods also measure specialisation using some form of gradient or relative comparison (e.g.
710 Julliard *et al.* 2006; Fridley *et al.* 2007; Peers *et al.* 2012).

711

712 **2.3 Defining Shark Specialisation: The Ecological Niche of a Shark**

713

714 To finalize a definition and contextual framework for shark specialisation, I must also define
715 the ecological niche of shark species. This is necessary to implement a test for specialisation
716 because it is the main component of the definition. However, there are a number of different
717 definitions and contradictory visions of the niche concept. It is also an unpopular term with
718 some ecologists (Chase and Leibold 2009). As a result there can be confusion as to what
719 ecologists mean when they discuss the niche of a species (Leibold 1995; Peterson *et al.*
720 2011). I will not enter into a broader discussion of niche theory, as I am only interested in
721 finding a functional definition for niche as it pertains to shark resource use. With that in mind,
722 I will briefly review the most prevalent contributions to niche theory to develop an
723 appropriate definition for shark specialisation.

724

725 Two major conceptual contributions to niche theory have dominated modern definitions. The
726 first was put forward by Grinnell (1917) and advanced by Hutchinson (1957). According to
727 these works a niche is defined as the range of resources a species uses in an environment or
728 the conditions in which it can survive. In other words, a Grinnellian niche is defined by “what
729 a species needs” or uses (Devictor *et al.* 2010). The second definition was developed by Elton
730 (1927), who defined a species niche as the role of a species in the environment or by “what
731 the species is doing” (Devictor *et al.* 2010). This definition requires measurements of all the
732 ways in which a species might directly and indirectly effect the environment (Leibold 1995;
733 Devictor *et al.* 2010).

734

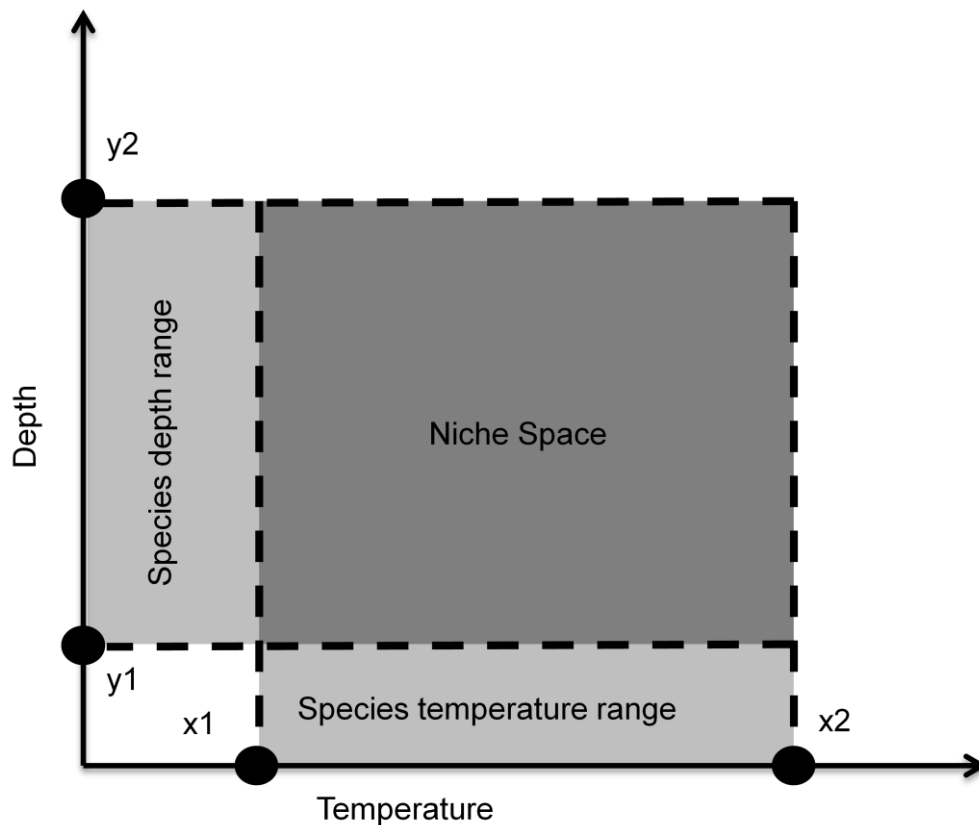
735 Hutchinson (1957) developed a physical schematic for his definition known as an n-
736 dimensional hyper-volume. Hutchinson proposed that a niche could be plotted on a Cartesian
737 coordinate system where the axes of the plot are environmental variables, such as temperature

738 (x) or depth (y) (Fig. 2.1). The limiting values or range of values a species uses can be plotted
739 on each axis. The overlapping ranges of each variable result in a figure that represents the
740 niche space for the species. Any point in this space represents an environmental state in
741 which the species can survive. Any point outside of it is not considered a part of the species
742 niche. The space can be defined by any number of relevant environmental variables that
743 affect the species (Hutchinson 1957; Chase and Leibold 2009), eventually creating a multi-
744 dimensional hyper volume. A less specialised species niche space would be distributed
745 among all or a large portion of the environment's resources and conditions (Vandermeer
746 1972; Fig. 2.2a). A highly specialised species niche space would be relatively concentrated
747 over one or a few resources or conditions (Vandermeer 1972; Fig. 2.2b). Modern applications
748 of niche theory often incorporate both Hutchinson's and Elton's contributions, defining the
749 ecological niche as the response that a species has to each point in Hutchinson's classical
750 environmental space (measured as species survival rate, growth rate, or per capita rate of
751 increase) and the effect that the species has on each point (measured as resource consumption,
752 competition, etc.) (Leibold 1995; Shea and Chesson 2002; Chase and Leibold 2009).

753 Information on species competition and predator-prey relationships is important when trying
754 to define the role of a species in the environment or its relationship with other species.

755 However, Hutchinson's definition is more appropriate when measuring resource
756 specialisation of sharks because it only considers the resource requirements of species, which
757 is precisely what resource use studies quantify. Moreover, although not impossible, it can be
758 difficult to measure the response of a species in the wild and relate that response to a
759 particular point and variable in the niche space. The advantage of using a "requirements only"
760 approach is researchers do not need to measure response variables (Peterson *et al.* 2011).

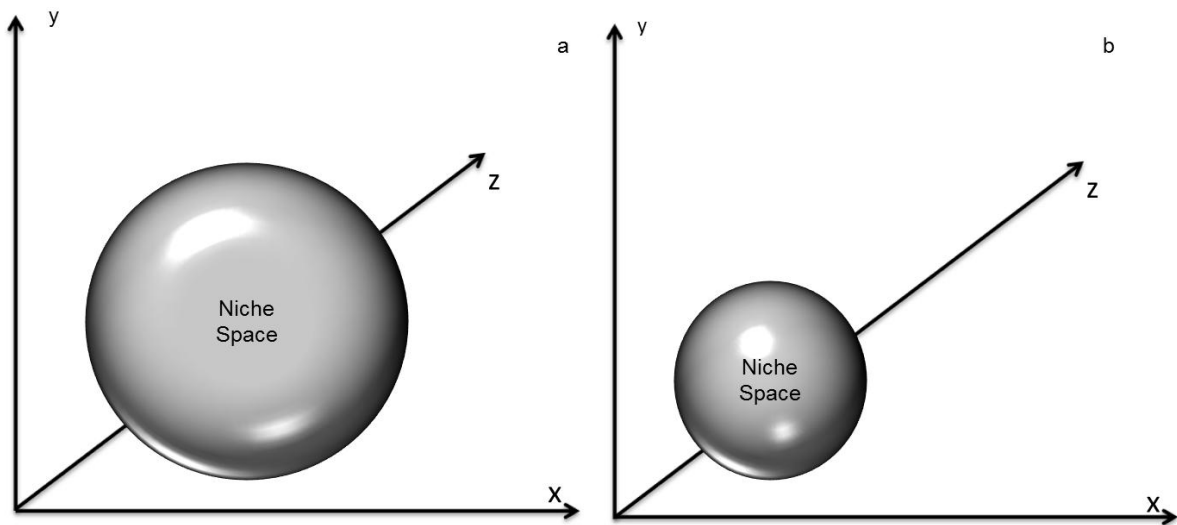
761



762

763 **Fig. 2.1.** A hypothetical example of Hutchinson's niche space with two commonly examined
764 shark habitat use variables, depth (y) and temperature (x). x_1 and x_2 denote a species'
765 temperature range, y_1 and y_2 denote a species' depth range. The dark grey space created by
766 these overlapping ranges is the niche space of an organism, any point within which it can
767 survive.

768



770
771

772 **Fig. 2.2.** (a) Hypothetical depiction of Hutchinson's (1957) niche hyper-volume of a species
773 with a large niche breadth, indicating a lesser degree of specialisation (b); Hypothetical
774 depiction of Hutchinson's (1957) niche hyper-volume of a species with a narrow niche
775 breadth, indicating a greater degree of specialisation.

776

777

778 There are also numerous methods available to measure specialisation that incorporate
779 Hutchinson's niche model. The best method for analysis will depend on the goals of the
780 experiment. Regardless of the method chosen, at their core tests will compare the range of
781 resources used by a species (the niche space) to the range of resources available in the
782 environment, resulting in the relative niche breadth of the species (e.g. Smith 1982; Basille *et*
783 *al.* 2008; Poisot *et al.* 2012). The inverse of the relative niche breadth can be used to measure
784 the degree of specialisation.

785

786 Using Hutchinson's definition of niche space and the continuum concept described earlier, it
787 is possible to develop a widely applicable definition for measuring shark specialisation. I
788 propose sharks, rather than being categorically defined, should be ranked along a continuum
789 as more or less specialised relative to an index and other species. Ranks or positions along
790 this continuum should be determined by measuring the resource niche breadth of a species as
791 described by Hutchinson's hyper-volume. Therefore, a highly specialised species can be
792 defined as one with a relatively narrow niche breadth compared to the range of resources or
793 conditions available within the environment. It is essential that the methods for measuring
794 niche breadth use a scale or index. Methodologies should not rely solely on a binary system
795 of classification or an arbitrarily value within an index above or below which a species is
796 classified as a specialist. This violates the continuum concept that is the core of this
797 definition. Note that the relevant environmental parameters will be based on the scale of the
798 experiment and are best left to the researcher to define. However, definitions of the available
799 environment will affect the interpretation of any measure of specialisation and its
800 comparability to other studies. Therefore environment and scale should be carefully

801 considered when utilizing ecological terminology, choosing appropriate methods, and making
802 assessments.

803

804 This definition is well suited to sharks as specialisation is often measured at different
805 temporal and physical scales. The environmental and temporal parameters of any study can
806 be adjusted to suit a ranking system. The use of a continuum concept is also more
807 ecologically realistic as many sharks are not likely to meet the traditional definition of a
808 specialist or generalists. A rank system avoids the use of definitive categorical language when
809 it is not appropriate. Finally, a ranking system which defines species as more or less
810 specialised also makes it easier to compare trends between groups, even those separated over
811 large distances that may be exposed to different resources. A ranking system that compares
812 niche breadth instead of selectivity for a single resource may better account for differences in
813 environmental circumstances.

814

815 Unfortunately, limited information on the diet or habitat use of many sharks currently
816 precludes the ability to fully assess species resource specialisation. Therefore, the ranking of
817 some species as more or less specialised will be unavoidably speculative. However, having a
818 clear understanding of what is meant by specialist gives researchers the ability to better
819 classify species, study their role in the environment, and communicate their findings.

820

821 **2.4 Shark Habitat Specialisation**

822

823 Species that have a high degree of habitat specialisation inhabit a smaller niche space
824 compared to their less specialised counter parts. As a result habitat restricted species will

825 occupy one end of the specialist continuum and those that use a large number of habitats will
826 occupy the other. More specific rankings of organisms are affected by the scale of the study
827 and the habitat parameters evaluated. Habitat use can be evaluated at large (e.g. Weng *et al.*
828 2005) or small scales (e.g. Morrissey and Gruber 1993) and specialisation can be measured
829 across numerous variables, including depth (Knip *et al.* 2011), temperature (Campana and
830 Joyce 2004), salinity (Heupel and Simpfendorfer 2008), and bottom type (Espinoza *et al.*
831 2011). Patterns in habitat use may also change over time (e.g. daily, seasonally) (Carlisle and
832 Starr 2009; Ortega *et al.* 2009). Therefore, when analysing habitat specialisation it is
833 important to clarify the scale of the study in relation to the continuum definition. On a local
834 scale, most sharks are thought to have relatively large habitat ranges; however, there are few
835 measurements of niche breadth or broad studies of habitat use for many shark species.
836 Moreover, scale and methodology differ widely between studies, making it difficult to rank
837 species against one another. Therefore, I will limit this discussion to better studied species
838 and research testing selection or specialisation to highlight the importance of defining scale in
839 resource use studies and the diversity among species.

840

841 On a global scale, species with low levels of habitat specialisation include the tiger shark,
842 *Galeocerdo cuvier* and the great white shark *Carcharodon carcharias*. *Galeocerdo cuvier* has
843 been found in temperate and tropical waters with reports of individuals being found as far
844 north as Iceland and the United Kingdom (Compagno 1984; Randall 1992). They are also
845 known to use coastal and offshore habitats (e.g. Randall 1992; Holland *et al.* 1999; Heithaus
846 *et al.* 2007; Meyer *et al.* 2009). *Carcharodon carcharias* also have large, cosmopolitan
847 geographic ranges and use both coastal and oceanic habitats (Compagno 2001; Bruce 2008;
848 Jorgensen *et al.* 2010; Carlisle *et al.* 2012), and are known to undertake wide ranging
849 migrations (Pardini *et al.* 2001; Boustany *et al.* 2002; Bonfil *et al.* 2005; Weng *et al.* 2007).

850 However, habitat use is more often evaluated at smaller scales (regionally and locally) and
851 low specialisation rankings can be applied to species if the scale of evaluation is clear. For
852 example, species may have low degrees of specialisation over a single environmental
853 parameter. Juvenile lemon sharks *Negaprion brevirostris* acoustically tracked in nursery
854 grounds in Bimini Island, Bahamas, showed no preference for specific water temperatures.
855 Instead, individuals selected for the warmest possible waters during the day and cooler waters
856 in the evening (DiGirolamo *et al.* 2012). As a result, individuals used the range of available
857 temperatures within its home range in a day (DiGirolamo *et al.* 2012). Based on the proposed
858 definition and measured at a local scale, the use of a wide range of temperatures relative to
859 the daily available range indicates this population has a low degree of temperature
860 specialisation. However, DiGirolamo *et al.* (2012) did not directly test species niche breadth
861 or selectivity. Also note that while juveniles have a low degree of specialisation, there was
862 selection for temperatures depending on the time of day. This highlights the importance of
863 separating the concepts of specialisation and selection.

864

865 Species that have moderate habitat niches will have neither high nor low degrees of
866 specialisation. Species with varying levels of resource selectivity, that may avoid some
867 habitats while using others, will likely equate to a moderate niche breadth. Such species
868 cannot be easily defined in categorical terms and demonstrate the usefulness of a continuum
869 scheme for specialisation. For example, young bull sharks, *Carcharhinus leucas*, tracked in
870 the Caloosahatchee River Estuary, Florida, showed strong avoidance for areas with salinities
871 less than 7 while showing affinity for salinities between 12 to 20 (Heupel and Simpfendorfer
872 2008). Heupel and Simpfendorfer (2008) suggested juveniles may have been selecting for
873 salinity ranges that reduced their osmoregulatory costs. The importance of salinity in
874 predicting the distribution of young *C. leucas* was also suggested by Simpfendorfer *et al.*

875 (2005), however, this study did not use selectivity indices to analyse behaviour. Although
876 niche breadth was not measured, the avoidance of some salinity ranges and the selection of
877 others would likely result in a moderate niche breadth and degree of specialisation for this
878 population. This example also demonstrates the importance of clarifying definitions for
879 selection and specialisation. *Carcharhinus leucas* may be selecting for specific salinities, but
880 they are not highly ecologically specialised along this parameter. Scale and location will also
881 affect how a ranking is determined. For example, although *G. cuvier* utilizes a wide range of
882 habitats globally, on a local scale they appear to select for specific microhabitats.
883 Acoustically tracked *G. cuvier* in Shark Bay, Western Australia, selected for shallow banks
884 covered in seagrass and avoided deeper sand habitats (Heithaus *et al.* 2006). Selectivity was
885 likely the result of increased prey availability within shallow areas (Heithaus *et al.* 2002;
886 Heithaus *et al.* 2006). As a result *G. cuvier* habitat niche breadth in nearshore areas is likely
887 smaller than that expected at a global level.

888

889 A lack of studies that measure the habitat niche breadth of sharks makes any speculative
890 rankings presented here potentially controversial. However, some of the best examples of
891 habitat specialisation among sharks are coral reef associated species, a notable exception
892 being the blacktip reef shark, *Carcharhinus melanopterus* (Chin *et al.* 2012). Species with
893 high degrees of specialisation likely include the whitetip reef shark *Triaenodon obesus*, the
894 grey reef shark *Carcharhinus amblyrhynchos*, and the Caribbean reef shark, *Carcharhinus*
895 *perezi*. *Triaenodon obesus* is most often found within or very near reef habitat and its
896 morphology indicates it is well adapted to forage in reef environments (Randall 1977; Last
897 and Stevens 2009; Whitney *et al.* 2012). *Carcharhinus amblyrhynchos* is almost exclusively
898 found on and near reef habitat (Compagno 1984; McKibben and Nelson 1986; Economakis
899 and Lobel 1998; Heupel and Simpfendorfer 2014), although individuals may make long

900 distance movements between habitat patches (Heupel *et al.* 2010). *Carcharhinus perezi* is
901 also closely associated with reef habitat (Pikitch *et al.* 2005; Garla *et al.* 2006a; Garla *et al.*
902 2006b; Chapman *et al.* 2007).

903

904 Unfortunately, determining where sharks should be placed along the specialist continuum is
905 limited as little is known about the habitat preferences of most species. Data collection is
906 hindered by the difficulties associated with habitat use evaluation, particularly in the marine
907 environment. Traditional techniques used to evaluate animal movements and habitat
908 preferences, such as tag and recapture and fishing surveys, only provide short-term (< 10 yr)
909 data and do not monitor the lifetime of an individual. While these studies can offer valuable
910 insight into animal distribution (Kohler and Turner 2001), population size (Dicken *et al.*
911 2008), and survival rates (Gruber *et al.* 2001), these techniques only provide snapshots of
912 individual habitat use (Gruber *et al.* 1988; Holland *et al.* 1993; Holts and Bedford 1993). It
913 may also be difficult to measure the availability of various habitat types and variables
914 meaningful to the animals in the environment. Therefore, accumulating data for habitat use
915 assessments can be a slow process and the degree of habitat specialisation of many species
916 will remain uncategorized in the near future. However, based on the previous examples it is
917 clear there is significant variance in the habitat specialisation and selection patterns of sharks.
918 These examples also demonstrate how defining scale and intent has a large influence on the
919 use of ecological terminology and its interpretation. By utilizing a continuum concept of
920 specialisation, parameters can be set according to the needs of the research and result in less
921 arbitrary use of terms. Detailed analysis of shark habitat use incorporating sound definitions
922 as well as new techniques may reveal that more species are highly selective or more
923 specialised than currently known.

924

925 **2.5 Shark Dietary Specialisation**

926

927 Optimal foraging theory states that individuals should attempt to forage at maximum
928 efficiency to ensure a large net energy return for their efforts (Townsend and Winfield 1985;
929 Stephens 1986). However, evaluation of foraging theory and dietary specialisation are context
930 dependant. Fluctuating factors such as competition may cause species that naturally adopt
931 opportunistic feeding patterns to become highly selective (e.g. Papastamatiou *et al.* 2006).
932 Diets may differ between species, populations of the same species, and between age classes
933 and sexes within populations (Bethea *et al.* 2006; Edwards *et al.* 2011; Sommerville *et al.*
934 2011). Reasons for this include differences in body shape and size, ability to locate and
935 capture prey, and ability to process and digest what has been caught (Lowe *et al.* 1996;
936 Heupel and Bennett 1998; Dean *et al.* 2005; Bethea *et al.* 2006; Brischoux *et al.* 2011).
937 Predator avoidance tactics may lead to a different diet than that predicted by optimal foraging
938 theory (Gill 2003) and individuals may forage sub-optimally if doing so reduces their
939 exposure to predation (Heithaus and Dill 2002). Finally, dietary analysis can be skewed by
940 temporal shifts in prey availability where predators switch between prey items as they
941 become more or less available (e.g. Lucifora *et al.* 2006). Therefore, evaluations over short
942 time scales may be misleading in fluctuating environments. The method of evaluation may
943 also affect definitions and interpretations of species dietary patterns. For example, traditional
944 methods such as stomach content analysis provide detailed information on dietary patterns
945 (e.g. Ba *et al.* 2013) , whereas stable isotope analysis, an increasingly popular method for
946 shark diet and movement analysis, can provide long term, integrated data (Hussey *et al.*
947 2012a). Therefore, when measuring niche breadth or diet selectivity it is necessary to consider
948 environmental and morphological variables and what is available to the predator, both in

949 terms of prey abundance and what it has the ability to capture (Backwell *et al.* 1998).
950 Therefore, similar to habitat analysis, it is important to clarify the temporal and spatial scale
951 and intention of a study when calculating diet specialisation and selection.

952

953 At a global scale, species that are traditionally considered to have low degrees of dietary
954 specialisation include the spiny dogfish *Squalus acanthias*, which feeds on a diverse array of
955 prey over its entire range (Jones and Geen 1977; Compagno 1984; Hanchet 1991; Tanasichuk
956 *et al.* 1991; Link and Ford 2006; Brodeur *et al.* 2009). In Argentinean waters, *S. acanthias*
957 feeds on teleosts, squid, ctenophores and molluscs (Alonso *et al.* 2002). In the southeastern
958 Black Sea, *S. acanthias* preys on teleosts, crustaceans, sea anemones and nematodes (Avsar
959 2001). Larger species with broad diets include *G. cuvier*. In Hawaiian waters, large
960 individuals (>200 cm) were found to have a varied diet that included teleosts, elasmobranchs,
961 crustaceans, birds, mammals, turtles, and cephalopods (Lowe *et al.* 1996). In Australian
962 waters, *G. cuvier* also has a wide dietary niche. Individuals on the western Australian coast
963 feed on teleosts, crustaceans, marine mammals and reptiles, elasmobranchs and cephalopods
964 (Heithaus 2001; Simpfendorfer *et al.* 2001a), while on the north eastern coast *G. cuvier* feed
965 on teleosts, marine reptiles, crustaceans, and to a lesser extent marine mammals and
966 cephalopods (Simpfendorfer 1992a). At a smaller regional scale, stomach content analysis of
967 the small spotted cat shark *Scyliorhinus canicula* from the north eastern Atlantic coast found
968 the species fed on a variety of prey, including crustaceans, teleosts, annelids, and molluscs
969 (Ellis *et al.* 1996). The dietary niche of *S. canicula* was calculated and equated to a low
970 degree of dietary specialisation. In comparison to other species similarly surveyed in the same
971 study, it had one of the largest dietary niches.

972

973 Variability in diet selectivity can result in niche breadths that rank species in between the two
974 extremes of the specialisation continuum. For example, Baremore *et al.* (2008) examined the
975 stomach content of Atlantic angel sharks *Squatina dumeril* captured in the northeastern Gulf
976 of Mexico and compared it to the trawl fishery catch composition to calculate *S. dumeril*
977 niche breadth index and dietary selectivity. Stomach content analysis revealed *S. dumeril* fed
978 on a variety of prey, but when compared to prey availability, the diet of *S. dumeril* equated to
979 a moderate niche breadth (Baremore *et al.* 2008). These results indicated *S. dumeril* was
980 neither highly specialised nor generalised. Thus, *S. squatina* demonstrated both opportunistic
981 and selective behaviours by feeding on fish in high abundance (opportunistic) as well as fish
982 and cephalopods found in relatively low abundance (selective). This example also highlights
983 the importance of using the continuum concept to measure specialisation rather than trying to
984 categorically define species as either specialists or generalists. Varied patterns in behaviours
985 and changing conditions may result in niche breadth values that cannot be easily designated
986 as one or the other.

987

988 Dietary selectivity can also change as the result of fluctuations in prey abundance and
989 availability over time and space. Changes in dietary patterns can affect niche breadth
990 measurements and alter the predicted placement of a species on a specialisation continuum.
991 For example, although juvenile *N. brevirostris* has a broad dietary niche, and therefore has a
992 low degree of specialisation, a controlled field-pen study found *N. brevirostris* may be a
993 highly selective predator. Caged *N. brevirostris* were fed varying ratios of two prominent
994 prey in their diet, the grey snapper *Lutjanus griseus* and the yellow fin mojarra *Gerres*
995 *cinereus*. Results showed *G. cinereus* was selected for over *L. griseus* and that selectivity for
996 *G. cinereus* increased as its relative abundance increased, highlighting the effect of changing
997 prey abundance on diet and indicating *G. cinereus* is a preferred prey (Reeve *et al.* 2009).

998 Both *N. brevirostris* dietary selectivity and preference for mojarra (Gerreidae) were supported
999 by field based stomach content analysis (Newman *et al.* 2010). Populations within the bays of
1000 Bimini Island selected for prey based on type and size when environmental conditions were
1001 favourable and prey abundance was high (Newman *et al.* 2010). However, populations fed
1002 more opportunistically in relatively poor conditions. This result suggests niche breadth and
1003 feeding strategies change in response to environmental fluctuations. High levels of selectivity
1004 for specific types of prey in favourable conditions may result in a more specialised diet than
1005 when conditions are poor.

1006

1007 Competition can also have a powerful influence on dietary selection. Co-occurring shark
1008 species surveyed in the coastal waters of Hawaii showed limited dietary overlap, but when the
1009 diets of these species were surveyed in areas where they did not co-occur, high dietary
1010 overlap was observed (Papastamatiou *et al.* 2006). This suggests that to reduce competition
1011 for resources, each species selected for a non-overlapping subset of resources in the
1012 environment. However, when competition was removed or reduced, species adopted wider
1013 dietary niches. Spatial variation in selection would create variable rankings for the same
1014 species and confound a binary attempt to define the species or adjacent populations.

1015

1016 It is important to note that some populations that have large dietary niches may be composed
1017 of individual specialists, where each individual uses a subset of resources within the
1018 population's broader dietary niche (Bolnick *et al.* 2002). The combination of individual non-
1019 overlapping, selective diets results in a wide dietary niche for the population. Surveys of
1020 populations that do not test for the presence of individual specialisation may incorrectly
1021 classify individuals as having large dietary niches (Bolnick *et al.* 2002; Bolnick *et al.* 2003).

1022 However, it is difficult to determine how common this strategy is among sharks as it has only
1023 recently been investigated among a few species. Matich *et al.* 2011 quantified the dietary
1024 patterns of *C. leucus* and *G. cuvier*, both of which have traditionally been defined as
1025 generalist predators, using several individual specialisation indices. The indices revealed that
1026 individual *G. cuvier* had wider ranging diets relative to individual *C. leucus* which had more
1027 specialised dietary profiles. This suggests the broad dietary range of *C. leucas* may be the
1028 result of individual specialisation, not each individual feeding on a large number of prey
1029 (Matich *et al.* 2011). More detailed analysis will hopefully determine if individual
1030 specialisation is a widespread or significant strategy within the taxon.

1031

1032 To date there are few studies that have been able to identify species that exhibit high degrees
1033 of dietary specialisation and that can be labelled as such without debate. However, there are
1034 some examples where species can be classified as highly specialised. On a global scale,
1035 dietary specialists include basking sharks *Cetorhinus maximus* and whale sharks *Rhincodon*
1036 *typus*, both of which have a highly specialised feeding mechanism (Hallacher 1977; Colman
1037 1997) evolved to capture prey of a specific size, namely zooplankton (Colman 1997; Sims
1038 and Quayle 1998; Stevens 2007). In the case of *R. typus*, prey include fish spawn and
1039 plankton (Martin 2007), crab larvae (Meekan *et al.* 2009), copepods (Clarke and Nelson
1040 1997; Motta *et al.* 2010), and krill (Jarman and Wilson 2004). On smaller geographic scales, a
1041 potential dietary specialist includes *F. macki*, a species endemic to western Australia (Last
1042 and Stevens 2009). Although the diet of *F. macki* was not compared to prey availability,
1043 stomach content analysis revealed this species feeds almost exclusively on octopus and other
1044 cephalopods. The diet of *F. macki* was significantly less varied than other shark species
1045 captured simultaneously in the same habitats (Simpfendorfer *et al.* 2001b). Stomach content
1046 analysis of the starry smooth-hound *Mustelus asterias* captured in trawl surveys on the north

1047 eastern Atlantic coast showed this species almost exclusively consumed brachyuran crabs.
1048 Niche breadth measurements revealed *M. asterias* had a high degree of dietary specialisation
1049 in comparison to almost all other elasmobranchs examined in the study (Ellis *et al.* 1996).

1050

1051 The term specialist can also be applied at smaller physical or temporal scales; however
1052 context must be clearly articulated. For example, the stomach contents of the school shark
1053 *Galeorhinus galeus* surveyed in Anegata Bay, Argentina revealed this species had a broad
1054 dietary niche (Lucifora *et al.* 2006). However, Lucifora *et al.* (2006) found that during the
1055 astral summer the diet of *G. galeus* became highly specialised when individuals fed almost
1056 exclusively on the benthic teleost the Atlantic midshipman *Porichthys porosissimus* (Lucifora
1057 *et al.* 2006). This seasonally small niche breadth was likely the result of opportunistic
1058 foraging on the seasonally vulnerable *P. porosissimus*. *Porichthys porosissimus* mates in the
1059 spring and summer when males use sound and bioluminescent displays to attract their mates.
1060 Lucifora *et al.* (2006) hypothesized these displays make *P. porosissimus* more vulnerable to
1061 predators and as a result are more easily targeted in summer than at other times of the year.
1062 However, caution should be used when considering a species highly specialised over small
1063 scales to ensure there is no confusion over intent.

1064

1065 **2.6 Implications of Resource Use Strategies**

1066

1067 Ecological theory states that the niche breadth differences between species are the product of
1068 an evolutionary trade-off between the ability to use multiple resources and the ability to use
1069 each one. Different strategies result in unique costs and benefits for species (Van Tienderen
1070 1991; Kawecki 1994). Ecological specialisation of sharks will also affect their role in an

1071 ecosystem, such as the intensity and spread of direct and indirect effects of predation (Heupel
1072 *et al.*2014). Therefore, defining the resource use patterns of sharks can help to determine the
1073 implications of their depletion or removal. Understanding the trade-offs and vulnerabilities
1074 associated with ecological specialisation can create a clearer picture of environmental
1075 dynamics and the role of sharks.

1076

1077 The ability to use multiple resources, and therefore have a low degree of specialisation, is
1078 advantageous during times of environmental instability (Chapman and Mackay 1984;
1079 Heithaus *et al.* 2006; Meyer *et al.* 2010). Species with wide niches can use resources
1080 unaffected by environmental fluctuations and/or use multiple resources to compensate for the
1081 decline in any one resource (Richmond *et al.* 2005; Julliard *et al.* 2006; Verberk *et al.* 2010).
1082 Thus, species with wide resource niches can maintain a high level of fitness in unstable
1083 environments and will not be as greatly affected by environmental changes than highly
1084 specialised species (Richmond *et al.* 2005; Julliard *et al.* 2006; Chin *et al.* 2010; Verberk *et*
1085 *al.* 2010). However, these plastic adaptations may incorporate anatomical, physiological, or
1086 behavioural mechanisms that require high levels of energy (DeWitt *et al.* 1998). If
1087 environments are stable, these high energetic costs may outweigh the benefits of being highly
1088 adaptive (Van Tienderen 1991; Wilson and Yoshimura 1994; DeWitt *et al.* 1998)

1089

1090 In contrast, high degrees of specialisation are associated with the reduction in or loss of
1091 physiological, morphological, or behavioural characteristics, which theoretically reduces
1092 energetic costs (Futuyma and Moreno 1988). By adapting to use only one or a few resources,
1093 species avoid the high costs of adaptive plasticity (Van Tienderen 1991). Resource detection
1094 performance is also greater when an animal is only searching for one item (Futuyma and

1095 Moreno 1988; Bernays and Wcislo 1994; Bernays and Funk 1999). Highly specialised species
1096 may also utilize resources and assimilate energy from preferred sources more efficiently than
1097 species with broad resource niches (Britt *et al.* 2006). Therefore, if fluctuations in the
1098 environment are limited and the preferred environmental state is abundant, specialisation may
1099 be a more successful life strategy (Futuyma and Moreno 1988; Richmond *et al.* 2005).
1100 However, species with high degrees of specialisation incur costs when their preferred
1101 resource is not available. Highly specialised species cannot easily switch between resources
1102 and as a result, when environmental conditions are in a state of flux, they may have difficulty
1103 adapting to the new environmental equilibrium (Dulvy *et al.* 2004; Munday 2004; Chin *et al.*
1104 2010). Thus, highly specialised shark species are at increased risk to population decline as a
1105 result of environmental and anthropogenic changes compared to species with large ecological
1106 niches.

1107

1108 **2.7 Conclusion**

1109

1110 Sharks have been historically described as roaming generalists that feed on whatever
1111 resources become available. In reality, various strategies are present among shark species,
1112 including selective and opportunistic feeding behaviours as well as highly specialised
1113 resource use patterns. As the field of shark ecology expands and advances, it is important to
1114 develop clear, inclusive, and theoretically sound definitions and methodologies to study
1115 resource use at small and large scales. Doing so will allow for efficient communication of
1116 ideas and more comparable research. Shark ecology, compared to other fields of ecological
1117 research, is in its early days, and detailed studies of resource selection and niche breadth have
1118 only recently become a prominent feature of the shark literature. This provides shark
1119 researchers with a unique opportunity. Shark ecologists can avoid some hurdles associated

1120 with applied ecological studies by deciding now what the most appropriate theoretical basis is
1121 to ground analytical studies.

1122

1123 Although it is difficult to quantify the resource use patterns of many shark species due to
1124 limited data, this will change as research progresses, and having a structured paradigm within
1125 which to evaluate behaviours will be invaluable to furthering research and communication.
1126 Improved knowledge of where shark species fall on the specialisation continuum will result in
1127 more accurate predictions of the effects of human induced changes and the development of
1128 more effective environmental management. Future research aimed at examining resource use
1129 and selectivity of sharks should ensure that tests are based in well supported theoretical
1130 schemes and authors clarify their intent by defining terminology and ensuring they are only
1131 used when appropriate.

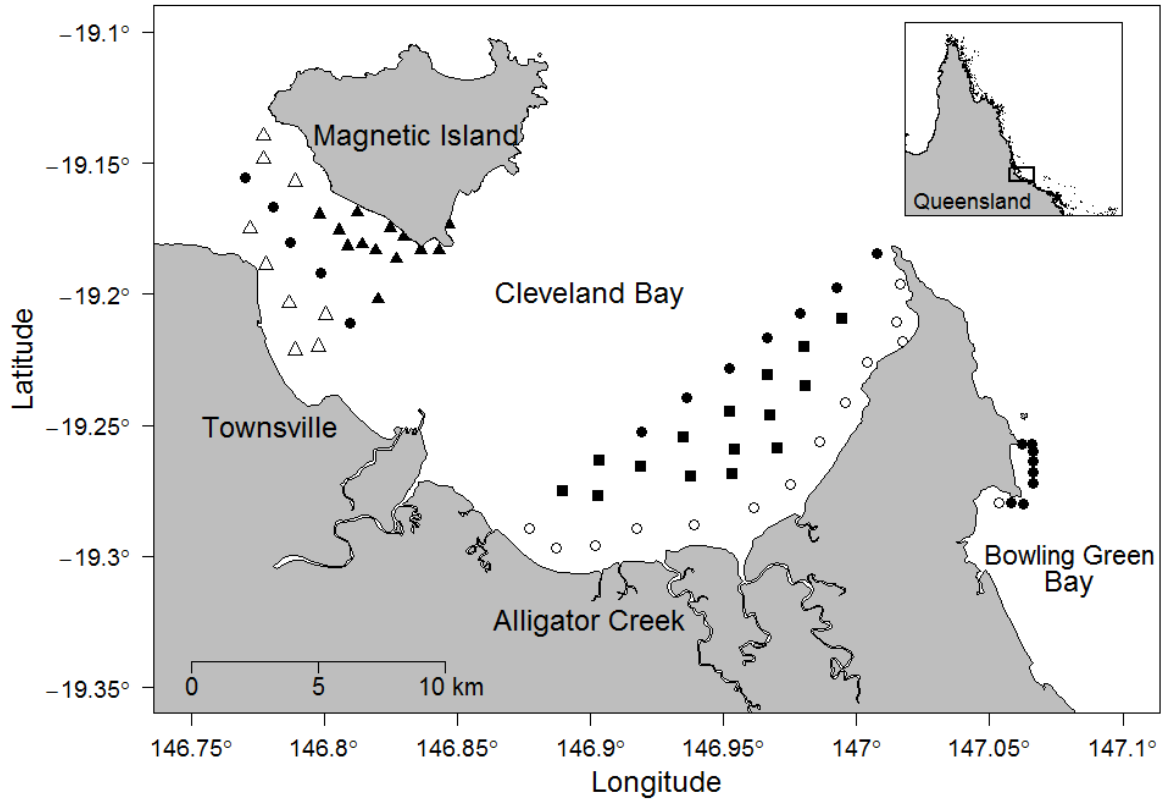
Chapter 3

General Methods

3.1 Acoustic Analysis

3.1.1 Study Site

Acoustic tracking was conducted in Cleveland Bay, Queensland, a shallow embayment on the northeast coast of Australia (Fig. 3.1). Cleveland Bay covers an area of approximately 225 km², is 27 km wide, and the majority of the bay has a depth of less than 10 m and a maximum tidal range of 4.2 m. The dominant habitat is soft mud substrate and to a lesser extent sandy substrate. The bay also contains patches of seagrasses (*Cymodocea serrulata*, *Halophila spp.*, *Halodule uninervis*) and coastal reefs. The southern shore of the bay is lined with mangroves. The main river outlets are on the southeastern side of the bay and are adjacent to intertidal mudflats and seagrass habitat. Sixty-three VR2W acoustic receivers (Vemco Ltd., Canada) were deployed inside Cleveland Bay to monitor shark movements. Receivers were deployed in primary habitat types within the bay, specifically intertidal mudflats, outer bay mud substrate (> 5 m depth), sandy inshore substrate, reefs, and potential seagrass (here after referred to as seagrass). Receiver habitat type was assessed by scuba divers during initial deployment. Benthic habitat assessments in Cleveland Bay by the James Cook University program Seagrass Watch (seagrasswatch.com.au) were also used to determine the habitat designation of each receiver. The distribution of intertidal mudflats, outer bay mud substrate, sandy inshore substrate, and reef was consistent. However, seagrass distribution can change on a seasonal basis. It was not possible to conduct detailed benthic surveys throughout the study, therefore seagrass habitat was designated as potential seagrass habitat to acknowledge potential changes in density of seagrass over time. Data were downloaded from receivers



1157

1158 **Fig. 3.1.** Cleveland Bay, Queensland, Australia, locations of receivers in intertidal mudflat

1159 (○), seagrass (■), outer bay mud substrate (●), inshore sand (Δ), and reef habitat (▲).

1160

1161 every three months. An additional nine receivers were deployed by the Australian Institute of
1162 Marine Science (AIMS) in Bowling Green Bay adjacent to the southeast of Cleveland Bay.
1163 The majority of these receivers were deployed between depths of 9.2 to 11.0 m with mud
1164 substrate. Therefore they were classified as outer bay mud substrate receivers. Data from
1165 these receivers were not included in habitat, space use, or residency analysis.

1166

1167 **3.1.2 Study Species**

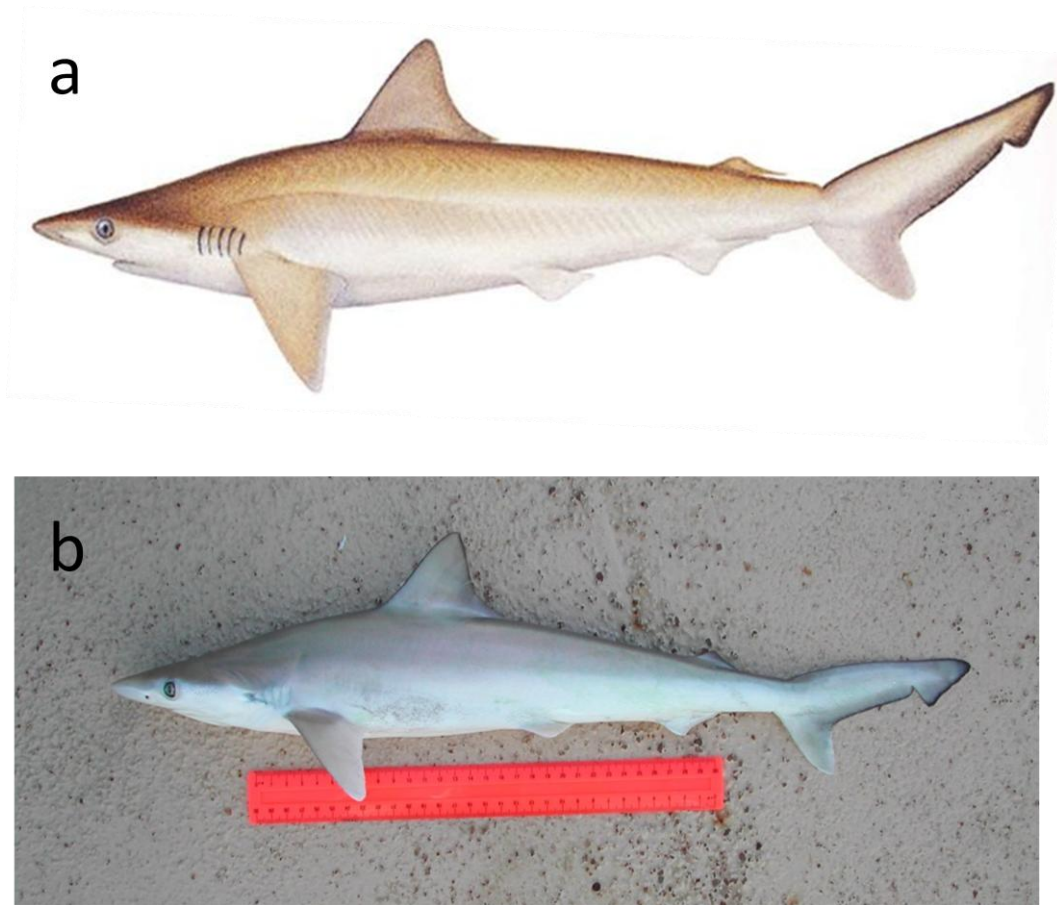
1168

1169 The study species for this research were the Australian sharpnose shark, *Rhizoprionodon*
1170 *taylori* (family Carcharhinidae; Fig. 3.2) and the creek whaler, *Carcharhinus fitzroyensis*
1171 (family Carcharhinidae; Fig. 3.3). *Carcharhinus fitzroyensis* is endemic to northern Australia
1172 although *R. taylori* can also be found on the southern coast of Papua New Guinea. The two
1173 species have closely overlapping coastal ranges, where *R. taylori* is found from Carnarvon
1174 (WA) to Moreton Bay (QLD) and *C. fitzroyensis* is found from Cape Cuvier (WA) to
1175 Gladstone (QLD) (Last and Stevens 2009). Both species most commonly inhabit turbid
1176 nearshore waters, but *R. taylori* has occasionally been captured on the outer continental shelf
1177 (Last and Stevens 2009). Neither species is a major component of northern Australian
1178 fisheries. *Rhizoprionodon taylori* is not directly targeted as it is too small to be of any value,
1179 however it is occasionally taken in large amounts as by catch in inshore gillnet and trawl
1180 fisheries. *Carcharhinus fitzroyensis* is taken in small numbers by the Australian gillnet fishery
1181 for meat. Teleosts and crustaceans constitute the majority of the diet of both species
1182 (Simpfendorfer 1998; Last and Stevens 2009). Although these species inhabit similar
1183 environments and have similarly broad diets, *R. taylori* and *C. fitzroyensis* have distinct life
1184 history strategies.

1185

1186

1187



1188

1189 **Fig. 3.2.** (a) *Rhizoprionodon taylori* from Last and Stevens (2009) and (b) photo of *R. taylori*
1190 (photo credit Centre for Sustainable Tropical Fisheries and Aquaculture).

1191

1192



1193

1194 **Fig. 3.3.** (a) *Carcharhinus fitzroyensis* from Last and Stevens (2009), (b) photo of *C.*
1195 *fitzroyensis* (photo credit Vinay Udyawer).

1196

1197 *Rhizoprionodon taylori* is an abundant, small-bodied, fast growing, highly productive species
1198 (Last and Stevens 2009). Size at birth is approximately 220-260 mm total length (TL); males
1199 and females mature at approximately 550 mm TL, and males grow to 690 mm TL and
1200 females to 810 mm TL (Simpfendorfer 1992b; Simpfendorfer 1993). Age at maturity is
1201 approximately one year and females can give birth to 1 to 10 pups per litter (mean = 4.5)
1202 (Simpfendorfer 1992b; Simpfendorfer 1993). This species has an annual reproductive cycle.
1203 Mating occurs in austral summer (December to February) and gestation lasts approximate
1204 11.5 months. *Rhizoprionodon taylori* is the only shark species known to incorporate a period
1205 of embryonic diapause in its reproductive cycle. This may allow *R. taylori* to delay giving
1206 birth until conditions are favourable (Simpfendorfer 1992b; Waltrick *et al.* 2012).

1207 In contrast, *C. fitzroyensis* is a medium-bodied species that is relatively slow growing and late to
1208 mature (Last and Stevens 2009). Size at birth is approximately 500 mm TL; males mature at
1209 approximately 800 mm TL and females 900 mm (Garrick 1982; Lyle 1987). Age at maturity
1210 is approximately six years. Adults grow to approximately 1350 mm. (Lyle 1987; Last and
1211 Stevens 2009). Age and growth estimates suggest females grow 200 mm larger than males
1212 (Smart *et al.* 2013). Female *C. fitzroyensis* give birth every year with 1 to 7 pups per litter
1213 (mean=3.7) following a gestation period of approximately 7 to 9 months (Lyle 1987).

1214

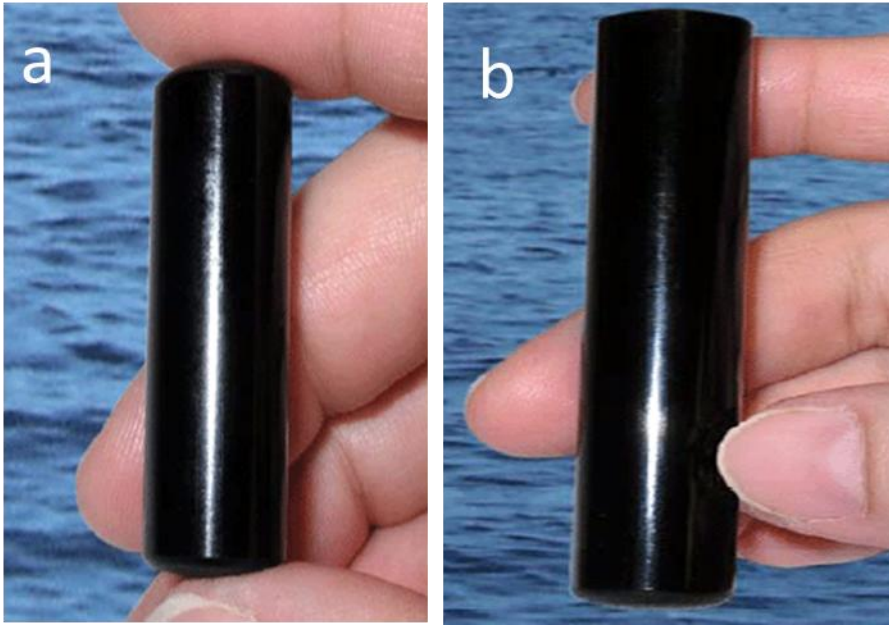
1215 Little is known about the movement and habitat use of either species. Previous catch data
1216 indicates that neonate *C. fitzroyensis* are found in intertidal zones, but appear to move out as
1217 they grow (Harry *et al.* 2011). There is no published data on *R. taylori* habitat use or
1218 movement. As both these species are closely associated with nearshore environments, *C.*
1219 *fitzroyensis* and *R. taylori* may influence nearshore ecosystem dynamics and may be affected
1220 by changes within nearshore areas.

1221 3.1.3 Field Methods

1222

1223 Study species were captured using bottom-set 400-m long-lines, 200-m long 11.45-cm mesh
1224 gillnets, and baited rod and reel. Long-lines were made of 6-mm nylon mainline that was
1225 anchored at both ends. Gangions were composed of 1 m of 4-mm nylon cord and 1 m of 1.5-
1226 mm wire leader. Approximately 50-70 size 14/0 Mustad tuna circle hooks were used per
1227 long-line and baited with butterfly bream (*Nemipterus* sp.), squid (*Loligo* sp.), blue threadfin
1228 (*Eleutheronema tetradactylum*), or mullet (*Mugil cephalus*). Long-lines were set for 45 to 60
1229 minutes, gillnets were set for 15 to 20 minutes. *Rhizoprionodon taylori* and *C. fitzroyensis*
1230 were fitted with V13 and V16 acoustic transmitters (Vemco Ltd., Canada) respectively (Fig.
1231 3.4). Transmitters were implanted into the body cavity (Fig. 3.5a) and the incision was closed
1232 with absorbable sutures. Individuals were measured to the nearest millimetre stretch total
1233 length (STL), sexed, tagged with an individually numbered Rototag in the first dorsal fin, and
1234 released (Fig. 3.5b). Umbilical scar condition, stretch total length (Last and Stevens 2009)
1235 and clasper calcification (males) were used to determine individual age and classify
1236 individuals as immature or mature. Range testing analysis found V13 and V16 transmitters
1237 had a maximum detection range of 525 m and 900 m respectively based on 0.05 probability
1238 of detection (Kessel *et al.* 2013) and emitted a unique code as a pulse series at 69 kHz.
1239 Unique transmitter codes allowed for the identification of individuals.

1240



1241

1242

1243 **Fig. 3.4.** (a) V13 and (b) V16 acoustic transmitters (photo credit Vemco Ltd., Canada).

1244

1245

1246

1247



1248

1249 **Fig. 3.5.** Surgery and processing procedures for sharks, (a) surgical implantation of V13

1250 acoustic transmitter (b) measuring and tagging of a captured shark.

1251

1252 **3.2 Stable Isotope Analysis**

1253

1254 **3.2.1 Study Site**

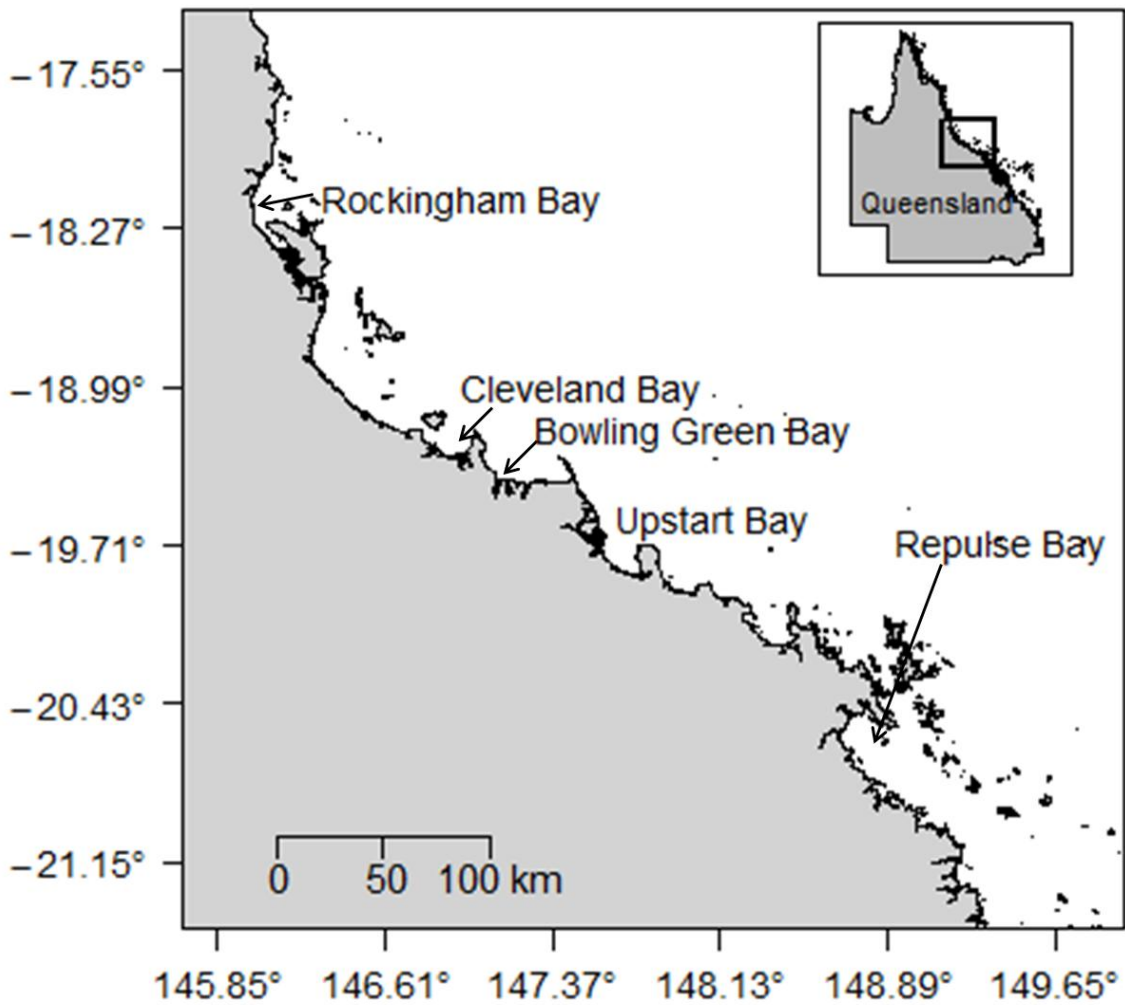
1255

1256 Shark tissue samples were collected from five bays on the northeast coast of Queensland,
1257 Australia between July 2012 and April 2013. The five bays (from south to north) were
1258 Repulse Bay (RE), Upstart Bay (UP), Bowling Green Bay (BG), Cleveland Bay (CB), and
1259 Rockingham Bay (RO) (Fig. 3.6). Cleveland Bay occupies a central location among the
1260 sample bays. Linear distances between adjacent bays ranged from 30 to 150 km.

1261

1262 The primary bottom type in all bays is mud and seagrass beds (GBRMPA 2011). Cleveland
1263 Bay and Repulse Bay also contain small patches of reef. Shorelines are primarily composed
1264 of sandy beaches, mudflats, and mangroves. Cleveland Bay is adjacent to Townsville, a
1265 moderately large north Queensland city. In contrast, the remaining four embayments abut
1266 large expanses of farm land that is primarily used to grow sugarcane. Average annual
1267 freshwater input is variable between bays. On average, Repulse Bay and Rockingham Bay
1268 receive higher volumes of freshwater input annually via rivers than Bowling Green Bay and
1269 Cleveland Bay (Furnas 2003). One of the largest rivers in north Queensland, the Burdekin
1270 River, drains into the coast at the mouth of Upstart Bay (Furnas 2003). However, the output
1271 from Burdekin River generally flows north. As a result, Upstart Bay, which is located south
1272 of Burdekin River, receives relatively little freshwater input. Terrestrial areas adjacent to
1273 Repulse Bay and Rockingham Bay also receive more rain fall (1600-2400 mm) annually than
1274 Cleveland Bay and Bowling Green Bay (1000-1200 mm), with Upstart Bay receiving the
1275 least (800-1000 mm) (Australian Government Bureau of Meteorology).

1276



1277

1278

1279 **Fig. 3.6.** Stable isotope sampling region for *Rhizoprionodon taylori* indicating the five sample

1280 bays. Inset indicates location along the north Queensland coast, Australia.

1281

1282 **3.2.2 Study Species**

1283

1284 Isotope analysis was limited to *R. taylori*. Unfortunately, due to the relative rarity of *C.*
1285 *fitzroyensis* the collection of sufficient samples for isotope analysis for this species was not
1286 possible in the available time frame.

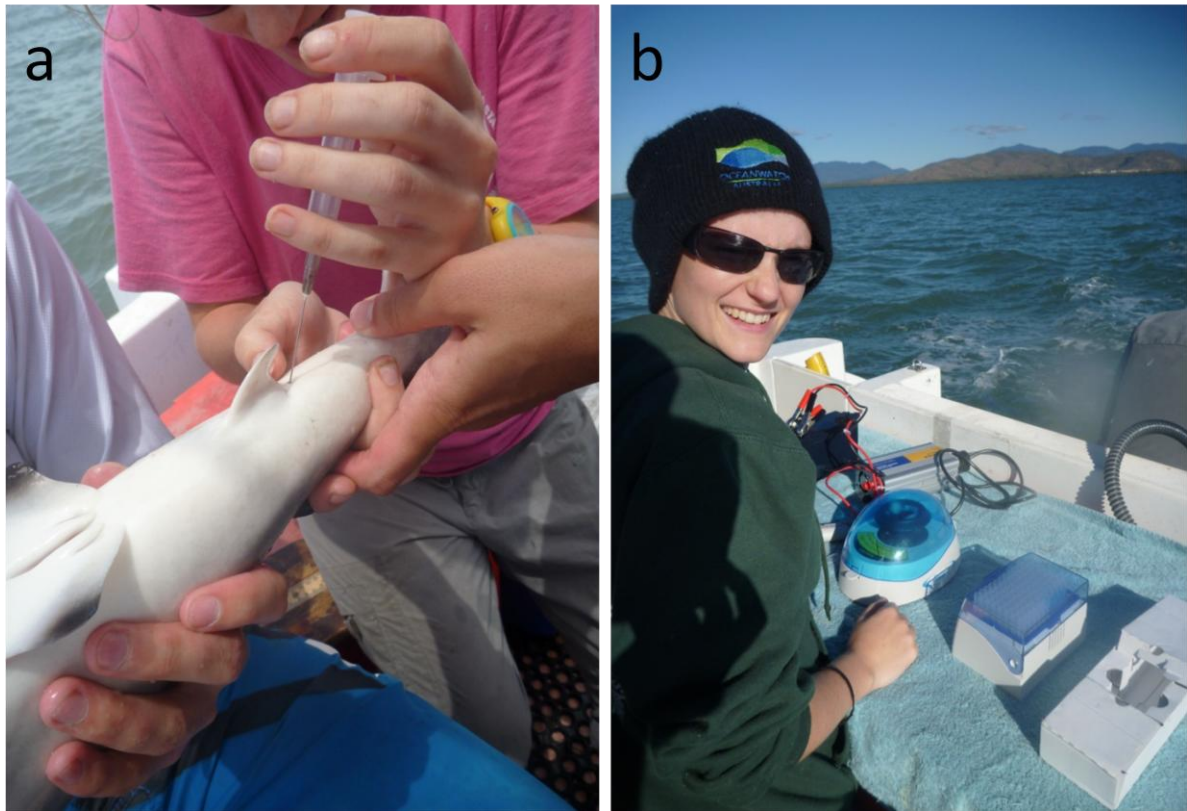
1287

1288 **3.2.3 Field Methods**

1289

1290 Each bay was sampled twice, once in the austral summer (November-March) and once in
1291 austral winter (June -August). Individuals were captured using a combination of bottom-set
1292 400-800 m long-lines and 200-400 m long, 11.45 cm mesh gillnets. Long-lines were
1293 constructed as described in Section 3.1.3. Approximately 50-70 size 14/0 Mustad tuna circle
1294 hooks were used per long-line and baited with butterfly bream (*Nemipterus* sp.), squid (*Loligo*
1295 sp.), blue threadfin (*Eleutheronema tetradactylum*) and mullet (*Mugil cephalus*). Long-lines
1296 and gillnets were set for 45 to 60 minutes. Captured sharks were measured to the nearest
1297 millimetre stretch total length (STL), sexed, and tagged with a uniquely numbered Rototag in
1298 the first dorsal fin. Muscle and plasma tissues were collected for stable isotope analysis and
1299 individuals were released. One cm³ of muscle was sampled from behind the first dorsal fin.
1300 Blood samples were collected using a heparinised needle and syringe from the caudal vein
1301 anterior to the tail. Two ml of blood were collected from each individual (Fig. 3.7a). A
1302 portable centrifuge was used on board the vessel to spin and separate blood samples into
1303 plasma and RBC (red blood cell) components (Fig. 3.7b). Plasma and RBC layers were
1304 pipetted into separate 1.5 ml Eppendorf safe lock microcentrifuge tubes. All shark samples
1305 collected in Cleveland Bay were kept on ice in the field and frozen (-20°C) upon return to the
1306 laboratory. Due to their remote locations, samples collected from the remaining four bays

1307 .



1308

1309 **Fig. 3.7.** (a) Blood extraction from *Rhizoprionodon taylori* and (b) on board centrifuge
1310 equipment.

1311

1312 were kept on ice in the field and stored in a Taylor-Wharton CX100 Dry Shipper (-80°C) until
1313 return to the laboratory where samples were frozen (-20°C).

1314 There is evidence to suggest that juvenile stable isotopes values may incorporate maternal
1315 feeding patterns (Olin et al. 2011). However, previous work has shown that *R. terraenovae*, a
1316 close relative of *R. taylori*, likely replaces the maternal isotope signature with its own dietary
1317 isotope signature by the time its umbilical scar has healed but is still visible (4 to 6 weeks;
1318 Olin et al. 2011). To help ensure maternal isotope values did not affect the isotope values of
1319 captured specimens, *R. taylori* were only sampled if the umbilical scar was no longer visible
1320 (Kinney et al. 2011). Although there is limited information available on how long it takes for
1321 umbilical scars to heal and are no longer be visible, previous work indicates this process may
1322 take approximately one year (Duncan and Holland 2006; Olin et al. 2011).

1323

1324 Data suggest *R. taylori* is a demersal predator, although it could not be conclusively
1325 determined if they forage from benthic and/or pelagic food chains within nearshore areas
1326 (Simpfendorfer, 1998). Therefore, baseline benthic and pelagic $\delta^{13}\text{C}$ ($^{13}\text{C}/^{12}\text{C}$) and $\delta^{15}\text{N}$
1327 ($^{15}\text{N}/^{14}\text{N}$) food web sources were collected from each bay to establish local values. Seagrass
1328 and macroalgae were used to establish benthic food web $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ sources and were
1329 sampled opportunistically from fishing locations in each bay. Plankton was used to establish
1330 pelagic $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ food web sources and were collected using horizontal surface tows
1331 with a 0.85 m long, 300-mm diameter plankton net (53 micron mesh). Plankton samples were
1332 collected from a central location in each bay approximately 5 km from shore. Plankton
1333 samples included zooplankton and some invertebrates. Samples of all plant and plankton
1334 material were kept on ice while in the field and frozen upon return to the laboratory as
1335 described for shark tissues

1336 3.2.4 Sample Preparation and Isotope Analysis

1337

1338 Shark tissue samples were freeze dried and ground into a powder with a mortar and pestle.

1339 Seagrass and macroalgae were thawed, rinsed in distilled H₂O, and cleaned of visible residue

1340 and epiphytes. After cleaning, seagrass and macroalgae were oven dried at 60°C for 48 hours

1341 and ground into a powder. Zooplankton and phytoplankton were not separated to ensure there

1342 was sufficient plankton sample volume for analysis. Plankton samples were filtered through

1343 GF/F Whatman glass micro-fibre filters (0.7 µm pore size) using a vacuum pump (300 mm

1344 Hg). Plankton samples were rinsed with dH₂O during filtration to remove any salt from the

1345 samples. After filtration, large detritus were removed from the filters. Filters were oven dried at

1346 60°C for 24 hours and stored in petri dishes prior to analysis.

1347

1348 Lipids in animal tissues are depleted in ¹³C in comparison to proteins and carbohydrates. The

1349 inclusion of lipids may result in unreliable isotope data where differences in the lipid content

1350 between organisms and tissues may produce more negative δ¹³C (Post *et al.* 2007). To correct

1351 for this, shark tissues and plankton samples underwent lipid extraction using a modified Bligh

1352 & Dyer (1959) method. 1.9 ml of 2:1 chloroform-methanol was combined with the powdered

1353 samples, agitated for 10 seconds and put in a water bath (30°C) for 24 hours. Lipid extracted

1354 samples were removed from the bath, centrifuged for three minutes, and decanted. 1.9 ml of

1355 2:1 chloroform-methanol was added a second time followed by another round of agitating and

1356 centrifuging before the final decant. The tissue pellet that was produced was left in a fume

1357 hood to dry for 48 hours. A separate urea extraction process was not carried out for shark tissue

1358 as previous work has shown that the lipid extraction process also removes soluble urea (Hussey

1359 *et al.* 2012b). 400-600 µg of dried shark muscle, 700-900 µg of dried plasma, 3000-4000 µg of

1360 dried plant material, and 4000-5000 µg of dried plankton were analysed for δ¹³C and δ¹⁵N

1361 using a continuous flow isotope ratio mass spectrometer (IRMS, Finnigan MAT Delta^{plus},
1362 Thermo Finnigan, San Jose, CA, USA) equipped with an elemental analyser (Costech,
1363 Valenica, CA, USA).

1364

1365 Stable isotope ratios were expressed in δ notation as deviations from standards in parts per
1366 thousand (‰) using the following calculation:

1367

$$1368 \quad \delta X = \left[\left(\frac{R_{\text{sample}}}{R_{\text{standard}}} \right) - 1 \right] \times 1000 \quad (3.1)$$

1369

1370 Where X is ¹³C or ¹⁵N, R_{sample} is the ratio (¹³C/¹²C or ¹⁵N/¹⁴N) in the sample, and R_{standard} is the
1371 ratio in the standard. The standard reference for carbon was Pee Dee Belemnite carbonate and
1372 nitrogen was atmospheric N₂. Laboratory and National Institute of Standards and Technology
1373 (NIST) standards were analysed every 12 samples to determine analytical precision. The
1374 analytical precision (standard deviation) for NIST standard 1577c (bovine liver, n =42) and an
1375 internal laboratory standard (tilapia muscle, n = 42) for $\delta^{13}\text{C}$ was 0.07‰ and 0.11‰,
1376 respectively, and for $\delta^{15}\text{N}$ was 0.16‰ and 0.14‰, respectively.

1377

1378 **3.3 Permits and Ethics**

1379

1380 All research was conducted in accordance with James Cook University animal ethics permit
1381 A1566 and Great Barrier Reef (G11/346181.1) and DEEDI (144482) permits for animal
1382 collection.

Chapter 4

Habitat and space use of an abundant nearshore shark, *Rhizoprionodon taylori*

4.1 Introduction

Quantifying shark habitat and space use is essential to understanding a species' relationship with the environment and other species (Clarke *et al.* 2011; Heupel and Simpfendorfer 2008; Knip *et al.* 2011a). Knowledge of habitat and space use can also reveal a great deal about shark ecology and life history. For example, shark activity space size and location have been shown to vary between sexes (Sims *et al.* 2001) and sizes (Goldman and Anderson 1999; Knip *et al.* 2011a). Movement data can also indicate how sharks respond to environmental fluctuations. Previous work has demonstrated factors such as salinity (Ubeda *et al.* 2009), temperature (DiGirolamo *et al.* 2012), bottom type (Morrissey and Gruber 1993a), and prey availability (Sims *et al.* 2006) can influence shark presence, space use, and habitat selection. Movement data has also been used to assess the efficiency of marine protected areas to manage and conserve shark populations (Garla *et al.* 2006a; Knip *et al.* 2012a).

Information on habitat and space use can also be used to determine vulnerability to environmental change and degradation. Species that use a limited array of habitat types may be more vulnerable to environmental change than species that have wider habitat niches (Futuyma and Moreno 1988; Carlton *et al.* 1991; Wilson and Yoshimura 1994). Selective or highly specialised species may not be able to use different habitats if their preferred habitat declines in health or abundance (Colles *et al.* 2009; Clavel *et al.* 2010; Curtis *et al.* 2013). Species with broader habitat niches can more easily use different habitats or locations

1408 (Marvier *et al.* 2004). Therefore, species that can use diverse habitats are more likely to
1409 maintain high levels of fitness in the face of environmental change (Marvier *et al.* 2004;
1410 Richmond *et al.* 2005).

1411

1412 Due to their high productivity, nearshore areas contain key habitats for many sharks and often
1413 function as important foraging and nursery grounds (Beck *et al.* 2001; Heupel *et al.* 2007;
1414 Knip *et al.* 2010; Gutteridge *et al.* 2011). Unfortunately, nearshore habitats also have some of
1415 the highest levels of exposure to sources of anthropogenic influence (Harley *et al.* 2006;
1416 Halpern *et al.* 2008; Bulleri and Chapman 2010; Chin *et al.* 2010). As a result, sharks that use
1417 nearshore areas may be susceptible to population decline (Chin *et al.* 2010). Vulnerability to
1418 decline or localised depletion may increase if the population demonstrates strong site
1419 attachment to specific locations or habitats within nearshore areas. Therefore, data on the
1420 habitat and space use of nearshore sharks is critical to species conservation and management.

1421

1422 In this chapter, I examined the movement patterns of *Rhizoprionodon taylori* in Cleveland
1423 Bay and the species' vulnerability to change. Passive acoustic telemetry was used to quantify
1424 *R. taylori* residency, space use, habitat use, and define whether increased wet season river
1425 discharge affected movement. Results from this chapter will increase our understanding of
1426 how small-bodied sharks use nearshore areas and how *R. taylori* responds to the environment.

1427

1428 **4.2 Data Analysis**

1429

1430 **4.2.1 Residency**

1431

1432 Presence was assessed each day and *R. taylori* were considered present if they were detected
1433 two or more times in the acoustic array in a given day. Residency was determined using a
1434 residency index that calculated the number of days an individual was present in the array as a
1435 proportion of the total days monitored (e.g. Abecasis and Erzini 2008; Werry *et al.* 2014).
1436 The index ranged from 1 to 0, indicating high to low residency, respectively. A two-factor
1437 ANCOVA was used to test for differences in residency between years and sexes with STL as
1438 a covariate.

1439

1440 Individuals were sorted into two groups based on the number of days they were detected in
1441 the array. Individuals that spent more than two weeks in the array were considered resident,
1442 individuals that spent less than two weeks were determined to be transitory. Due to the
1443 potential bias of including individuals with low presence, transitory animals were not
1444 included in space use analysis and some habitat use analysis.

1445

1446 **4.2.2 Space Use**

1447

1448 Individual positions were estimated using the mean position algorithm described by
1449 Simpfendorfer *et al.* (2002) which determines individual centre of activity (COA) locations.
1450 The COA was a weighted mean position for each 30 minute interval the animal was detected
1451 within the acoustic array. COA locations were used to calculate individual monthly activity
1452 space for resident individuals as 50% and 95% kernel utilisation distributions (KUDs) using
1453 the *adehabitatHR* package in R version 3.0 (Calenge 2006). An impassable boundary was
1454 added to the KUD calculations to represent the Cleveland Bay coastline and prevent
1455 overestimation of KUD size. A smoothing parameter of 0.008 was used in all KUD

1456 calculations. A linear mixed effects model was used to determine if there were differences in
1457 50% and 95% KUD size between months with individual as a random factor using the *nlme*
1458 package in R (Pinheiro *et al.* 2013). Linear regression analysis was used to determine if there
1459 was a relationship between 50% and 95% KUD size and animal size (STL). Activity space
1460 overlap between consecutive months was calculated for each *R. taylori* as a percent using the
1461 *adehabitatHR* package in R (Calenge 2006).

1462

1463 **4.2.3 Habitat Selection**

1464

1465 Cleveland Bay was divided into regions based on the distribution of five distinct habitat
1466 types. Receivers were assigned a habitat type based on their location in the bay. Habitat
1467 availability was determined by calculating the proportion of receivers in each habitat.

1468 Proportional habitat use by individuals was determined by comparing the amount of time
1469 spent in a habitat to the total amount of time spent in the array.

1470

1471 The proportion of time spent in each habitat by individuals was compared to habitat
1472 availability using Strauss's (1979) linear selectivity index (L_i) to determine if *R. taylori* were
1473 selecting for or avoiding habitats:

$$1474 \quad L_i = r_i - p_i \quad (4.1)$$

1475 where r_i was the proportion of habitat i used and p_i was the proportion of habitat i available in
1476 the study site. $L_i < 0$ indicated negative selection or avoidance. $L_i > 0$ indicated positive
1477 selection. $L_i = 0$ indicated the habitat was neither positively nor negatively selected for and
1478 was used opportunistically (Strauss 1979). Habitat selection analysis was carried out for both
1479 resident and transitory individuals. The mean of resident individual L_i values was calculated

1480 for each year and each two week time period within each year to assess population habitat
1481 selection over time. A chi-squared goodness-of-fit test was used to determine if the
1482 proportion of time spent in each habitat by the resident population was significantly different
1483 from habitat availability, and hence if mean annual selection was significant.

1484

1485 Spearman's rank correlation coefficient was used to compare habitat selection values of
1486 resident individuals and river discharge rates (m^3s^{-1}). North Queensland is subject to high
1487 rainfall and river discharge during the Austral summer (November to March). Increased river
1488 discharge increases freshwater input into the bay which decreases salinity in waters adjacent
1489 to rivers, especially the southeastern portion of Cleveland Bay (Walker 1981). River
1490 discharge rates from Alligator Creek, a large creek that drains into southeastern Cleveland
1491 Bay, were used to evaluate changes in freshwater input to the bay. River discharge data was
1492 provided by the Queensland Government Department of Natural Resources and Mines.

1493

1494 **4.2.4 Habitat Niche Breadth**

1495

1496 Niche breadth of resident individuals was measured using Hutchinson's niche definition
1497 which only included which habitats a species used and in what proportions (Hutchinson 1957;
1498 Devictor *et al.* 2010). Based on this definition, a modified Freeman-Tukey statistic was used
1499 to calculate population niche breadth (Smith 1982):

$$1500 \quad FT = \sum_{i=1}^R (p_i q_i)^{1/2} \quad (4.2)$$

1501 where q_i was the proportion of habitat i available in the study site, p_i was the proportion of
1502 habitat i used, and R was the total number of habitats available. The output ranged from 1,

1503 which indicated a large niche, to 0, which indicated a narrow niche and a highly specialised
1504 species.

1505

1506 A variation of equation (4.2) was used to calculate individual niche overlap (Arlettaz 1999):

$$1507 \quad FT = \sum_{i=1}^R (p_{ik} p_{jk})^{1/2} \quad (4.3)$$

1508 where p_{ik} and p_{jk} were the proportions of habitat R used by individual i and j respectively. The
1509 output similarly ranged from 1 to 0. A value of 1 indicated complete overlap in habitat niche
1510 breadth between a pair of individuals and 0 indicated no overlap in habitat niche breadth
1511 between a pair of individuals. Where applicable, all data was checked for normality using
1512 normality and homogeneity of variance diagnostics in R, version 3.0 (R Development Core
1513 Team: www.r-project.org) and data were \log_{10} transformed if necessary.

1514

1515 **4.3 Results**

1516

1517 Forty *R. taylori* with acoustic transmitters were released in Cleveland Bay between
1518 September 2011 and November 2012. The majority of individuals ($n = 34$) were captured and
1519 released on the eastern side of Cleveland Bay. Twenty *R. taylori* (7 male, 13 female) were
1520 released in year one of this study (September 2011 to September 2012). Twenty *R. taylori* (7
1521 males, 13 female) were released in year two (September 2012-April 2013). Four *R. taylori*
1522 released in year one and one released in year two died or were not detected following release
1523 and were excluded from analysis. Animal size ranged from 489 to 771 mm STL (mean \pm SE
1524 = 657 ± 21.0) in year one and 485 to 763 mm (mean \pm SE = 659 ± 15.2) STL in year two.
1525 Size ranges indicated that the majority of *R. taylori* were either mature or nearing sexual

1526 maturity (Simpfendorfer 1993). There were no significant differences in sizes between years
1527 (ANOVA, $F_{(1,31)} = 0.0193$, $P > 0.05$), however, females were significantly larger than males
1528 (ANOVA, $F_{(1,31)} = 27.45$, $P < 0.05$).

1529

1530 **4.3.1 Residency**

1531

1532 *Rhizoprionodon taylori* were present in Cleveland Bay for 1-106 days (mean \pm SE = $11.4 \pm$

1533 7.4) in year one and 1-112 days (mean \pm SE = 20.6 ± 6.6) in year two. Two *R. taylori*

1534 released in year one (2 female) and seven released year two (2 male, 5 female) were present

1535 for more than two weeks. The remaining 26 individuals spent less than two weeks in the

1536 array. The residency index was low in both years and ranged from 0.00-0.40 (mean \pm SE =

1537 0.053 ± 0.03) in year one and 0.00-0.56 (mean \pm SE = 0.11 ± 0.04) in year two (Fig. 4.1).

1538 Residency data was not normal and was \log_{10} transformed. Animal size had no effect on *R.*

1539 *taylori* residency (ANCOVA, $F_{(1,27)} = 0.727$, $P > 0.05$). There was a significant difference in

1540 residency between years (ANCOVA, $F_{(1,27)} = 4.48$, $P < 0.05$), but not between sexes

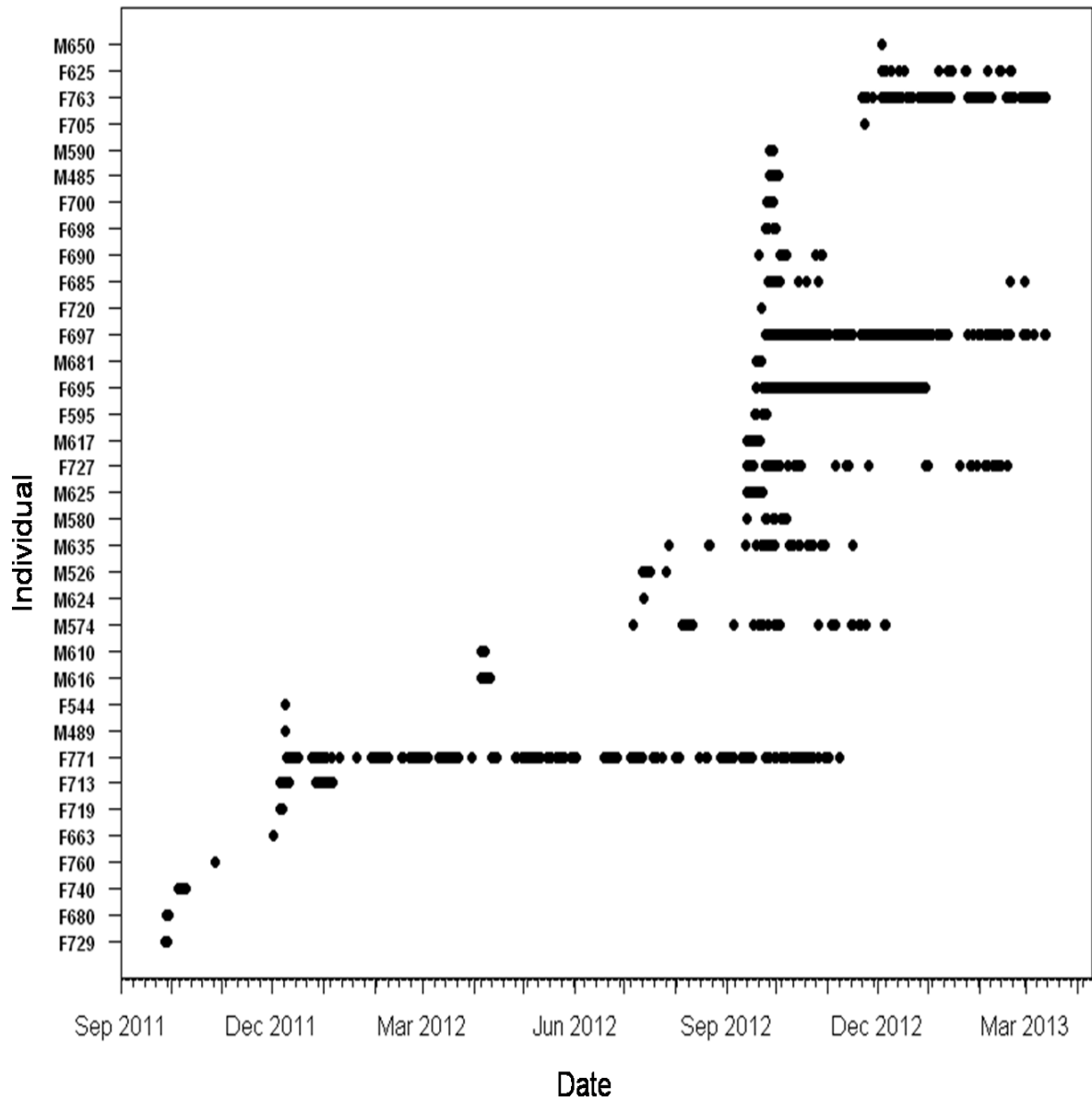
1541 (ANCOVA, $F_{(1,27)} = 0.284$, $P > 0.05$). There was no seasonal pattern in *R. taylori* movement

1542 out of Cleveland Bay. After last detection in Cleveland Bay, seven *R. taylori* (3 male, 4

1543 female) were detected on receivers inside Bowling Green Bay for a maximum of seven

1544 consecutive days (Heupel unpubl. data).

1545



1546

1547 **Fig. 4.1.** Daily presence of *Rhizoprionodon taylori* released with acoustic transmitters in
 1548 Cleveland Bay in 2011- 2013. Individuals are identified by sex and stretch total length (mm).

1549

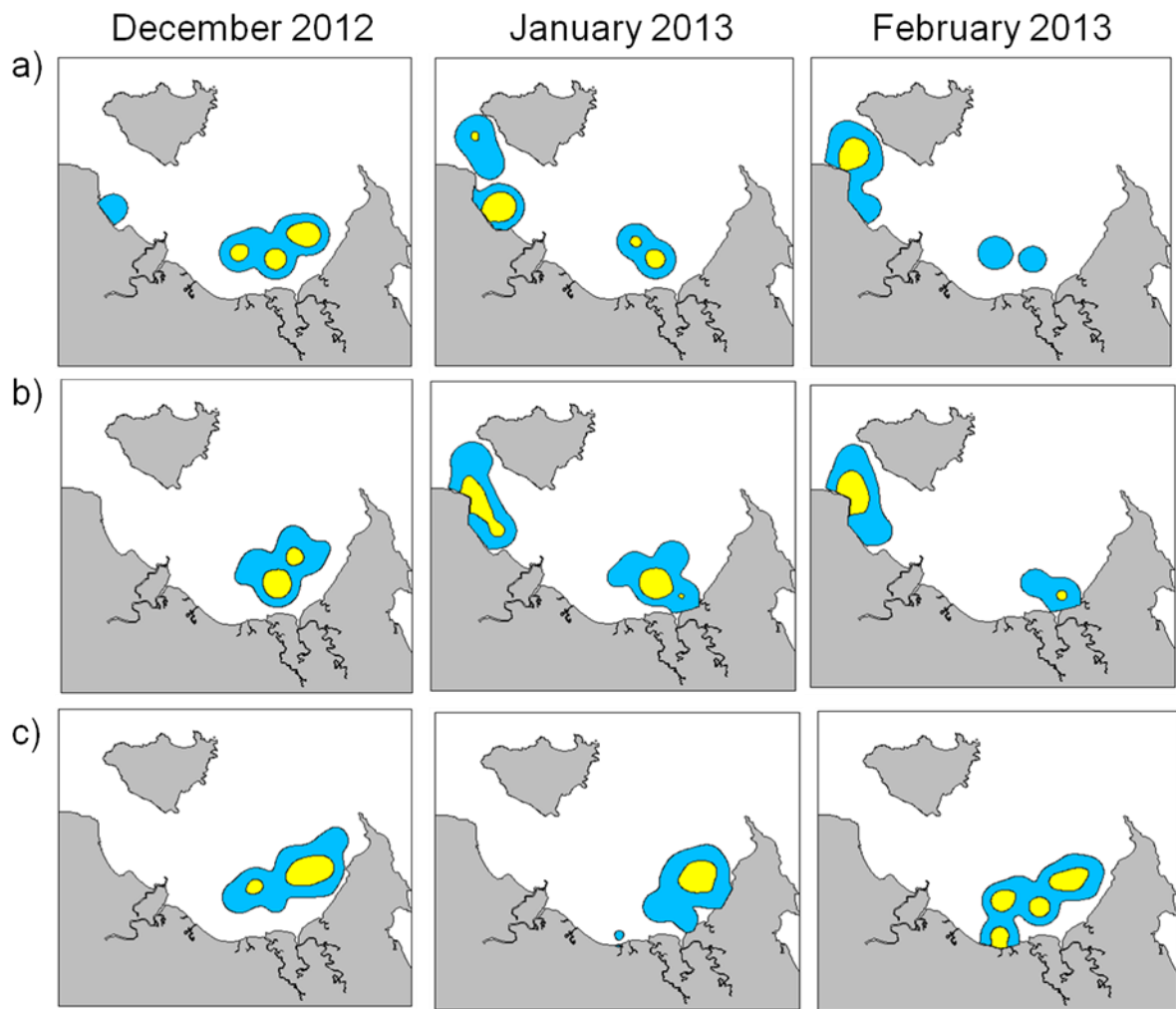
1550 4.3.2 Space Use

1551

1552 Due to the low number of resident individuals, it was not possible to perform population
1553 analysis of the activity space patterns of *R. taylori* in year one. Therefore, all activity space
1554 analysis was based on data from sharks monitored in year two. Individual monthly activity
1555 space of resident individuals ranged between 4.3 and 21.4 km² (mean \pm SE = 11.3 km² \pm
1556 0.90) for 50% KUDs and 21.5 and 80.4 km² (mean \pm SE = 51.0 km² \pm 3.9) for 95% KUDs.
1557 There was no significant difference in KUD size between months for 50% (Linear mixed
1558 effects model, $F_{(6,18)}=0.883$, $P > 0.05$) or 95% (Linear mixed effects model, $F_{(6,18)} = 1.043$, P
1559 > 0.05) KUDs. There was also no relationship between animal size and activity space size for
1560 either 50% (Linear regression, $r^2=0.006$, $F_{(1,23)}=0.136$, $P > 0.05$) or 95% (Linear regression,
1561 $r^2=0.041$, $F_{(1,23)}=0.971$, $P > 0.05$) KUDs.

1562

1563 The majority of *R. taylori* movements were on the eastern side of the bay, specifically in
1564 seagrass habitat. However, 57% of individuals were detected on both sides of the bay.
1565 Individual monthly KUD overlap was highly variable and ranged between 0.0-88.6 % (mean
1566 \pm SE = 34.1 \pm 6.2, $n = 17$) for 50% KUDs and 34.2-92.7% (mean \pm SE = 61.0 \pm 3.8, $n = 17$)
1567 for 95% KUDs. The most distinct shift in *R. taylori* KUD location occurred between months
1568 of low (December 2012) and high river discharge (January and February 2013). Monthly
1569 KUD locations of some individuals (all female) shifted from the southeastern to the
1570 northwestern side of Cleveland Bay between December 2012 and February 2013 (Fig. 4.2a-
1571 b), resulting in low space use overlap for those individuals during that time. However, one
1572 individual remained on the eastern side of Cleveland Bay in January and February 2013 (Fig.
1573 4.2c). Individual monthly KUD overlap was recalculated with months of low and high river



1574

1575 **Fig. 4.2.** *Rhizoprionodon taylori* monthly activity spaces of three individuals (a, b, c) in
 1576 December 2012, January 2013 and February 2013. Each panel shows the 95% (blue fill) and
 1577 50% (yellow fill) kernel utilisation distributions.

1578

1579 discharge excluded to remove their effect on the results. However, secondary results were
1580 similar to the original calculations. Individual monthly KUD overlap again ranged between
1581 0.0-88.57% (mean \pm SE = 30.5 \pm 6.7, $n = 14$) for 50% KUDs and 34.2-92.7% (mean \pm SE =
1582 60.85 \pm 4.5, $n = 14$) for 95% KUDs. There was also no consistent pattern in the degree of
1583 activity space overlap of each individual. Highly variable KUD overlap values indicate
1584 individual *R. taylori* did not use the same core areas between months, regardless of freshwater
1585 input. Most activity space relocations during periods of low river discharge were on the
1586 eastern side of the bay. However, one individual made regular trips between the eastern and
1587 western side of the bay when river discharge was low.

1588

1589 **4.3.3 Habitat Selection and Niche Breadth**

1590

1591 The majority of transient *R. taylori* selected for seagrass habitat (Table 4.1). Of the 26
1592 transient individuals, 11 exclusively selected for seagrass while seven selected for seagrass
1593 and at least one other habitat (intertidal mudflat and/or outer bay mud substrate). Eight
1594 transient *R. taylori* avoided seagrass. Reef was avoided by all transient individuals except for
1595 one adult female.

1596

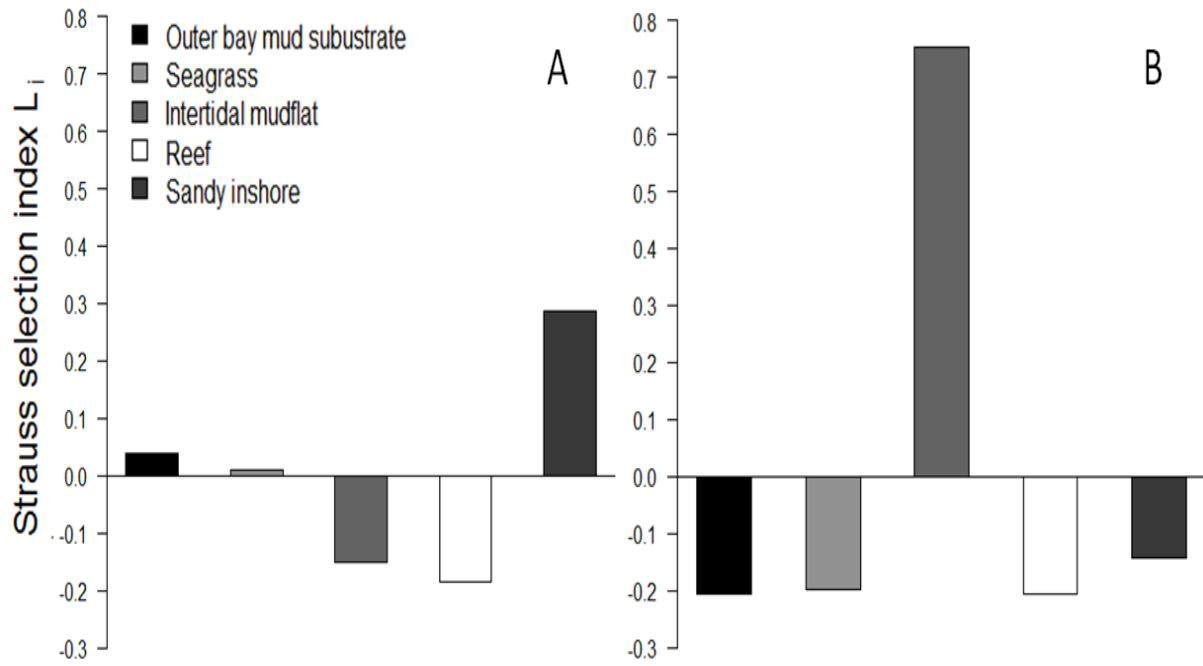
1597 The two resident females monitored in year one had contrasting selection patterns. One
1598 female selected for sandy inshore habitat, outer bay mud substrate, and seagrass (Fig. 4.3a)
1599 while the other only selected for mudflat habitat (Figure 4.3b). Resident individuals in year
1600 two were detected in all five primary habitat types at least once during the monitoring period,
1601 but on average spent the majority of time in seagrass habitat (Table 4.2). Mean individual

1602 **Table 4.1.** Strauss selectivity index values of low residency *Rhizoprionodon taylori* (< two
1603 weeks detected), for each primary habitat in the bay, indicating sex as M (male) or F (female)
1604 and size as stretch total length in mm for each individual.

Sex	Size	Days detected	Seagrass	Outer bay mud substrate	Reef	Sandy inshore	Intertidal mudflat
M	580	8	0.16	-0.01	-0.14	-0.04	0.03
M	620	10	0.54	-0.21	-0.21	-0.14	0.02
M	617	9	0.67	-0.21	-0.21	-0.14	-0.11
F	595	5	0.64	-0.21	-0.21	-0.14	-0.08
M	681	4	0.75	-0.21	-0.21	-0.14	-0.19
F	700	4	0.40	0.15	-0.21	-0.14	-0.21
M	485	6	0.72	-0.21	-0.21	-0.14	-0.16
F	705	1	0.76	-0.21	-0.21	-0.14	-0.21
M	650	1	0.16	0.39	-0.21	-0.14	-0.21
F	698	5	0.69	-0.13	-0.21	-0.14	-0.21
F	720	1	0.36	-0.21	-0.21	-0.14	0.19
M	590	3	0.76	-0.21	-0.21	-0.14	-0.21
F	685	13	0.62	-0.06	-0.21	-0.14	-0.21
F	690	8	-0.11	0.30	-0.20	0.18	-0.17
F	663	1	0.76	-0.21	-0.21	-0.14	-0.21
F	760	1	-0.24	0.13	-0.21	-0.14	0.46
F	719	2	-0.24	0.79	-0.21	-0.14	-0.21
M	616	7	-0.06	-0.21	-0.21	-0.14	0.61
M	610	4	0.73	-0.21	-0.21	-0.14	-0.17
M	489	1	-0.24	-0.21	-0.21	-0.14	0.79
M	624	1	-0.24	0.79	-0.21	-0.14	-0.21
F	544	1	-0.24	-0.21	-0.21	-0.14	0.79
F	740	6	0.16	0.39	-0.21	-0.14	-0.21
F	680	3	0.35	0.21	-0.21	-0.14	-0.21
F	729	3	-0.24	-0.21	0.45	0.20	-0.21

1605

1606



1607

1608

1609 **Fig. 4.3.** Strauss linear selection index values of resident (> 2 weeks) *Rhizoprionodon taylori*

1610 released in Cleveland Bay in year one (September 2011 to September 2012) a) female 574

1611 mm stretch total length (STL) and b) female 713 mm STL.

1612

1613 **Table 4.2.** Mean and standard deviation of time spent in each habitat by high residency (> 2
1614 weeks) *Rhizoprionodon taylori* (n=7) released in year two (September 2012 to April 2013) in
1615 Cleveland Bay, measured as a percent with standard error (SE).

Habitat Type	Mean Time Spent (%) ± SE
Seagrass	48.4 ± 9.6
Outer bay mud substrate	21.2 ± 3.1
Reef	0.1 ± 0.07
Sandy inshore	24.3 ± 8.9
Intertidal mudflat	5.7 ± 2.2

1616

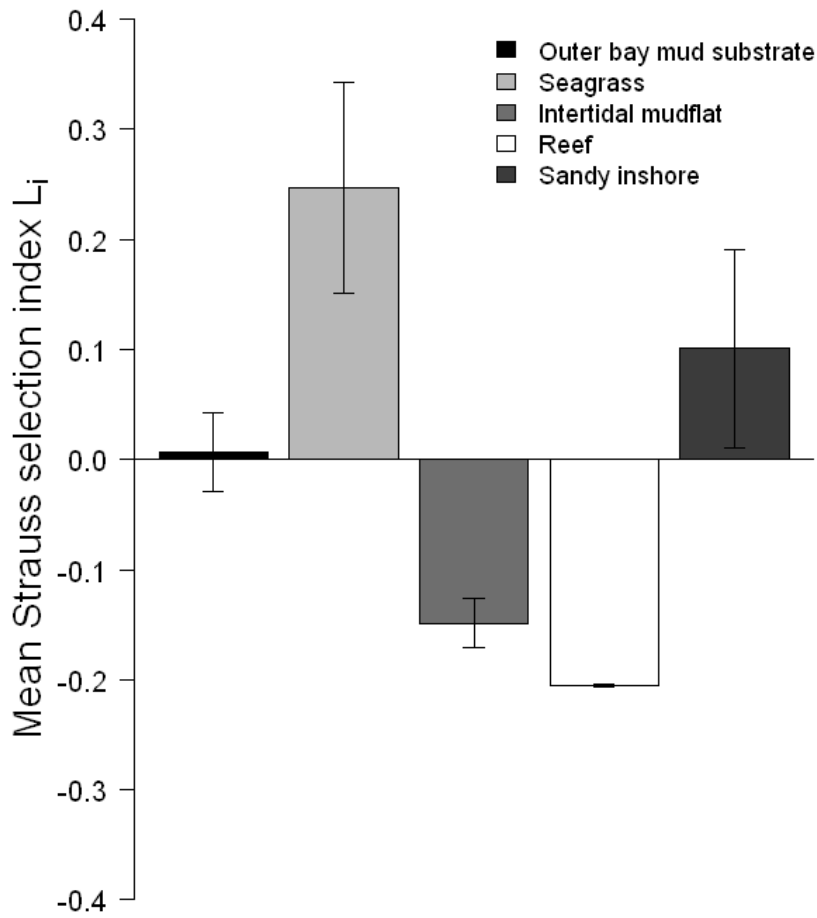
1617

1618 Strauss selection values of resident individuals in year two indicated that on an annual basis
1619 outer bay mud substrate was used opportunistically, reef and mudflat habitats were avoided,
1620 and seagrass and sandy inshore habitat were positively selected (Fig. 4.4). A chi-squared
1621 goodness-of-fit test showed that selection was significant ($X^2_4=63.888, P < 0.05$). Mean
1622 fortnightly selection values revealed that during year two reefs and intertidal mudflats were
1623 consistently avoided while seagrass was consistently favoured, except in January 2013 when
1624 river discharge increased. Selection for sandy inshore substrate and outer bay mud substrate
1625 was more variable (Fig. 4.5).

1626

1627 Selection of outer bay mud substrate of resident individuals fluctuated between low values of
1628 negative and positive selection with individuals generally spending little time in the area (<1
1629 consecutive day). Selection for sandy inshore habitat was also irregular and varied between
1630 negative and positive values over time and among individuals. The majority of resident
1631 individuals in year two did not enter sandy inshore habitat prior to December 2012, except for
1632 one female that made regular excursions into sand habitat for variable periods of time (7-14
1633 consecutive days). However, between December and February 2013, the majority of resident
1634 individuals abruptly selected sandy inshore habitat while avoiding seagrass and this coincided
1635 with increased river discharge. Spearman's rank correlation revealed a strong negative
1636 correlation between sandy inshore and seagrass selection by resident individuals ($r_s = -0.694,$
1637 $N = 14, P < 0.05$). Spearman's rank correlation also revealed a positive relationship between
1638 increased river discharge and selection for sandy inshore habitat ($r_s = 0.305, N = 14, P < 0.05$).
1639 There was a negative relationship between selection for seagrass and increased river
1640 discharge (Spearman's rank correlation, $r_s = -0.308, N = 14, P < 0.05$).

1641

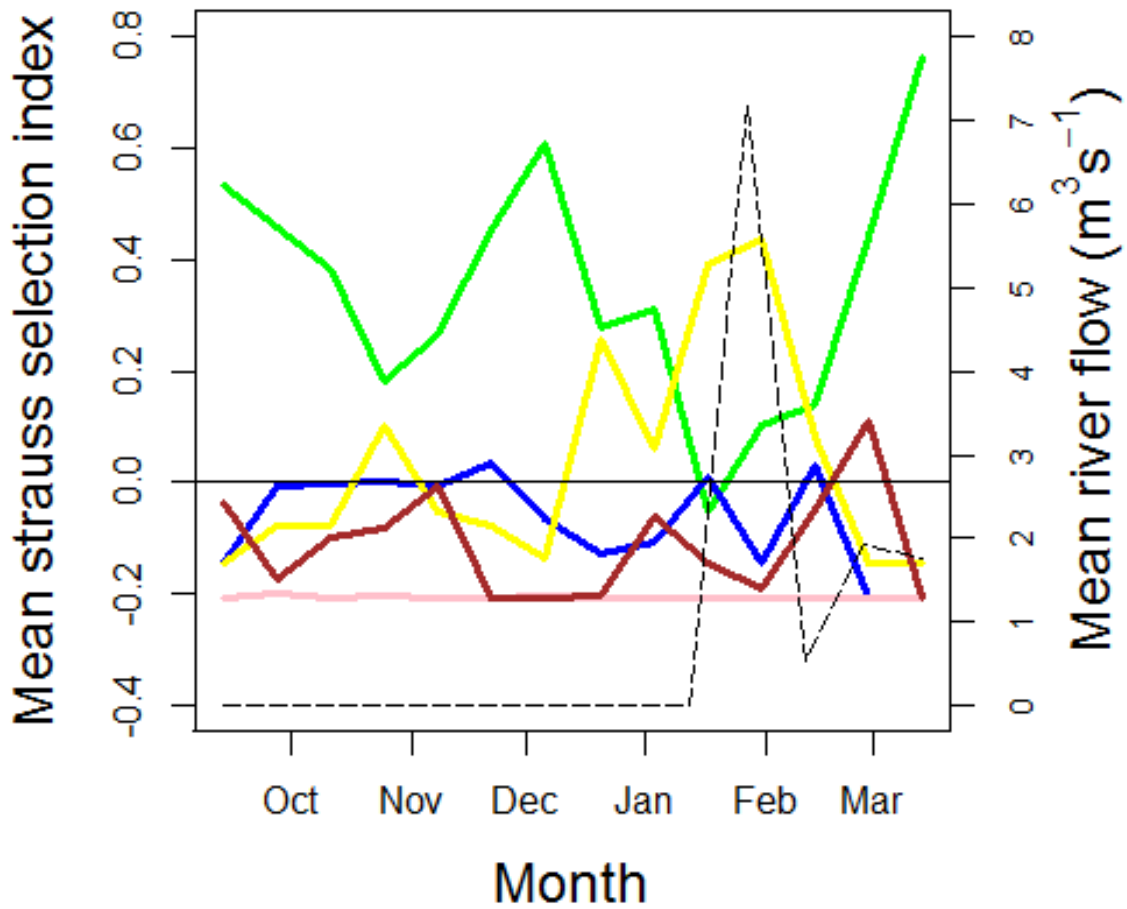


1642

1643 **Fig. 4.4.** Mean Strauss linear selection index values of resident (> 2 weeks) *Rhizoprionodon*
 1644 *taylori* in Cleveland Bay ($n = 7$) between September 2012 to April 2013. Bars indicate
 1645 standard error.

1646

1647



1648

1649

1650 **Fig. 4.5.** *Rhizoprionodon taylori* mean fortnightly individual Strauss linear selection values
 1651 for resident individuals from year two (September 2012-April 2013) for seagrass (green line),
 1652 sand (yellow line), outer bay mud substrate (blue line), intertidal mudflat (brown line), and
 1653 reef (pink line). Mean fortnightly river discharge was measured in m^3s^{-1} (thin black dashed
 1654 line).

1655

1656 Annual mean individual niche breadth for resident *R. taylori* in the second year of this study
1657 was moderately large and ranged from 0.73 to 0.90 (mean \pm SE = 0.79 ± 0.02 , $n = 7$). These
1658 values indicate that individuals had wide habitat niches and therefore had low degrees of
1659 habitat specialisation. Mean individual overlap was also relatively large and ranged from 0.62
1660 to 0.95 (mean \pm SE = 0.83 ± 0.03 , $n = 7$).

1661

1662 **4.4 Discussion**

1663

1664 The movement patterns and low degree of habitat specialisation exhibited by *R. taylori*
1665 contrasts with some other small-bodied sharks that have demonstrated high residency and site
1666 fidelity to nearshore areas, including juvenile lemon sharks *Negaprion brevirostris*
1667 (Morrissey and Gruber 1993b), juvenile blacktip sharks *Carcharhinus limbatus* (Heupel *et al.*
1668 2004), and juvenile pigeye sharks *Carcharhinus amboinensis* (Knip *et al.* 2011a). It has been
1669 suggested that small-bodied sharks, in particular juveniles, are highly resident in shallow
1670 nearshore habitats to avoid large predators (Heupel *et al.* 2007; Knip *et al.* 2010). However,
1671 *R. taylori* residency patterns were consistent with the closely related Atlantic sharpnose shark
1672 *Rhizoprionodon terraenovae*, which has a similar life history to *R. taylori*. Monitored *R.*
1673 *terraenovae* in a northwest Florida bay were only present between 1 and 37 days and were
1674 absent for extended periods of time (Carlson *et al.* 2008). Both *R. taylori* and *R. terraenovae*
1675 are small-bodied species, but are fast growing and highly productive. Therefore, selection for
1676 long-term residency may not provide the same advantages as for juveniles of large-bodied,
1677 slow growing species (Carlson *et al.* 2008; Knip *et al.* 2010). Moreover, given that there are
1678 large-bodied juvenile sharks within Cleveland Bay, *R. taylori* predation risk may be similarly
1679 high inside and outside of nearshore habitats. Therefore, nearshore areas may not provide *R.*

1680 *taylori* with a significantly greater level of protection from predators than offshore habitats.
1681 As a result, there may be limited benefit for *R. taylori* to stay in one bay for long periods.
1682
1683 Low residency values suggest that *R. taylori* move in and out of nearshore areas and may
1684 have used several bays. Detections of *R. taylori* in Bowling Green Bay support the conclusion
1685 that individuals regularly move between different nearshore areas, potentially along the
1686 shoreline. Tag-recapture data in north Queensland also indicate that *R. taylori* move between
1687 neighbouring bays (Simpfendorfer unpubl. data). The use of multiple bays may increase
1688 individual access to prey resources, potentially increasing their foraging success (Knip *et al.*
1689 2010).
1690
1691 *Rhizoprionodon taylori* habitat and space use patterns within Cleveland Bay may have also
1692 been adopted to increase foraging success. Activity space overlap results suggest that most
1693 resident *R. taylori* exhibited roaming movement patterns, typically within seagrass habitat.
1694 However, a few resident individuals ranged throughout the monitoring area. Similar patterns
1695 in space use were observed among bonnethead sharks *Sphyrna tiburo* in a Florida estuary
1696 (Heupel *et al.* 2006). Like *R. taylori*, individual *S. tiburo* used consistent amounts of space
1697 but activity space locations were highly variable. Heupel *et al.* (2006) suggested the roaming
1698 movement patterns exhibited by *S. tiburo* may have been a prey search strategy. By moving
1699 to new areas within the bay, *R. taylori* may increase their prey encounter rates, particularly if
1700 resources have been depleted in a previously occupied area. Seagrass habitat is productive
1701 and usually abundant in small fish and demersal prey (Connolly 1994; Jackson *et al.* 2001).
1702 *Rhizoprionodon taylori* primarily feeds on small teleosts as well as crustaceans and squid
1703 (Simpfendorfer 1998). Therefore, the large abundance of preferred prey in seagrass habitat

1704 may explain why both resident and transient individuals primarily used this habitat. Other
1705 shark species have also selected for seagrass habitat because it functioned as productive
1706 foraging grounds (Heithaus *et al.* 2002; Heithaus *et al.* 2006; Heupel *et al.* 2006). Overall, the
1707 use of multiple bays in combination with roaming movements and the use of seagrass habitat
1708 may combine to provide a more effective foraging strategy than long-term residency in a
1709 single site.

1710

1711 Although seagrass habitat was consistently positively selected by both resident and transitory
1712 individuals, the population's overall low presence and degree of habitat specialisation, as well
1713 as the expansive roaming movement patterns of individuals, suggests *R. taylori* are probably
1714 not dependant on a single habitat. *Rhizoprionodon taylori* also exhibited low levels of
1715 individual specialisation, indicating that overall individuals used similar proportions of the
1716 same habitats as each other. Large individual niches and low levels of individual
1717 specialisation imply the *R. taylori* population in Cleveland Bay is composed of individual
1718 habitat generalists. However, as previously indicated, *R. taylori* did not use all habitats
1719 opportunistically. Avoidance of reef and mudflat by the majority of resident and transitory
1720 individuals suggests these habitats did not fulfil biological requirements or are suboptimal in
1721 some way, such as insufficient prey abundance. Individuals may have also avoided mudflat
1722 habitat because larger sharks (e.g. *C. amboinensis*) that may prey on *R. taylori* utilise this
1723 habitat (Knip *et al.* 2011a). Thus avoidance of these regions may reduce predation risk or
1724 competition with other species using this habitat. Low sample size and residency made it
1725 difficult to determine why *R. taylori* used outer bay mud substrate. Opportunistic use of outer
1726 bay mud substrate by highly resident individuals may be the result of short-term foraging
1727 excursions or a response to short-term environmental fluctuations. Detections in outer bay
1728 mud substrate may have also resulted from *R. taylori* passing through while moving between

1729 sides of the bay and because some outer bay receivers fringe seagrass habitats. Selectivity
1730 indices cannot measure the importance of a habitat, only its relative use, therefore outer bay
1731 mud substrate may serve an important but undefined short-lived purpose.

1732

1733 As discussed earlier, seagrass may provide beneficial foraging habitat, which could explain
1734 the consistent positive selection of this habitat. There is also some evidence to suggest
1735 changes in seagrass availability and abundance may affect *R. taylori* habitat use and presence.
1736 Significantly lower residency in year one occurred at the same time as a substantial decline in
1737 seagrass within Cleveland Bay beginning in 2010. Heavy rainfall in 2010-2011 in
1738 combination with category 5 Cyclone Yasi destroyed much of the seagrass in Cleveland Bay
1739 (Devlin *et al.* 2012; Marshall *et al.* 2011; Mckenzie *et al.* 2012; Seagrass Watch 2013). Low
1740 seagrass abundance may have precipitated a decline in *R. taylori* prey and thus shark
1741 presence. In 2012-2013, seagrass cover increased to levels similar to those prior to the 2010-
1742 2011 wet season (Seagrass Watch 2013). Greater seagrass cover, and potentially higher prey
1743 resource levels, may explain the increased presence of *R. taylori* in year two of the study.
1744 Low seagrass availability during the first year of this study may also explain the contrasting
1745 selection patterns of resident *R. taylori* in year one and year two. The two resident females
1746 monitored in year one may not have selected for seagrass because the habitat was either in
1747 poor condition and/or had decreased prey abundance.

1748

1749 Selection for sandy inshore habitat by resident *R. taylori* in year two may have been
1750 influenced by freshwater input. Some *R. taylori* transitioned from seagrass to sandy inshore
1751 habitat during a large increase in river discharge into Cleveland Bay. Individuals may have
1752 moved to the western side of the bay and used sandy inshore habitats as a secondary foraging

1753 ground in an effort to avoid increased freshwater input into eastern seagrass habitat.
1754 Movement in response to changes in freshwater has been observed in other shark species,
1755 including *C. amboinensis* (Knip *et al.* 2011b), bull sharks *Carcharhinus leucas* (Heupel and
1756 Simpfendorfer 2008), and *S. tiburo* (Ubeda *et al.* 2009). However, not all *R. taylori* moved to
1757 the western side of the bay when freshwater input increased. For that reason, low tolerance to
1758 reduced salinity may not fully explain this movement. It is also possible that increased
1759 freshwater discharge altered prey species distribution and some *R. taylori* may have followed
1760 while others remained to feed on less mobile or euryhaline prey. It is also possible that sandy
1761 inshore habitat selection was related to reproduction. *Rhizoprionodon taylori* give birth
1762 between December and January each year (Simpfendorfer 1992b), but it is not known where
1763 parturition occurs. It may be that some *R. taylori* moved into sandy inshore areas to give birth
1764 before leaving the bay or returning to seagrass habitat. Few transitory individuals selected for
1765 sandy inshore habitat, supporting the suggestion that use of sandy inshore habitat may be less
1766 common or irregular among the *R. taylori* population. Although several possible explanations
1767 exist, more data is needed to better understand movements such as the spatial shift from the
1768 eastern to western part of the bay.

1769

1770 Overall, results of this chapter indicate *R. taylori* is a species that has a low degree of habitat
1771 specialisation and large activity spaces that likely span multiple bays. As a result, *R. taylori*
1772 are probably resilient to localised environmental change and can avoid potentially detrimental
1773 environmental changes by moving to different areas or habitats. However, the presence and
1774 accessibility of seagrass may affect residency and habitat use patterns. Increased coastal
1775 development and the effects of climate change have been highlighted as major ongoing and
1776 future contributors to seagrass decline (Duarte 2002). Severe regional declines in seagrass
1777 abundance as a result of large storms, sea level rise, urban runoff, and development may

1778 decrease local *R. taylori* presence and fitness (Orth *et al.* 2006; Hughes *et al.* 2008; Waycott
1779 *et al.* 2009). Decreased seagrass availability is a potential concern for this species, but further
1780 study of *R. taylori* regional movement and habitat use in other nearshore areas is needed to
1781 assess this possibility. Understanding the dynamics of how small-bodied sharks use nearshore
1782 areas and how environmental change may affect their movement and habitat use will help
1783 define the resilience of coastal shark communities.

1784

Chapter 5

1785

1786 **Regional Movement Patterns of a Small-bodied Shark Revealed** 1787 **by Stable Isotope Analysis**

1787

1788

1789 **5.1 Introduction**

1790

1791 Mobile sharks link distant environments by moving between them and exerting predatory
1792 pressures on local food webs (Weng *et al.* 2007; Weng *et al.* 2008; Chin *et al.* 2013). As a
1793 result, some species connect otherwise separated food webs and ecosystem processes
1794 (Lundberg and Moberg 2003). Fast-growing, small-bodied sharks may be a particularly vital
1795 ecological and energetic link between food webs because they are both predators and prey
1796 items. Therefore, data on the movements of small-bodied sharks can increase understanding of
1797 marine ecosystem function and connectivity. Understanding shark movement patterns may also
1798 help predict how species will respond to environment change. For example, species that can
1799 use multiple, distinct, and potentially distant habitats will be less vulnerable to environmental
1800 change than species that are highly specialised and exhibit high site fidelity (Thomas *et al.*
1801 2004; Araújo *et al.* 2006; Williams *et al.* 2008; Curtis *et al.* 2013).

1802

1803 The regional movement patterns (< 500 km) of most small-bodied sharks are poorly
1804 understood. The current lack of information is due in part to the limitations inherent in methods
1805 previously used to study movement. For example, mark-recapture studies have been used to
1806 investigate the regional and large-scale movements of several smaller coastal sharks, such as
1807 the spottail shark *Carcharhinus sorrah*, the Atlantic sharpnose shark *Rhizoprionodon*
1808 *terraenovae*, and the milk shark *Rhizoprionodon acutus*, but failed to supply large movement

1809 data sets due to a low number of tag returns (Kohler *et al.* 1998; Stevens *et al.* 2000). To have a
1810 better understanding of the regional movements of small-bodied sharks, methods that can
1811 evaluate movement at a regional scale and provide larger, continuous data sets over shorter
1812 timeframes need to be utilized.

1813

1814 Stable isotope analysis (SIA) is a commonly employed technique in ecology that can be used to
1815 define the regional movements of animals (Hobson 2008). Isotope analysis evaluates the ratio
1816 of heavy to light isotopes of carbon ($^{13}\text{C}/^{12}\text{C}$; $\delta^{13}\text{C}$) and nitrogen ($^{15}\text{N}/^{14}\text{N}$; $\delta^{15}\text{N}$). The $\delta^{13}\text{C}$ and
1817 $\delta^{15}\text{N}$ values in an ecosystem vary beginning at the base of the food web (i.e. primary
1818 producers). This variation is the result of different local biogeochemical processes (Boutton
1819 1991). In animal tissue, $\delta^{13}\text{C}$ values increase in small amounts from prey to predator and are
1820 therefore conserved up the food chain (Post 2002). In contrast, $\delta^{15}\text{N}$ tissue values increase from
1821 prey to predator at a significantly greater rate than $\delta^{13}\text{C}$ (Deniro and Epstein 1981; Peterson
1822 and Fry 1987). Therefore, consumers assimilate the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ value of their prey and local
1823 environment with minimal and/or predictable rates of change (Graham *et al.* 2010). As a result,
1824 the isotopic values of resident shark populations should be similar to or a reflection of the
1825 isotopic value of local prey and primary producers (Graham *et al.* 2010). Resident populations
1826 from isotopically distinct habitats or regions should have similarly distinct $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$
1827 values. In contrast, the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of highly mobile, non-resident populations will be a
1828 blend of the different areas or habitats they have foraged in. Thus, the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of
1829 migratory populations should be similar between locations that have been linked via foraging
1830 and movement (Hobson 2008). Therefore, $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values provide a type of intrinsic
1831 geographic tag (Rubenstein and Hobson 2004). Metabolically active tissues, such as liver or
1832 plasma, respond to changes in diet more quickly than tissues with a lower metabolic rate, such
1833 as muscle (Hobson and Clark 1992; Buchheister and Latour 2010). As a result, $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$

1834 from different tissues can reveal if an animal's feeding location has changed over time
1835 (Newsome *et al.* 2009).

1836

1837 There are several benefits from using isotope analysis to assess the long-range movements of
1838 animals. Isotope samples can be processed relatively quickly and each animal sampled will
1839 return data, as opposed to tagged and released animals that have to be recaptured or detected.
1840 Isotope analysis can also provide comparatively long- and short-term assessments of animal
1841 movement (Dalerum and Angerbjörn 2005), depending on the tissue sampled. Therefore, in the
1842 appropriate situation, SIA may be an effective technique to study the regional movements of
1843 small-bodied sharks. However, despite these benefits, isotope-based shark movement studies
1844 are limited and have primarily examined coarse-scale movements between offshore and
1845 nearshore areas (Kerr *et al.* 2006; Abrantes and Barnett 2011; Carlisle *et al.* 2012).

1846

1847 In Chapter 4, acoustic tracking of *R. taylori* in Cleveland Bay found individuals were mostly
1848 transient and several individuals were detected moving into an adjacent bay approximately 30
1849 km south of the primary study site. Based on these findings, I proposed the home range of
1850 individual *R. taylori* encompassed multiple bays. However, due to the constraints of the acoustic
1851 array, it was not possible to determine how far *R. taylori* moved from Cleveland Bay following
1852 release. As a highly abundant coastal mesopredator, *R. taylori* may have a significant influence
1853 on nearshore ecosystem connectivity, function, and diversity.

1854

1855 The aim of this chapter was to evaluate the nearshore regional residency and movements of *R.*
1856 *taylori* using SIA. To determine the physical and temporal extent of *R. taylori* movements,

1857 plasma and muscle $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of captured *R. taylori* were compared to the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$
1858 baselines (established using seagrass, macroalgae and plankton) from five embayments on the
1859 northeast coast of Queensland (Fig. 3.6), including Cleveland Bay. Cleveland Bay occupied a
1860 central location among the sample bays to better understand the movement of *R. taylori* in and
1861 out of Cleveland Bay. The results of this chapter will improve understanding of the regional
1862 residency and movement of small-bodied coastal sharks.

1863

1864 **5.2 Data Analysis**

1865

1866 Bayesian inferences were used for all data analysis in this study because there was concern that
1867 more traditional methods of spatial analysis may not be able to detect some of the subtle
1868 geographic difference in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values. Bayesian analysis is better able to incorporate
1869 uncertainty (due to variability in isotope ratios) and the small and variable sample size in some
1870 locations (Bernardo and Smith 1994; Berger 2006). A Bayesian ANOVA (Gelman 2007) was
1871 used to assess differences between bays in benthic (seagrass and macroalgae) and pelagic
1872 (plankton) $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values. The Bayesian ANOVA used vague, non-informative priors
1873 and was calculated according to the following:

1874 The Likelihood

$$1875 y_{ij} \sim \text{Normal}(\mu + \alpha_i, \sigma^2) \quad (5.1)$$

1876 The Priors

$$1877 \mu \sim \text{Normal}(0, 10^{-6}) \quad (5.2a)$$

$$1878 \alpha_i \sim \text{Normal}(0, 10^{-6}) \quad (5.2b)$$

1879 Where μ was the overall mean and α_i was the effect due to the i -th sample bay. Differences
 1880 between locations were considered significant if the 95% credibility intervals of posterior
 1881 draws did not overlap.

1882

1883 Linear Bayesian correlation analysis (Gelman 2007; McCarthy 2007) was used to determine if
 1884 there was a correlation between benthic and pelagic $\delta^{13}\text{C}$ or $\delta^{15}\text{N}$ values based on geographic
 1885 location. The results were used to establish if there was a consistent geographic pattern in
 1886 benthic and pelagic $\delta^{13}\text{C}$ or $\delta^{15}\text{N}$ that could be compared to *R. taylori* $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values to
 1887 assess regional movement. Correlation analysis was calculated according to the following:

1888 The Likelihood

$$1889 C_i \sim \text{MultivariateNormal}([\mu_1, \mu_2], \Sigma) \quad (5.3)$$

1890 Where $C_i = (C_{i,1}, C_{i,2})$

1891 The Priors

$$1892 \mu_1, \mu_2 \sim \text{Normal}(0, 10^3) \quad (5.4a)$$

$$1893 \sigma_1, \sigma_2 \sim \text{Normal}(0, 10^3) \quad (5.4b)$$

$$1894 \rho \sim \text{Uniform}(-1, 1) \quad (5.4c)$$

$$1895 \Sigma = \begin{pmatrix} \sigma_1^2 & \rho\sigma_1\sigma_2 \\ \rho\sigma_1\sigma_2 & \sigma_2^2 \end{pmatrix} \quad (5.5)$$

1896 Where $C_{i,1}$ and $C_{i,2}$ were the $\delta^{13}\text{C}$ or $\delta^{15}\text{N}$ values of group one and two respectively, μ_1 and μ_2
 1897 were the means of group one and group two, Σ was the covariance matrix and ρ defined the
 1898 correlation parameter. ρ ranged from -1 to 1, indicating the strength and direction of the

1899 correlation. Correlations were considered significant if the 95% credibility intervals of the
1900 posterior draws did not overlap with 0.

1901

1902 There were no estimates for the isotopic incorporation rates of *R. taylori*. Therefore tissue
1903 turnover rates were approximated based on data available for other elasmobranchs. Logan and
1904 Lutcavage (2010) found that complete isotopic $\delta^{13}\text{C}$ turnover for captive juvenile sandbar
1905 sharks *Carcharhinus plumbeus* was > 300 days in whole blood and > 500 days in muscle. Kim
1906 *et al.* (2012) found complete isotopic $\delta^{13}\text{C}$ turnover for captive leopard sharks *Triakis*
1907 *semifasciata* Girard 1855 was somewhat longer with approximately 300 days in plasma and >
1908 700 days in muscle. However, Malpica-Cruz *et al.* (2012) found that the isotopic turnover rates
1909 of small, faster-growing captive *T. semifasciata* were faster than turnover rates of the larger,
1910 more slowly growing individuals. Moreover, Olin *et al.* 2011 found that *R. terraenovae*, a close
1911 relative of *R. taylori*, had high isotopic turnover rates due to the species' high growth rate.
1912 Previous studies of other taxa have also demonstrated that small body size, fast growth rate,
1913 and high metabolic rate increase $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ turnover (Trueman *et al.* 2005; Tarboush *et al.*
1914 2006; Carleton and Del Rio, 2010; Weidel *et al.* 2011). Due to their relatively small size and
1915 fast growth rate, *R. taylori* likely have faster $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ turnover rates than slow growing,
1916 larger-bodied sharks (Olin *et al.* 2011). Therefore, based on estimates of previous work, *R.*
1917 *taylori* plasma isotopic $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ turnover was estimated to take approximately 6 months
1918 (~180 days) and muscle was estimated to take approximately one year (~365 days) (Olin *et al.*
1919 2011; Malpica-Cruz *et al.* 2012).

1920

1921 A 2-factor Bayesian ANOVA (Gelman 2007) was used to test for differences in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$
1922 between bays and sexes in muscle and plasma. The 2-way Bayesian ANOVA was calculated
1923 according to the formulations:

1924 The Likelihood:

$$1925 \quad y_{ijk} \sim \text{Normal}(\mu + \alpha_i + \beta_j + \gamma_{ij}, \sigma^2) \quad (5.6)$$

1926 The Priors

$$1927 \quad \mu \sim \text{Normal}(0, 10^{-6}) \quad (5.7a)$$

$$1928 \quad \alpha_i \sim \text{Normal}(0, 10^{-6}) \quad (5.7b)$$

$$1929 \quad \beta_j \sim \text{Normal}(0, 10^{-6}) \quad (5.7c)$$

$$1930 \quad \gamma_{ij} \sim \text{Normal}(0, 10^{-6}) \quad (5.7d)$$

1931

1932 Where β_j was the effect due to sex, and γ_{ij} was the effect due to the interaction between the i -th
1933 bay and sex.

1934

1935 A 3-way ANCOVA was considered to examine the effect of size (STL), sex, and sample bay
1936 on shark tissue $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, however, preliminary analysis showed the effect of size was
1937 highly variable between bays. Therefore, a series of linear Bayesian regressions were used to
1938 determine if there was a relationship between muscle and plasma $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ and animal
1939 size for each bay. Regression analysis used vague, non-informative priors and was calculated
1940 according to the following:

1941 The Likelihood

1942 $y_{ij} \sim \text{Normal}(\mu + S_i, \sigma^2)$ (5.8)

1943 The Priors

1944 $\mu \sim \text{Normal}(0, 10^{-6})$ (5.9a)

1945 $S_i \sim \text{Normal}(0, 10^{-6})$ (5.9b)

1946 Where S_i was the effect due to animal size. Relationships between size and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ were
1947 considered significant if the 95% credibility intervals of the posterior draws did not overlap
1948 with 0.

1949 Linear Bayesian correlation analysis was used to determine if there was a correlation between
1950 benthic and pelagic, and shark tissue $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values based on location. All posterior
1951 draws were built using three Markov chains with 10000 iterations per chain and a thinning
1952 interval of 10. Chain mixing trace plots and autocorrelation values were used to assess each
1953 applied version of the models. All Bayesian models were fitted using the package R2jags (Su
1954 and Yajima 2014) in R version 3.0.2 (R Development Core Team: www.r-project.org) and
1955 JAGS, version 3.4.0 (Plummer 2003).

1956

1957 **5.3 Results**

1958

1959 **5.3.1 Study Site $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$**

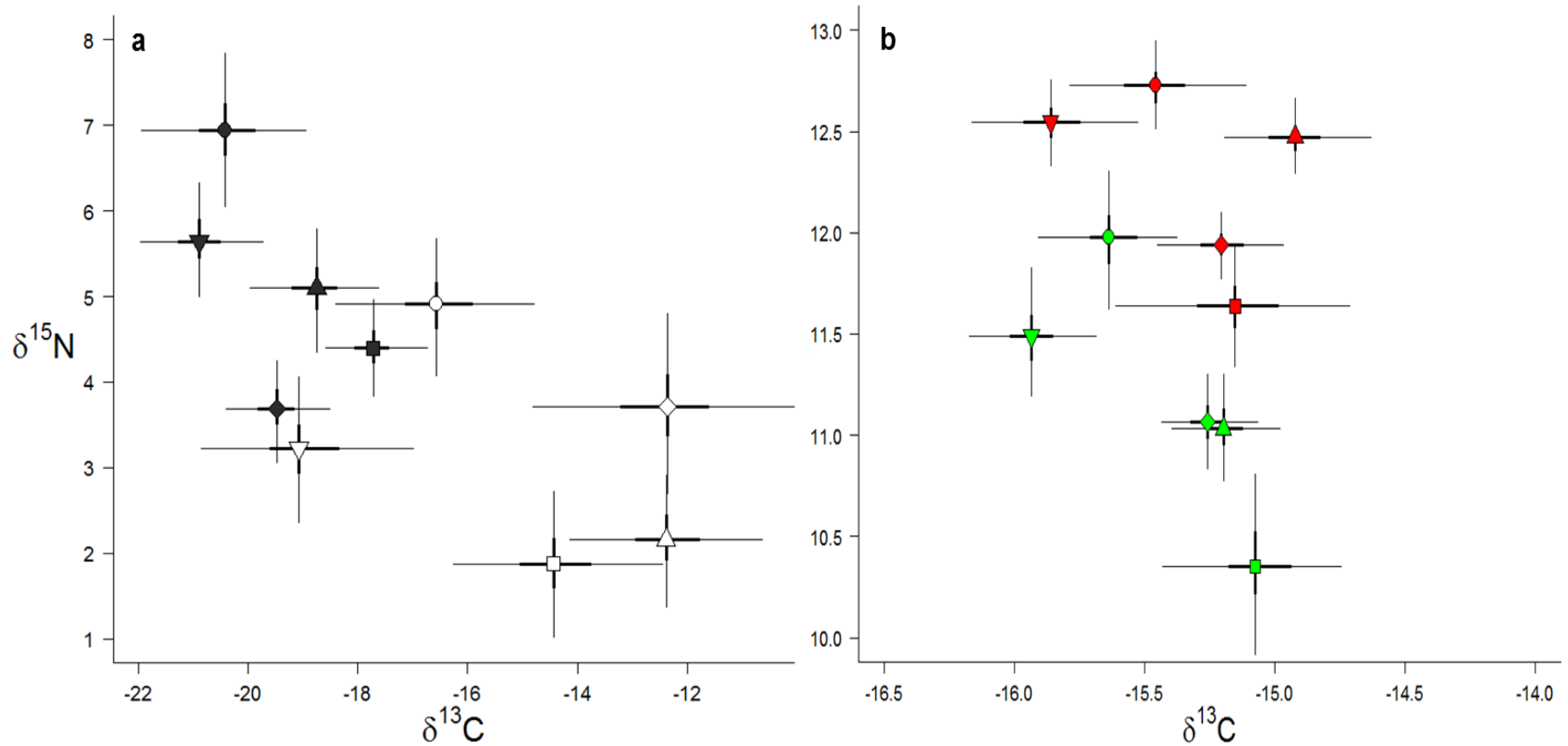
1960

1961 Forty-seven pelagic (plankton) and 55 benthic (seagrass and macroalgae) samples were
1962 collected from the five bays (Table 5.1). Benthic $\delta^{13}\text{C}$ values were higher than pelagic values in
1963 all bays (Table 5.1; Fig. 5.1). Across all sample bays, benthic and pelagic $\delta^{13}\text{C}$ values ranged
1964 from -23.2 to -8.4 and -23.3 to -14.6 respectively. Sample bay accounted for 45.3% and 45.4%

1965 **Table 5.1** Sample size (n) and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ range (mean \pm SD) of benthic and pelagic sources from each location, Repulse Bay (RE), Upstart
 1966 Bay (UP), Bowling Green Bay (BG), Cleveland Bay (CB), and Rockingham Bay (RO).

Location	Benthic (n)	Pelagic (n)	Benthic $\delta^{13}\text{C}$	Benthic $\delta^{15}\text{N}$	Pelagic $\delta^{13}\text{C}$	Pelagic $\delta^{15}\text{N}$
RE	12	5	-19.6 - -12.1 (-16.1 \pm 2.3)	2.9-6.8 (4.9 \pm 1.3)	-20.8 - -19.9 (-20.4 \pm 0.3)	6.7-7.6 (6.9 \pm 0.4)
UP	11	13	-20.5 - -10.3 (-14.4 \pm 4.0)	0.62-4.5 (2.1 \pm 1.7)	-20.52 - - 14.63 (-17.7 \pm 2.6)	3.7-5.0 (4.4 \pm 0.5)
BG	7	12	-19.3- -9.5 (-12.4 \pm 4.3)	2.4-5.7 (3.7 \pm 1.4)	-20.1- -18.7 (-19.5 \pm 0.4)	1.22-5.9 (3.7 \pm 1.8)
CB	13	8	-17.2- -8.4 (-12.4 \pm 2.2)	1.3-3.6 (2.2 \pm 0.8)	-19.4- - 18.2(-18.8 \pm 0.5)	4.7-5.7 (5.1 \pm 0.3)
RO	11	9	-23.2- -15.2 (-19.1 \pm 2.8)	1.3-4.9 (3.7 \pm 1.0)	-23.3- -18.7 (-21.0 \pm 2.0)	5.2-6.2 (5.6 \pm 0.3)

1967



1968

1969 **Fig. 5.1.** Median $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values from Bayesian ANOVA of (a) benthic (white), pelagic (dark grey), (b) *Rhizoprionodon taylori* muscle
1970 (red), and plasma (green) samples in Repulse Bay (●), Upstart Bay (■), Bowling Green Bay (◆), Cleveland Bay (▲) and Rockingham Bay (▼),
1971 black lines show 50% and 95% credibility intervals of posterior draws.

1972 of the estimated variance components (% standard deviation) in benthic and pelagic $\delta^{13}\text{C}$
1973 values respectively. Benthic $\delta^{13}\text{C}$ values from UP, BG and CB were significantly higher than
1974 the benthic $\delta^{13}\text{C}$ values from RO (Fig. 5.1). Benthic $\delta^{13}\text{C}$ values in BG and CB were also
1975 significantly higher than the $\delta^{13}\text{C}$ values from RE. Although the absolute $\delta^{13}\text{C}$ values were
1976 different, pelagic samples exhibited similar geographic patterns in relative $\delta^{13}\text{C}$ values
1977 compared to benthic samples. Pelagic $\delta^{13}\text{C}$ values from UP and CB were significantly higher
1978 than the $\delta^{13}\text{C}$ values from RO. Pelagic $\delta^{13}\text{C}$ values from UP were also significantly higher than
1979 the $\delta^{13}\text{C}$ values from RE. Linear Bayesian correlation analysis indicated there was no
1980 significant correlation between benthic and pelagic $\delta^{13}\text{C}$ based on location (median, 95%
1981 credibility intervals = 0.50, -0.406-0.999).

1982

1983 Benthic $\delta^{15}\text{N}$ values were lower than pelagic $\delta^{15}\text{N}$ values in all bays, most likely because
1984 pelagic samples contained some zooplankton. Benthic and pelagic $\delta^{15}\text{N}$ values ranged from
1985 and 0.62 to 6.8 and 1.2 to 7.6 respectively. Sample bay accounted for 46.3% and 49.0% of the
1986 estimated variance components (% standard deviation) in benthic and pelagic $\delta^{15}\text{N}$ values
1987 respectively. Benthic $\delta^{15}\text{N}$ in RE was significantly higher than the $\delta^{15}\text{N}$ values from BG and
1988 CB. Benthic RO $\delta^{15}\text{N}$ was also higher than CB and UP $\delta^{15}\text{N}$, but the difference was not
1989 significant. Pelagic $\delta^{15}\text{N}$ values in RE were significantly higher than pelagic $\delta^{15}\text{N}$ from UP,
1990 BG and CB. Pelagic RO $\delta^{15}\text{N}$ was also significantly higher than $\delta^{15}\text{N}$ values from UP and BG.
1991 Linear Bayesian correlation analysis indicated there was no significant correlation between
1992 benthic and pelagic $\delta^{15}\text{N}$ based on location (median, 95% credibility intervals = 0.39, -0.57 -
1993 0.98).

1994

1995 Despite the lack of significant geographic correlations across sample locations, there was
1996 evidence of consistent geographic patterns in the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in primary producers.
1997 Specifically, benthic and pelagic $\delta^{13}\text{C}$ from UP, BG, and CB was higher than $\delta^{13}\text{C}$ from RO
1998 and RE. Repulse Bay had the highest $\delta^{15}\text{N}$ values compared to any other bay for both benthic
1999 and pelagic sources. RO also had relatively high benthic and pelagic $\delta^{15}\text{N}$ compared to other
2000 bays. Moreover, sample location accounted for a large component of the estimated variance,
2001 suggesting location was a strong determinant of benthic and pelagic $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$. Therefore,
2002 the results indicated there was a relatively consistent geographic trend in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ that
2003 could be used to assess *R. taylori* residency and movement between bays.

2004

2005 **5.3.2 Shark tissue $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$**

2006

2007 One hundred and forty six *R. taylori* (30 male, 116 female) were sampled from the five study
2008 bays from 2012 to 2013 (Table 5.2). Across all sample bays, *R. taylori* size ranged from 415 to
2009 780 mm STL (mean \pm SD = 663 \pm 66). Muscle $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ ranged from -18.1 to -12.8 and
2010 10.6 to 13.8 respectively. Plasma $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ ranged from -16.8 to -13.7 and 8.3 to 12.7
2011 respectively. An insufficient number of males were captured in RE and UP to investigate the
2012 effect of sex on $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in these bays (Table 5.2).

2013

2014 Stretched total length had a small, inconsistent, and mostly insignificant influence on *R. taylori*
2015 $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$. Linear Bayesian regression analysis showed only BG muscle $\delta^{13}\text{C}$ (median,
2016 95% credibility intervals; 0.006, 0.004 - 0.008), BG plasma $\delta^{13}\text{C}$ (median, 95% credibility
2017 intervals; 0.003, 0.0006 - 0.006 $\delta^{13}\text{C}$), CB muscle $\delta^{13}\text{C}$ (median, 95% credibility intervals;
2018 0.005, 0.0007 - 0.009), CB plasma $\delta^{13}\text{C}$ (median 95% credibility intervals; 0.003, 0.0008 -
2019 0.005), and RE muscle $\delta^{13}\text{C}$ (median 95% credibility intervals; 0.006, 0.001 - 0.01) had a

2020 **Table 5.2.** *Rhizoprionodon taylori* male (M) and female (F) sample size, combined stretch total length in mm (STL mm), and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$
 2021 tissue range (mean \pm SD) from Repulse Bay (RE), Upstart Bay (UP), Bowling Green Bay (BG), Cleveland Bay (CB), and Rockingham Bay
 2022 (RO).

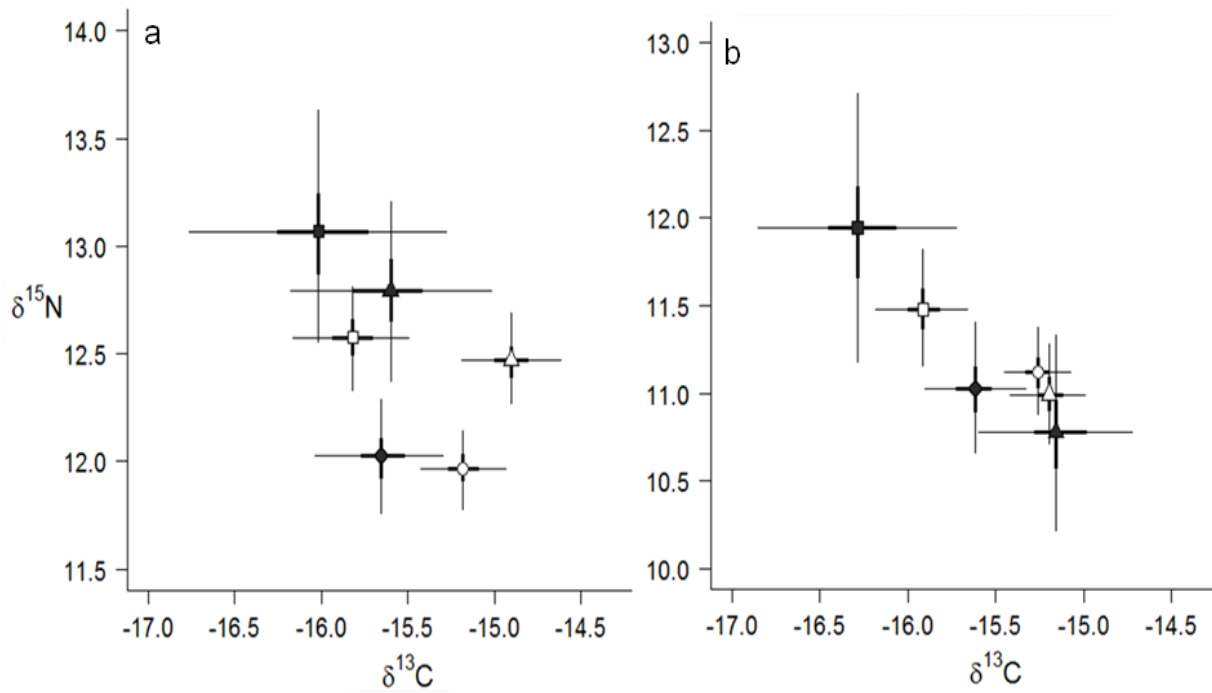
Bay	M	F	STL (mm)	Muscle $\delta^{13}\text{C}$	Muscle $\delta^{15}\text{N}$	Plasma $\delta^{13}\text{C}$	Plasma $\delta^{15}\text{N}$
RE	1	20	595-755 (699 \pm 41)	-16.6 - -14.5 (-15.5 \pm 0.5)	12.0-13.4 (12.7 \pm 0.4)	-16.7 - -14.7 (-15.6 \pm 0.5)	10.2-12.7(11.9 \pm 0.8)
UP	0	11	674-780 (714 \pm 35)	-16.0 - -14.2 (-15.2 \pm 0.5)	11.1-12.3 (11.6 \pm 0.5)	-15.9 - -14.5 (-15.1 \pm 0.5)	9.19-11.4 (10.4 \pm 0.8)
BG	18	37	462-753 (641 \pm 68)	-16.6 - -12.8 (-15.3 \pm 0.7)	10.6-13.2 (12.0 \pm 0.6)	-16.7- -13.7 (-15.4 \pm 0.6)	9.6-12.0 (11.1 \pm 0.7)
CB	7	28	415-744 (650 \pm 80)	-18.1 - -13.3 (15.04 \pm 1.0)	10.7-13.5 (12.5 \pm 0.6)	-16.5 - -14.2 (-15.2 \pm 0.5)	8.3-12.3 (11.0 \pm 0.9)
RO	4	20	625-755 (678 \pm 38)	-17.0 - -14.5 (-15.8 \pm 0.6)	11.6-13.8 (12.7 \pm 0.5)	-16.8- -14.5 (-16.0 \pm 0.5)	10.0-12.5 (11.6 \pm 0.6)

2023 significantly positive relationship with STL. Only BG plasma $\delta^{15}\text{N}$ was significantly correlated
2024 with *R. taylori* STL (median 95% credibility intervals; 0.003, 0.0 - 0.005). Sex also had an
2025 inconsistent effect on *R. taylori* $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$. Female muscle and plasma $\delta^{13}\text{C}$ from BG and
2026 muscle from CB was significantly higher than male $\delta^{13}\text{C}$ (Fig. 5.2). However, there was no
2027 significant difference in $\delta^{13}\text{C}$ between sexes in RO or plasma tissue from CB. There was no
2028 significant difference in $\delta^{15}\text{N}$ between sexes for muscle or plasma tissue (Fig. 5.2). As a result
2029 of the inconsistent effect of sex on $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ and the small, uneven sampling of males
2030 between locations, males were excluded from between bay $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ comparisons.

2031 When compared to the primary produces, female *R. taylori* muscle and plasma $\delta^{13}\text{C}$ values fell
2032 within range of the combined benthic and pelagic $\delta^{13}\text{C}$ values of their respective capture
2033 locations. Sample bay accounted for 33.0% and 38.2% of the estimated variance components
2034 (% standard deviation) in muscle and plasma $\delta^{13}\text{C}$, respectively. The results of the Bayesian
2035 ANOVA showed female *R. taylori* muscle $\delta^{13}\text{C}$ from UP, BG, and CB was significantly higher
2036 than muscle $\delta^{13}\text{C}$ values from RO (Fig. 5.1b). Muscle $\delta^{13}\text{C}$ from CB was also significantly
2037 higher than muscle $\delta^{13}\text{C}$ from RE. Female *R. taylori* plasma $\delta^{13}\text{C}$ in UP, BG, and CB was
2038 significantly higher than plasma $\delta^{13}\text{C}$ from RO and RE (Fig. 5.1b). It was not possible to
2039 differentiate between the $\delta^{13}\text{C}$ values of individuals captured in UP, BG and CB for either
2040 muscle or plasma. These results show there was a high degree of similarity in *R. taylori* $\delta^{13}\text{C}$
2041 between adjacent bays (i.e. those within 100 km), while *R. taylori* from more distant bays (>
2042 100 km separation) had less similar $\delta^{13}\text{C}$ values. Overall, differences in *R. taylori* $\delta^{13}\text{C}$ between
2043 locations were more pronounced in plasma than muscle.

2044

2045



2046

2047 **Fig. 5.2.** Median $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of Bayesian ANOVA of female (white) and male
2048 (dark grey) *Rhizoprionodon taylori* for muscle (a) and plasma (b) tissue in Bowling Green Bay
2049 (●), Cleveland Bay (▲) and Rockingham Bay (■). Black lines show 50% and 95% credibility
2050 intervals of posterior draws.

2051

2052 The results of the Bayesian ANOVA showed sample bay accounted for 45.6% and 45.1% of
2053 the estimated variance components (% standard deviation) in muscle and plasma $\delta^{15}\text{N}$,
2054 respectively. Female *R. taylori* muscle $\delta^{15}\text{N}$ from RE and RO was significantly higher than
2055 muscle $\delta^{15}\text{N}$ from BG and UP (Fig. 5.1b). Female *R. taylori* plasma $\delta^{15}\text{N}$ from RE was also
2056 significantly higher than plasma $\delta^{15}\text{N}$ in UP, BG and CB. Plasma $\delta^{15}\text{N}$ from RO was
2057 significantly higher than plasma $\delta^{15}\text{N}$ in UP. Muscle and plasma $\delta^{15}\text{N}$ values from UP, BG and
2058 CB closely overlapped. The differences in *R. taylori* $\delta^{13}\text{N}$ between sample bays were more
2059 pronounced in plasma than muscle. Similar to the $\delta^{13}\text{C}$ analysis, $\delta^{15}\text{N}$ analysis showed that *R.*
2060 *taylori* from more distant bays (> 100 km separation) had more distinct $\delta^{15}\text{N}$ values.

2061

2062 Linear Bayesian correlation analysis indicated significant positive correlation between muscle
2063 and benthic $\delta^{13}\text{C}$ values based on location (Table 5.3). However, correlation between muscle
2064 and pelagic $\delta^{13}\text{C}$ was not significant. Plasma $\delta^{13}\text{C}$ was significantly positively correlated with
2065 pelagic $\delta^{13}\text{C}$ values; however correlation between plasma and benthic $\delta^{13}\text{C}$ was not significant.
2066 None of the $\delta^{15}\text{N}$ tissue-primary producer geographic correlations were significant. Although
2067 not all correlations between tissues and study site $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ based on location were
2068 significant, the geographic patterns in female *R. taylori* $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ were similar to the
2069 geographic patterns in benthic and pelagic $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$. Shark, benthic and pelagic $\delta^{13}\text{C}$
2070 values from UP, BG, and CB were often significantly higher than the $\delta^{13}\text{C}$ values in RE and
2071 RO. Similarly, shark, benthic, and pelagic samples from RE and RO had the highest $\delta^{15}\text{N}$
2072 values compared to UP, BG, and CB. These results suggest that individuals in RE and RO did
2073 not forage extensively in the centralized bays within the study area.

2074 **Table 5.3.** Results of linear Bayesian correlation analysis between the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of
 2075 *Rhizoprionodon taylori* and the benthic and pelagic producers across sample bays. ρ defined
 2076 the correlation parameter. Correlations were considered significant if the 95% credibility
 2077 intervals (CIs) of the posterior draws did not overlap with 0.

Isotope	Comparison	Median ρ value	95% CIs ρ value
$\delta^{13}\text{C}$	Benthic-Muscle	0.815	0.063 - 0.996
$\delta^{13}\text{C}$	Pelagic-Muscle	0.626	-0.262 - 0.992
$\delta^{13}\text{C}$	Benthic-Plasma	0.723	-0.081 - 0.0991
$\delta^{13}\text{C}$	Pelagic-Plasma	0.825	0.123 - 0.998
$\delta^{15}\text{N}$	Benthic-Muscle	0.383	-0.503 - 0.960
$\delta^{15}\text{N}$	Pelagic-Muscle	0.661	-0.239 - 0.994
$\delta^{15}\text{N}$	Benthic-Plasma	0.700	-0.1452 - 0.996
$\delta^{15}\text{N}$	Pelagic-Plasma	0.603	-0.278 - 0.996

2078

2079

2080 5.4 Discussion

2081

2082 The results of this Chapter suggest that female *R. taylori* did not forage between areas that are
2083 more than 100 km apart on the Queensland coast over short periods of time (~ 6 months). Less
2084 pronounced geographic differences in *R. taylori* muscle $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ compared to plasma could
2085 indicate individuals foraged between more distant bays (> 100 km) over six months to one year.
2086 However, *R. taylori* muscle $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ also reflected the geographic trends in benthic and
2087 pelagic $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values. Therefore, while some *R. taylori* likely foraged between more
2088 distant bays, most may have spent extended periods of time (~ 1 year) near their respective
2089 capture locations. Therefore, these results suggest *R. taylori* may not make long distance
2090 movements (> 100 km) over a period of time spanning approximately six months to one year.

2091 The regional movement patterns demonstrated by *R. taylori* contrasts with the regional
2092 movement patterns of the Atlantic sharpnose shark, *R. terraenovae*, which has a similar life
2093 history (Loefer and Sedberry 2003) and nearshore residency patterns (see Chapter 4; Carlson *et*
2094 *al.* 2008). In contrast to *R. taylori*, *R. terraenovae* is known to move broadly over short periods
2095 of time (Kohler *et al.* 1998; Carlson *et al.* 2008; Suárez-Moo *et al.* 2013). Tag and recapture data
2096 of *R. terraenovae* from the Gulf of Mexico showed one individual travelled 169 km in 35 days,
2097 and another individual travelled 322 km in 228 days (Carlson *et al.* 2008). Given the biological
2098 and behavioural similarities between *R. taylori* and *R. terraenovae*, it was anticipated that *R.*
2099 *taylori* would exhibit similar large-scale regional movement patterns. Instead, female *R. taylori*
2100 appeared to be similar to the closely related milk shark, *R. acutus*, another small-bodied, fast-
2101 growing species that has a similar geographic range to *R. taylori* (Last and Stevens 2009).
2102 Although data on the movement and site fidelity of *R. acutus* are limited, stock structure analysis

2103 indicates this species exhibits moderate site fidelity with some long distance movements
2104 (Ovenden *et al.* 2011; Schroeder 2011).

2105

2106 Although muscle and plasma $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values suggest limited regional foraging and
2107 dispersal of *R. taylori*, the results do not necessarily indicate long-term residency within
2108 individual bays. Female *R. taylori* sampled from adjacent, central bays (UP, BG, and CB) had
2109 indistinguishable plasma and muscle $\delta^{13}\text{C}$ values. The $\delta^{15}\text{N}$ values of *R. taylori* from UP, BG,
2110 and CB were all relatively low and closely overlapped, particularly in plasma. There are several
2111 possible explanations for the similarity in values between these locations. The first is that
2112 similarity in isotopic baselines between UP, BG, and CB resulted in similar isotope values for
2113 sharks feeding in those areas. A second explanation is that *R. taylori* captured in UP, BG, and
2114 CB regularly moved between and fed within these bays, thus accumulating similar isotope
2115 profiles. The observed similarity could be the result of a combination of these two explanations.
2116 Acoustic tracking from Chapter 4 indicated *R. taylori* move between CB and BG, therefore, it is
2117 likely that the similar isotope values in sharks sampled in UP, BG, and CB were, at least in part,
2118 the result of regular movement between these neighbouring areas.

2119

2120 Given that *R. taylori* likely moved between bays within 100 km in the central part of the study
2121 region, it is unlikely that individuals captured in RE (south) and RO (north) only used their
2122 respective capture bays over one year. If the movement patterns of *R. taylori* in RE and RO are
2123 consistent with those in UP, CB, and BG, it is probable that *R. taylori* captured in RE and RO
2124 moved to other nearby bays within six months to one year. Therefore, it is likely that the
2125 baseline $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values used to establish dietary sources did not account for all possible
2126 $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ that *R. taylori* in RO and RE could have been exposed to. However, the similar

2127 geographic pattern in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values, in combination with relatively fast tissue turnover in
2128 *R. taylori*, supports the general conclusion that *R. taylori* in RE and RO were not travelling as far
2129 as CB, BG, or UP within six months to one year.

2130

2131 It has been suggested that small-bodied, fast growing, productive sharks use multiple nearshore
2132 areas because it increases individual access to resources (Heupel *et al.* 2007; Carlson *et al.* 2008;
2133 Knip *et al.* 2010). The limited regional range indicated by *R. taylori* isotope values in this
2134 chapter suggests that sufficient resources to support *R. taylori* survival, growth, and annual
2135 reproduction were located within 100 km of capture locations. Regional dispersal may have also
2136 been limited by the high energetic cost associated with long-range movement (Roff 1988;
2137 Nøttestad *et al.* 1999; Alerstam *et al.* 2003). Therefore, female *R. taylori* movement patterns
2138 may represent an energetic balance between maximising the benefits of using multiple bays and
2139 minimising the reciprocal costs of long-distance movement. As a result, *R. taylori* may serve as
2140 an important ecological link between bays < 100 km apart, but may not be as significant a link
2141 between more distant bays (> 100 km) over approximately 6-12 months. However, it should be
2142 noted that SIA in sharks is not temporally refined enough to detect sporadic or short-term long-
2143 range dispersal (Logan and Lutcavage 2010; Kim *et al.* 2012). Individuals may have travelled to
2144 more distant bays but if *R. taylori* did not forage in those bays, isotope analysis would not be
2145 able to detect the presence of *R. taylori* in those areas. Moreover, although *R. taylori* in CB had
2146 higher $\delta^{13}\text{C}$ values compared to other populations, several individuals had low $\delta^{13}\text{C}$ values
2147 relative to locally available $\delta^{13}\text{C}$ sources and the majority of the population. This could suggest
2148 longer range movement into areas with lower $\delta^{13}\text{C}$ source values, such as RE and RO. Therefore,
2149 this work cannot rule out the possibility that *R. taylori* made occasional long-range movements.

2150 There was also insufficient data to assess male movement patterns and it is possible that male *R.*
2151 *taylori* exhibit different movement patterns.

2152

2153 The regional movement patterns demonstrated by female *R. taylori* may provide resilience to
2154 local (i.e. single bay) fluctuations in environmental conditions. For example, the results of
2155 Chapter 4 indicated *R. taylori* in CB select for seagrass habitat over other primary habitats.
2156 Individuals may have been selecting for seagrass because that habitat is typically abundant in
2157 small prey (Jackson *et al.* 2001; Gillanders 2006). Local declines in seagrass could negatively
2158 affect *R. taylori* access to resources. However, the results of Chapter 4 and 5 suggest individuals
2159 move between areas within 100 km. The ability to move into different nearby bays to exploit
2160 adequate resources may be a successful survival strategy. By moving between different
2161 locations, *R. taylori* are "buffered" against unproductive conditions in one bay by potentially
2162 more productive conditions in another bay (Yates *et al.* 2012). Thus, the use of multiple
2163 locations may help to stabilize the *R. taylori* population (Secor *et al.* 2009; Yates *et al.* 2012).
2164 However, without more detailed information on resource availability in each area and the dietary
2165 patterns of *R. taylori*, it is difficult to say how changes in any one area may affect the population.

2166

2167 The variability in environmental $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values may have been the result of variable
2168 amounts of freshwater input into each bay. Freshwater runoff into nearshore areas, primarily
2169 from rivers, reduces the $\delta^{13}\text{C}$ value in the environment (Boutton 1991; Hobson 1999).
2170 Freshwater has lower $\delta^{13}\text{C}$ because it contains ^{13}C depleted CO_2 derived from the decomposition
2171 of terrestrial organic matter (Boutton 1991; Hobson 1999). RO and RE typically have high to
2172 moderate amounts of freshwater input respectively (see Chapter 3). This may explain why RE
2173 and RO baseline samples had lower $\delta^{13}\text{C}$ compared to the other locations. Higher $\delta^{15}\text{N}$ in RE and

2174 RO may be the result of large expanses of sugarcane farms in the area in combination with high
2175 rates of freshwater input (Thorburn *et al.* 2011 and refs therein). Nitrogen runoff from fertilizer
2176 used on these farms may have drained into RE and RO and the surrounding areas, resulting in
2177 higher local $\delta^{15}\text{N}$. It should be noted that because zooplankton and phytoplankton were not
2178 separated prior to analysis, comparisons of pelagic $\delta^{15}\text{N}$ values between bays may be biased as a
2179 result of different amounts of zooplankton being collected from each sample location. If certain
2180 bays had a higher volume of zooplankton, the baseline pelagic $\delta^{15}\text{N}$ values reported for those
2181 bays would be artificially high in comparison to other locations (Montoya 2008). However, the
2182 similar geographic pattern in $\delta^{15}\text{N}$ demonstrated by benthic and pelagic samples suggest that the
2183 presence of zooplankton in the samples did not obviously bias the results. As $\delta^{13}\text{C}$ trophic
2184 discrimination factors are relatively small, it was unlikely that different amounts of zooplankton
2185 between locations would affect the interpretation of the $\delta^{13}\text{C}$ results.

2186

2187 Until recently, isotope analysis on elasmobranchs has primarily been used to directly study diet
2188 (MacNeill *et al.* 2005; McMeans *et al.* 2010; Kinney *et al.* 2011). Studies that have used isotopes
2189 to investigate elasmobranch movement have been conducted at either relatively small scales,
2190 such as islands (Papastamatiou *et al.* 2010) and individual bays and inlets (Dale *et al.* 2011;
2191 Reum and Essington 2013), or at coarse scales across isoscapes that span thousands of km
2192 (Carlisle *et al.* 2012). Abrantes and Barnett (2011) assessed the movement patterns of the
2193 broadnose sevengill shark *Notorynchus cepedianus* at a similar geographic scale to this
2194 dissertation, however, that study focused on nearshore and offshore population segregation. The
2195 successful application of isotope analysis to define the regional movement patterns of *R. taylori*
2196 between multiple bays demonstrates this technique may be an affordable and valid alternative to
2197 more traditional methods used to study regional movement. However, isotope analysis may not
2198 be able to define similarly precise regional movements for all species and in all circumstances.

2199

2200 As previously established, slow growing, large-bodied sharks, have relatively slow isotope
2201 incorporation rates (Logan and Lutcavage 2010). As a result, sharks that move quickly through
2202 different habitats may not have enough time to assimilate local isotope values. Effective isotope
2203 movement analysis requires some previous knowledge of the movement and/or diet of the study
2204 species (i.e. catch data and/or acoustic tracking) to guide sampling procedures (Hussey *et al.*
2205 2012a). In this case, catch and acoustic tracking data were available and helped guide several
2206 aspects of this study's methodology, such as study site selection. Regional movement studies
2207 require well defined isotopic baselines that allow for reasonable differentiation between habitats
2208 (Hobson 1999; Rubenstein and Hobson 2004; Hussey *et al.* 2012a). Ecological differences
2209 researchers aim to assess may not always be reflected in the isotopic baselines of the region. In
2210 such cases, researchers could consider examining sulphur isotope ratios in addition to carbon and
2211 nitrogen (West *et al.* 2006; McCauley *et al.* 2014). Differences in $\delta^{34}\text{S}$ between environments
2212 and producers may be present even when $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ are the same (Connolly *et al.* 2004).
2213 Overall, ecological circumstances should be carefully considered before using SIA to study the
2214 movement and home range of elasmobranchs.

2215

2216 These results have provided new information on the residency and movement of a small-bodied
2217 nearshore shark. The use of multiple bays with limited long distance (> 100 km) movements
2218 suggest *R. taylori* may be an important link between adjacent bays and habitats over shorter
2219 periods of time, and potentially more distant bays over longer periods of time. The movement
2220 strategies exhibited by *R. taylori* will have a significant effect on how this species responds to
2221 environmental fluctuations. Ultimately, *R. taylori* may be adaptive to environmental change if
2222 changes are localised to a single bay and/or habitat (Yates *et al.* 2012).

2223
2224
2225
2226
2227
2228
2229
2230
2231
2232
2233
2234
2235
2236
2237
2238
2239
2240
2241
2242
2243
2244
2245
2246
2247

Chapter 6

Geographic and Temporal Variation in the Trophic Ecology of a Small-bodied Shark

6.1 Introduction

Lethal effects of sharks on prey populations via direct predation is essential to maintaining food web structure and population size (Heithaus *et al.* 2008). Indirect effects on prey populations, such as altering prey behaviour through risk avoidance, are also important to ecosystem function (Lima and Dill 1990; Heithaus 2005; Heithaus *et al.* 2012; Klages *et al.* 2014). Variation in diet over time and space can indicate if species play different roles in different environments or through time. Variation in shark diet can also signify changes in local environmental conditions. Predators may alter their diet and hunting strategies to maximise energy intake in response to changing environmental circumstances (Ben-David *et al.* 1997; Eide *et al.* 2005). Therefore, defining trophic ecology of sharks over time and space is critical to understanding ecosystem function and species interaction.

Understanding shark dietary patterns can also help to determine how species will respond to changes in prey availability and biodiversity. For example, highly specialised predators may experience severely reduced foraging efficiency when preferred prey populations have decreased (see Chapter 2; Terraube *et al.* 2011). As a result, diet specialists may experience a decrease in growth, reproduction, and population size (Suarez and Case 2002; Graham 2007; Graham *et al.* 2009). In contrast, generalist predators are more likely to maintain stable levels of prey capture success when specific prey populations decline (Terraube *et al.* 2011). Therefore, generalists

2248 will probably be less vulnerable to population decline as a result of fluctuations in prey
2249 availability.

2250

2251 Stable isotope analysis is an increasingly common method to evaluate the temporal and spatial
2252 variation in elasmobranch diets (Hussey *et al.* 2012a). The two most commonly used isotopes
2253 are $\delta^{13}\text{C}$ ($^{13}\text{C}/^{12}\text{C}$) and $\delta^{15}\text{N}$ ($^{15}\text{N}/^{14}\text{N}$) as they provide complementary information on species
2254 dietary patterns (Shiffman *et al.* 2012). The $\delta^{13}\text{C}$ in animal tissues increases in relatively small
2255 amounts between prey and predators but more obviously varies between different primary
2256 producers and environments as a result of different local biogeochemical processes (Tieszen *et*
2257 *al.* 1983; Peterson and Fry 1987; Boutton 1991). Therefore tissue $\delta^{13}\text{C}$ can be used to estimate
2258 the carbon sources of a consumer (DeNiro and Epstein 1978; Peterson and Fry 1987). In
2259 contrast, $\delta^{15}\text{N}$ values predictably increase from prey to predator (Deniro and Epstein 1981;
2260 Peterson and Fry 1987). As a result, $\delta^{15}\text{N}$ in animal tissues can be used to estimate the trophic
2261 position of an individual (Post 2002). The $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of individuals can also be used to
2262 estimate the isotopic niche of a population (Layman *et al.* 2012). Collectively, this information
2263 can be used to estimate the dietary specialisation of a population in a given area and/or a species
2264 as a whole, depending on the geographic range of the study. Different tissues with different
2265 metabolic rates will integrate isotopes from prey over different periods of time, ranging from
2266 months to years (Logan and Lutcavage 2010; Kim *et al.* 2012). Therefore $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ from
2267 different tissues can be used to evaluate changes in dietary sources over time. Although isotope
2268 analysis provides less detailed data on prey composition than stomach content analysis, isotope
2269 analysis is a more cost effective and, under most circumstances, non-lethal alternative
2270 (Hammerschalg and Sulikowski 2011; Hussey *et al.* 2011).

2271

2272 Chapters 4 and 5 established that *Rhizoprionodon taylori* is best defined as a species with a low
2273 degree of habitat specialisation; however, *R. taylori* has demonstrated a strong preference for
2274 seagrass habitat, potentially because seagrass is typically highly productive and abundant in
2275 small teleost prey. Therefore, benthic food web sources may be a primary contributor to *R.*
2276 *taylori* diet. Previous stomach content analysis of *R. taylori* indicated this species fed on a wide
2277 variety of prey types, including teleosts, crustaceans, and cephalopods (Simpfendorfer 1998).
2278 Unfortunately a large proportion of empty stomachs hindered analysis and the source of prey
2279 was not able to be determined (i.e. benthic or pelagic food webs) (Simpfendorfer 1998). Chapter
2280 5 showed *R. taylori* move between bays < 100 km apart, but more distant populations are likely
2281 separated for approximately one year. It is possible that *R. taylori* in different locations may
2282 have distinct diets resulting in unique relationships with local environments. Geographically
2283 distinct populations of marine mammals (e.g. *Mirounga leonine*; Banks *et al.* 2014), birds (e.g.
2284 *Larus audouinii* and *Larus argentatus*; Oro *et al.* 1996; Herbet *et al.* 1999), and reptiles (e.g.
2285 *Thamnophis validus*; de Queiroz *et al.* 2001) have been shown to have distinct diets, likely due
2286 to spatial differences in food availability.

2287

2288 The aim of this chapter was to define the trophic ecology of *R. taylori* across multiple
2289 environments and time scales using stable isotope analysis. Plasma and muscle $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of
2290 *R. taylori* were compared to $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ baselines (seagrass, macroalgae, and plankton) from
2291 multiple embayments to determine the isotopic niche, trophic position, and the benthic and
2292 pelagic contribution to *R. taylori* diet in each area and over time. This chapter will improve
2293 understanding of the diets of small-bodied sharks and how this species may respond to
2294 variability in environmental conditions.

2295

2296 **6.2 Data Analysis**

2297

2298 The combined findings of Chapters 4 and 5 indicated female *R. taylori* captured in UP, BG, and
2299 CB probably move between these areas over the course of at least one year. Thus UP, BG, and
2300 CB likely represent a single potential feeding area for *R. taylori* captured in any one of these
2301 bays. The results of the acoustic analysis in Chapter 4 support the conclusion that *R. taylori*
2302 move between these locations. Chapter 5 analysis also indicated that female *R. taylori* captured
2303 in UP, BG, and CB were not likely to move to RE or RO within the time span of plasma and
2304 muscle tissue turnover. Therefore, to accurately represent the likely extent of dietary sources
2305 available to *R. taylori*, isotopic values of environmental baselines and *R. taylori* were grouped
2306 into three areas, RO, RE, and the Cleveland Bay Unit (CBU), which included UP, BG, and CB
2307 (Fig. 6.1). These groupings were referred to as sampling or sample areas. Large-scale movement
2308 patterns could only be established for female *R. taylori*, therefore males were excluded from
2309 analyses (see Chapter 5). *Rhizoprionodon taylori* plasma $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ turnover was estimated
2310 to take approximately 6 months while muscle was estimated to take one year (see Chapter 5).

2311

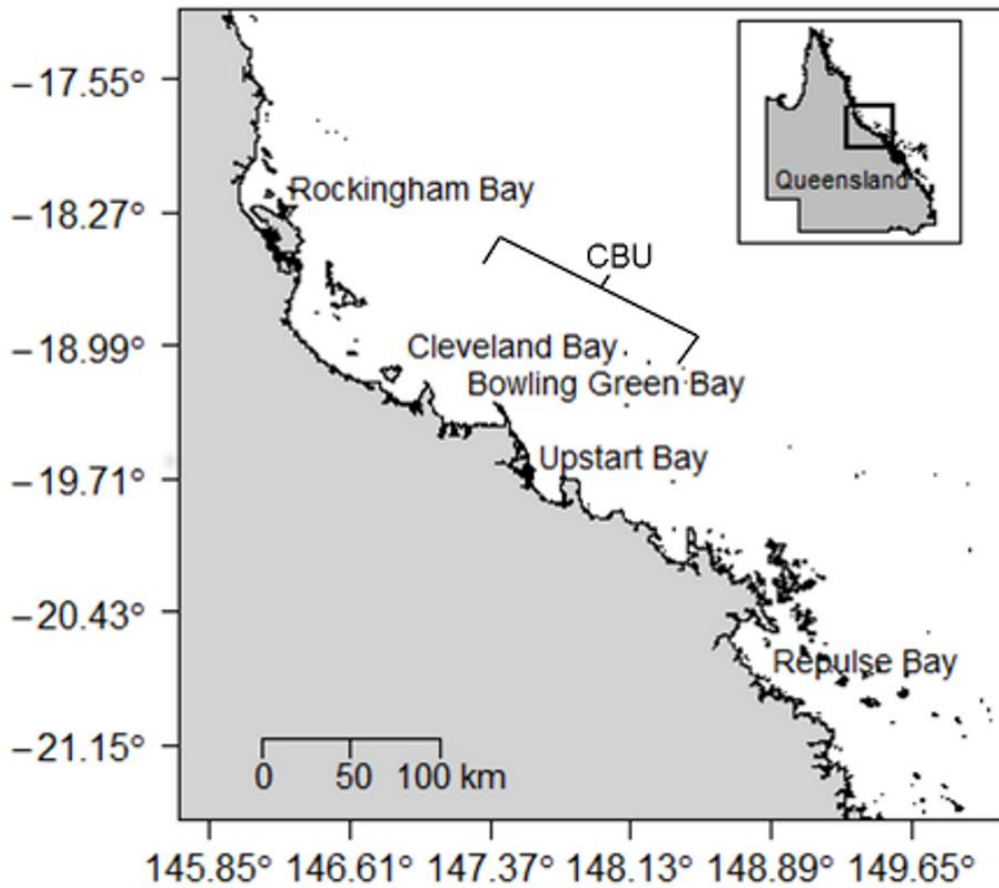
2312 A Bayesian ANOVA (Gelman 2007) was used to assess differences between sample areas in
2313 benthic and pelagic $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ baselines. The Bayesian ANOVA used non-informative priors
2314 and was calculated according to the formulations:

2315 The Likelihood

$$2316 y_{ij} \sim \text{Normal}(\mu + \alpha_i, \sigma^2) \quad (6.1)$$

2317 The Priors

$$2318 \mu \sim \text{Normal}(0, 10^{-6}) \quad (6.2a)$$



2319

2320 **Fig. 6.1.** Map of stable isotope sampling region indicating the five sampling locations and three

2321 designated feeding areas, Rockingham Bay, Cleveland Bay Unit (CBU), and Repulse Bay for

2322 *Rhizoprionodon taylori*. Inset indicates location along the north Queensland coast, Australia.

2323

2324 $\alpha_i \sim \text{Normal}(0, 10^{-6})$ (6.2b)

2325 Where σ was the sample variance, μ was the mean response, and α was the effect due to sample
2326 area. Differences between locations were significant if the 95% credibility intervals of posterior
2327 draws did not overlap. A Bayesian ANOVA was also used to test for differences between sample
2328 areas in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in muscle and plasma. Results of Chapter 5 showed that animal size had
2329 limited and inconsistent effects on *R. taylori* $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$. Therefore, size was not included in
2330 this analysis.

2331

2332 Individual trophic positions (TP) were calculated for each tissue in each sample area according
2333 to Post (2002) using a constant $\delta^{15}\text{N}$ diet tissue discrimination factor of 3.2:

2334

2335
$$TP_{individual} = TP_{baseline} + \frac{\delta^{15}\text{N}_{individual} - \delta^{15}\text{N}_{baseline}}{3.2}$$
 (6.3)

2336 Where $TP_{baseline}$ and $\delta^{15}\text{N}_{baseline}$ were the known TP and median $\delta^{15}\text{N}$ value of environmental
2337 baselines (based on the results of Bayesian analysis). Seagrass $\delta^{15}\text{N}$ (TP 1) and plankton (TP 1.5)
2338 were calculated separately and the range was combined. Plankton was given a TP of 1.5 because
2339 it was a combination of phytoplankton and zooplankton.

2340

2341 *Rhizoprionodon taylori* $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values were used to calculate the isotopic niche for each
2342 tissue in each sample area. The isotopic niche was calculated using the package *SIAR* (Parnell *et*
2343 *al.* 2010) in R version 3.0.2 (R Development Core Team, 2013) as described by Jackson *et al.*
2344 (2011). This method uses Bayesian inference techniques to produce (1) the smallest convex hulls
2345 that contain all individual $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values within a group (i.e. sample area) to represent

2346 maximum niche breath (Layman *et al.* 2007), and (2) Bayesian standard ellipses (SEA_b) which
2347 incorporate the 40% densest $\delta^{13}\text{C}/\delta^{15}\text{N}$ data points within a group and thus better represents the
2348 “average” isotopic niche breadth of the population (Jackson *et al.* 2011). This method was
2349 chosen because a Bayesian framework for isotopic niche calculations better accounts for sources
2350 of uncertainty and variability inherent in stable isotope analysis and allows for more robust
2351 comparisons between groups, particularly for small and/or variable sample sizes (Parnell *et al.*
2352 2010).

2353

2354 Relative contributions of benthic and pelagic sources to *R. taylori* diet for each tissue in each
2355 sample area was calculated using a two source Bayesian mixing model with the *SIAR* package
2356 (Jackson *et al.* 2011) in R version 3.0.2 (R Development Core Team, 2013). All other Bayesian
2357 models were fitted using the package R2jags (Su and Yajima 2014) in R version 3.0.2 (R
2358 Development Core Team: www.r-project.org) and JAGS, version 3.4.0 (Plummer 2003).
2359 Posterior draws were built using three Markov chains with 10000 iterations per chain and a
2360 thinning interval of 10. Chain mixing trace plots and autocorrelation values were used to assess
2361 each applied version of the models.

2362

2363 **6.3 Results**

2364

2365 **6.3.1 Study Site $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$**

2366

2367 Forty-seven pelagic and 55 benthic samples were collected from across the three sampling areas.
2368 The CBU had a considerably larger combined benthic and pelagic $\delta^{13}\text{C}$ range than RO and RE
2369 samples (Table 6.1). The CBU also had a slightly larger range of $\delta^{15}\text{N}$ values.

2370 **Table 6.1.** The $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ range (mean \pm SD) of combined pelagic and benthic samples
2371 from each sample area, Repulse Bay (RE), Cleveland Bay Unit (CBU), and Rockingham Bay
2372 (RO).

2373

Sample area	Sample size	$\delta^{13}\text{C}$ range	$\delta^{15}\text{N}$ range
RE	17	-23.28 - -15.15 (-19.9 \pm 2.5)	1.33-6.22 (5.5 \pm 1.4)
CBU	64	-20.54- -8.44 (-15.9 \pm 3.9)	0.62-6.78 (3.4 \pm 1.7)
RO	20	-21.46- -12.05 (-17.7 \pm 2.9)	2.94-7.26 (4.6 \pm 1.3)

2374

2375

2376 Rockingham Bay and RE had relatively similar baseline $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ ranges. Benthic samples
2377 had higher $\delta^{13}\text{C}$ values than pelagic samples in all areas (Fig. 6.2a). In contrast, pelagic samples
2378 had higher $\delta^{15}\text{N}$ values than benthic samples in all areas. CBU benthic and pelagic samples had
2379 higher $\delta^{13}\text{C}$ than RO and RE samples. RE and RO had higher $\delta^{15}\text{N}$ values than CBU samples.

2380

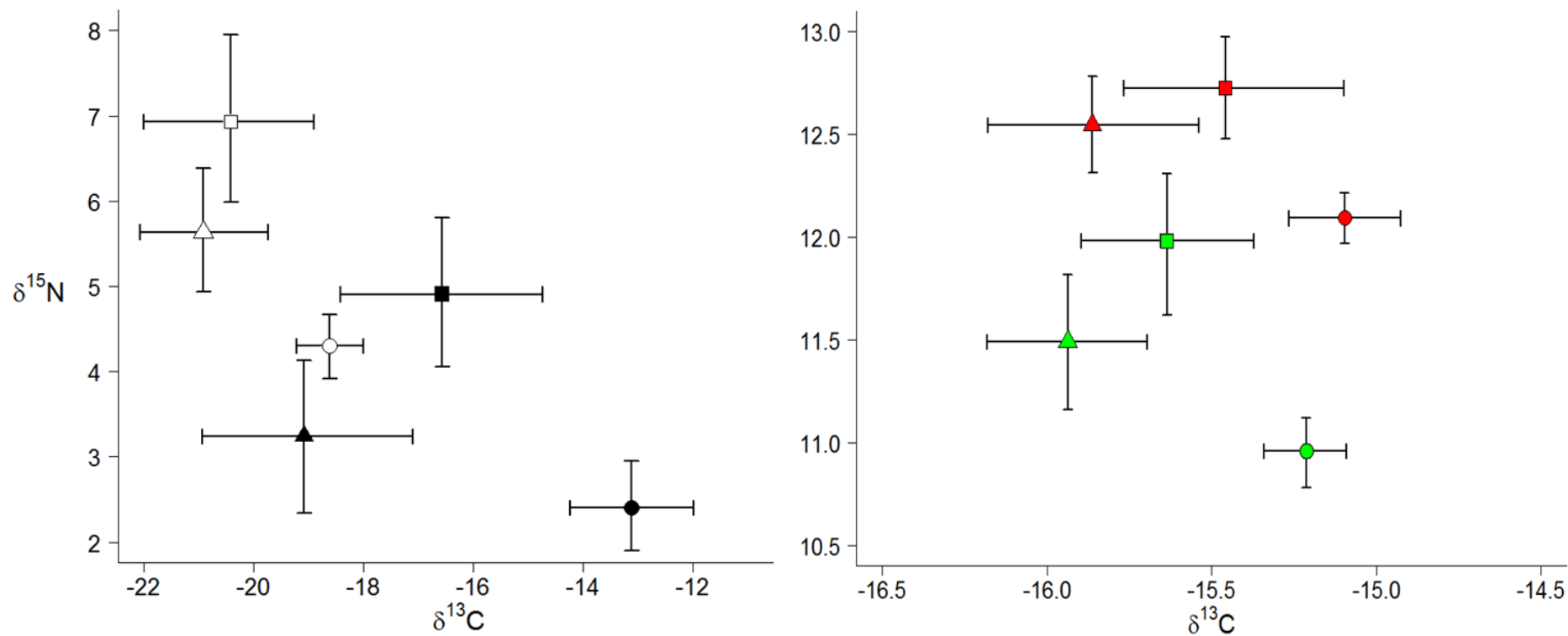
2381 **6.3.2 Shark $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$**

2382

2383 One hundred and sixteen female *R. taylori* were sampled from across the three sample areas
2384 (Table 6.2); sizes ranged from 543 to 780 mm (mean \pm SE = 681 \pm 5.0). *Rhizoprionodon taylori*
2385 $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ followed similar geographical patterns to environmental isotope baselines (Fig.
2386 6.2b). Plasma and muscle $\delta^{13}\text{C}$ from female *R. taylori* captured in CBU was higher than the $\delta^{13}\text{C}$
2387 values in RO and RE. Plasma and muscle $\delta^{15}\text{N}$ from *R. taylori* in RE and RO was higher than the
2388 $\delta^{15}\text{N}$ in CBU. The trophic position of each *R. taylori* sample population spanned more than one
2389 trophic level ($\sim 3.2\text{‰}$) and indicated each population was composed of secondary and/or
2390 tertiary consumers (Table 6.2). Trophic position varied between tissues but less so between
2391 locations. Muscle TPs were higher than plasma TPs in all three locations. The magnitude of
2392 decrease in TP from muscle to plasma was similar in each location. *Rhizoprionodon taylori* in
2393 RE had a lower range of TPs than *R. taylori* in RO and CBU.

2394

2395 Isotopic niche breadth of *R. taylori* varied between locations and tissues. Analysis of muscle
2396 $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ indicated the CBU population had the largest isotopic niche (Fig. 6.3a). However,
2397 credibility intervals of niche breadth calculations indicated that the CBU population only had a
2398 significantly larger isotopic niche than *R. taylori* in RE (Fig. 6.3c). Analysis of plasma $\delta^{13}\text{C}$ and
2399 $\delta^{15}\text{N}$ revealed all three populations had similar niche breadth sizes, although CBU was still the



2400

2401

2402 **Fig. 6.2.** Median $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ results of Bayesian ANOVA of (a) benthic (black) and pelagic samples (white), and *Rhizoprionodon taylori*

2403 muscle (red) and plasma (green) in Repulse Bay (■), the Cleveland Bay Unit (●) and Rockingham Bay. Black lines show 95% credibility

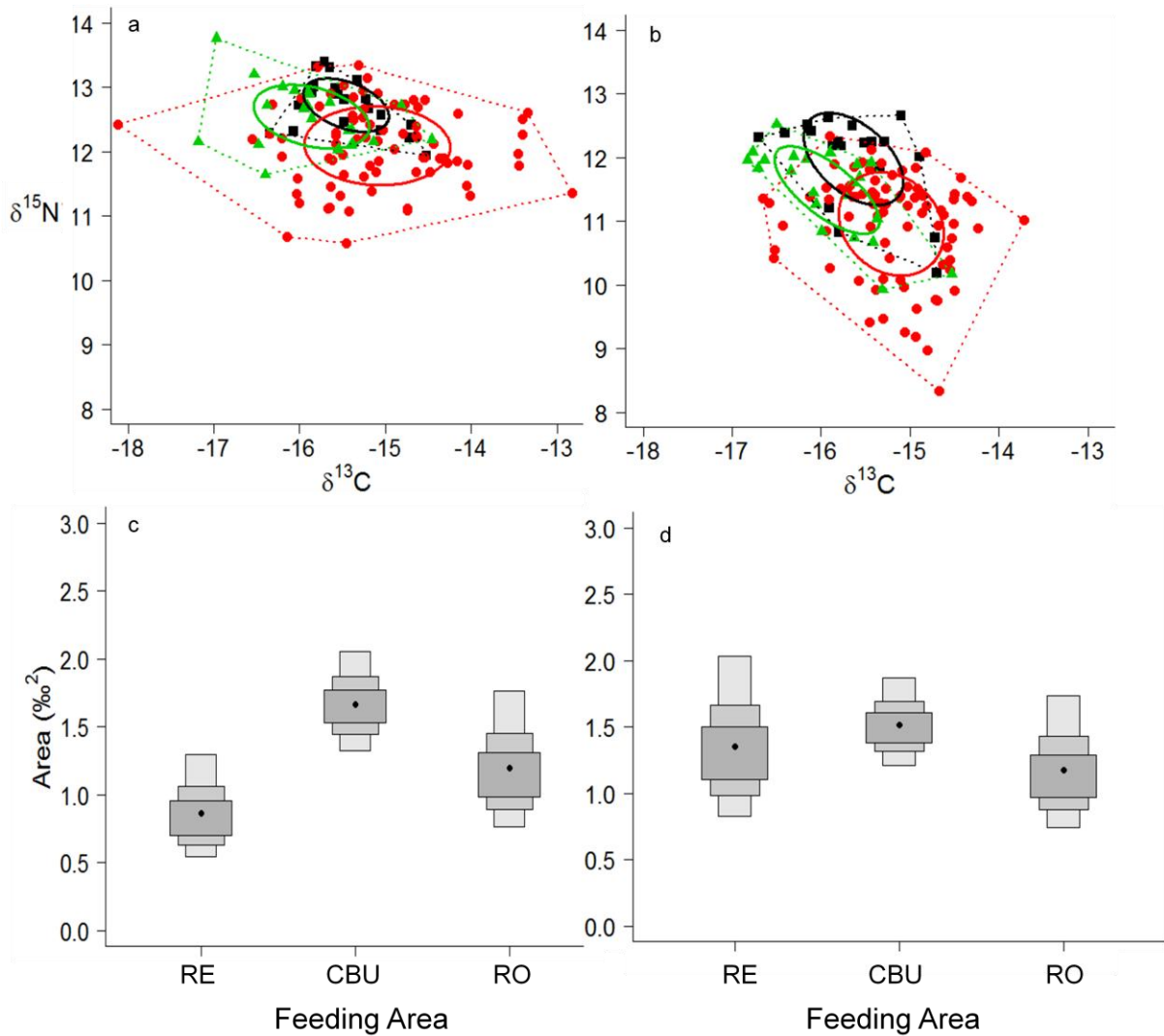
2404 intervals of posterior draws.

2405

2406 **Table 6. 2.** Total catch, $\delta^{13}\text{C}$ range, $\delta^{15}\text{N}$ range, trophic position (TP) range and mean with
 2407 standard error (SE) of female *Rhizoprionodon taylori* in each sample area (Area), Repulse
 2408 Bay (RE), Cleveland Bay Unit (CBU), and Rockingham Bay (RO).

Area	Total Catch	Tissue	$\delta^{13}\text{C}$ range	$\delta^{15}\text{N}$ range	TP Range	Mean TP \pm SE
RE	20	Muscle	-16.6 - -14.5	11.94-13.39	3.2-4.1	3.7 \pm 0.04
		Plasma	-16.7 - -14.7	10.19-12.66	2.7-3.9	3.5 \pm 0.05
CBU	76	Muscle	-18.1 - -13.3	10.57-13.35	3.6-4.9	4.3 \pm 0.02
		Plasma	-16.5- - 13.7	8.33-12.34	2.9-4.6	3.9 \pm 0.03
RO	20	Muscle	-17.0 - -14.5	11.64-13.76	3.6-4.8	4.2 \pm 0.05
		Plasma	-16.8- -14.5	9.92-12.52	3.1-4.4	3.8 \pm 0.05

2409



2410

2411 **Fig. 6.3.** Isotopic niche breadth of *Rhizoprionodon taylori*. Convex hulls of total niche width

2412 of muscle (a) and plasma (b) are dotted lines. Bayesian Standard Ellipses (SEA_b) isotope

2413 niches are shown for Repulse Bay (RE; ■/black), Cleveland Bay Unit (CBU; ●/red), and

2414 Rockingham Bay (RO; ▲/green). SEA_b area calculations are also given as 50, 75, 95

2415 credibility intervals (dark to light grey) of posterior draws for muscle (c) and plasma (d),

2416 black dots indicate median values.

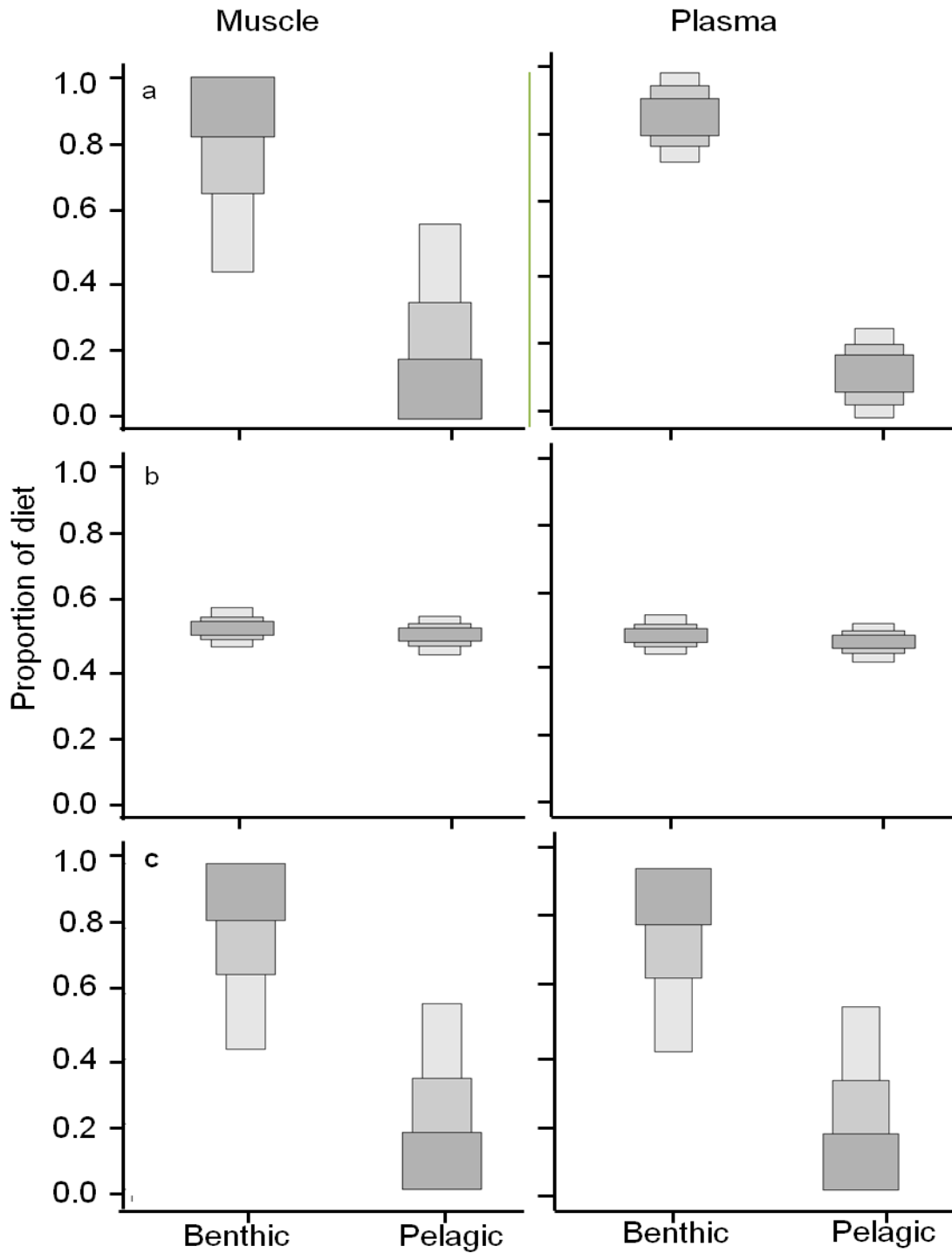
2417

2418 largest (Fig. 6.3b,d). Isotopic niche size remained relatively constant in CBU and RO between
2419 muscle and plasma. In contrast, the niche breadth of *R. taylori* in RE substantially increased
2420 from muscle to plasma. This large increase in RE niche breadth was primarily the result of an
2421 increase in the range of *R. taylori* $\delta^{15}\text{N}$ in that area.

2422

2423 Pelagic and benthic contributions to *R. taylori* diet varied between locations (Fig. 6.4). In CBU
2424 the mixing model showed that the diet was split equally between benthic and pelagic sources
2425 for both muscle and plasma. In contrast, the diets of *R. taylori* in RE and RO were primarily
2426 composed of benthic sources. The constrained credibility intervals of the RE plasma mixing
2427 model (Fig. 6.4a) strongly indicates benthic prey were the primary dietary source in this area
2428 more recently. However, wide ranging credibility intervals from posterior draws of RE muscle
2429 and RO muscle and plasma mixing models suggest *R. taylori* in these areas likely still consume
2430 prey from pelagic food webs.

2431



2432

2433 **Fig. 6.4.** Proportional contributions of benthic and pelagic food web sources to
 2434 *Rhizoprionodon taylori* diet using a two-source Bayesian mixing model for plasma and
 2435 muscle tissue in a) Repulse Bay, b) Cleveland Bay Unit, and c) Rockingham Bay. Shaded
 2436 boxes are 50, 75, 95 (from dark to light grey) credibility intervals of posterior draws of SEA_b .

2437

2438 **Discussion**

2439

2440 Small-bodied, highly productive, moderately mobile predators such as *R. taylori*
2441 (Simpfendorfer 1993) represent an important link in marine food webs. Abundant, small-
2442 bodied sharks can connect habitats and environments through movement and serve as both as
2443 predator and prey items (Lundberg and Moberg 2003). Geographic and/or temporal changes in
2444 the diet of species like *R. taylori* can provide valuable information on species ecological roles
2445 in different marine communities, species vulnerability to environmental change, and indicate
2446 variation in environmental conditions between areas. Therefore, data on the diet of small-
2447 bodied species is critical to a better understanding of marine ecosystems.

2448

2449 Chapter 4 showed *R. taylori* selected for nearshore seagrass habitat, potentially because this
2450 habitat is highly productive and abundant in suitable prey. As a result, it was expected that
2451 benthic or seagrass-based prey would represent a large component of *R. taylori* diet. Results of
2452 this chapter have confirmed benthic sources are a significant and in some areas a majority
2453 contributor to *R. taylori* diet, however, it is also clear that *R. taylori* consume prey from pelagic
2454 sources. The wide range of trophic positions of *R. taylori* in each area also suggests this species
2455 consumes a variety of prey. These findings are consistent with *R. taylori* stomach content
2456 analysis that indicated individuals fed on a variety of prey types, including teleosts, crustaceans
2457 and cephalopods (Simpfendorfer 1998). Stomach content analysis also concluded that
2458 approximately half of *R. taylori* diet in Cleveland Bay was composed of demersal prey, while
2459 the other half included pelagic prey types (Simpfendorfer 1998). Demersal and pelagic prey
2460 types do not necessarily stem from benthic and pelagic carbon sources respectively, but the
2461 presence of both prey types in *R. taylori* stomachs supports the conclusions of this Chapter. An
2462 even division of prey types in *R. taylori* diet in Cleveland Bay is also consistent with mixing

2463 model results within the CBU, supporting the accuracy of these results. Therefore, although the
2464 analysis present in this chapter is not a direct measure population specialisation (see Chapter
2465 2), the results presented here indicate *R. taylori* has a broad dietary niche and is likely best
2466 defined as a mesopredator with a low degree of dietary specialisation, at least at a population
2467 level (Matich *et al.* 2010).

2468

2469 The broad dietary niche and range of trophic positions exhibited by *R. taylori* collectively
2470 across all sampling regions is similar to other species within this genera such as the Atlantic
2471 sharpnose shark *Rhizoprionodon terraenovae* (Gelsleichter *et al.* 1999; Bethea *et al.* 2006), the
2472 Brazilian sharpnose shark *Rhizoprionodon lalandii* (Bornatowski *et al.* 2012), and the milk
2473 shark *Rhizoprionodon acutus* (White 2004). Previous isotope analysis of elasmobranchs and
2474 teleosts in Cleveland Bay also found that *R. taylori* had similar carbon ranges as similarly sized
2475 generalist predators, specifically the hardnose shark *Carcharhinus macloti*, the milk shark *R.*
2476 *acutus*, and the barramundi *Lates calcarifer* (Kinney *et al.* 2011). These results suggest that *R.*
2477 *taylori* in Cleveland Bay likely consumed similar carbon sources as other local generalist
2478 mesopredators. The niche breadth of *R. taylori* is also comparable to other small-bodied
2479 mesopredators in distant locations. The isotopic niche breadth of the generalist mesopredator
2480 the southern stingray, *Dasyatis americana*, was similar to the niche breadth of *R. taylori* in the
2481 CBU (Tilley *et al.* 2013). As generalists, these small-bodied species are likely important
2482 maintainers of ecosystem function and biodiversity (Richmond *et al.* 2005). *Rhizoprionodon*
2483 *taylori* likely influences the population size and structure of numerous nearshore species in
2484 both benthic and pelagic food webs.

2485

2486 The structural influence of *R. taylori*, however, probably differs based on location as there was
2487 considerable geographic variation in source contribution to diet and niche breadth. Geographic
2488 variation in diet has been documented in a number of shark species, including the bonnethead
2489 shark *Sphyrna tiburo* (Bethea *et al.* 2007), *R. terraenovae* (Drymon *et al.* 2012), the
2490 narrownose smooth-hound, *Mustelus schmitti*, (Belleggia *et al.* 2012), the lemon shark
2491 *Negaprion brevirostris* (Cortés and Gruber 1990), the sandbar shark *Carcharhinus plumbeus*
2492 (McElroy *et al.* 2006), and the Starrspotted-dogfish *Mustelus manazo* (Yamaguchi and
2493 Taniuchi 2000). A common inference among these studies is that geographic variation in diet is
2494 the result of geographic variation in prey availability and the opportunistic feeding strategies of
2495 the predators. As *R. taylori* consume a wide range of prey they will most likely consume prey
2496 that is highly abundant or most beneficial to them in each area (Mittelbach *et al.* 1992; Salini *et*
2497 *al.* 1992; Simpfendorfer *et al.* 2001; Reeve *et al.* 2009). As a result, the diet of female *R.*
2498 *taylori* will likely fluctuate based on changes in local prey availability. Therefore, it is probable
2499 that benthic prey in RE and RO were more abundant or easily accessible. It is also possible
2500 benthic prey are a better source of energy in RE and RO than in the CBU and *R. taylori* may
2501 actually be adopting selective strategies. Not all prey found in *R. taylori* stomachs in Cleveland
2502 Bay were consumed in equal proportions to local abundance (Simpfendorfer 1998). Therefore
2503 either situation could explain why female *R. taylori* consumed a larger proportion of benthic
2504 prey in RE and RO. However, it should be noted it is unlikely all possible $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$
2505 dietary sources were accounted for in RO and RE. In Chapter 5 it was established that although
2506 *R. taylori* in RO and RE were spending large periods of time feeding in their respective capture
2507 sites, they were probably moving into other nearby areas. Therefore, benthic and pelagic
2508 sampling in RO and RE did not fully account for the dietary $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ available to *R.*
2509 *taylori* sampled in those areas, particularly over the time span incorporated by muscle tissue.
2510 This would bias the results of the mixing models and may help to explain why there were large

2511 credibility intervals in mixing model outputs from RE and RO. Therefore, the results of the RO
2512 and RE muscle mixing models, and the implications they have for geographic variation in diet,
2513 should be interpreted with caution.

2514

2515 The geographic variation in isotope niche breadth may have also been due to variable patterns
2516 in selection or prey abundance. The *R. taylori* in CBU had a broad diet composed of an equal
2517 mix of benthic and pelagic prey, while *R. taylori* in RE and RO had smaller niche breadths and
2518 fed primarily on benthic prey. The less specialised diet of *R. taylori* in the CBU could result in
2519 a larger isotopic niche. However, it is also possible that the differences in niche breadth are due
2520 to differences in isotopic baselines between locations. The CBU had the largest range in
2521 baseline $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values. If *R. taylori* were opportunistic and/or broad predators,
2522 presumably the isotopic niche of *R. taylori* would increase as the range in baseline $\delta^{13}\text{C}$ and
2523 $\delta^{15}\text{N}$ values also increased. This would be most obvious in tissues with shorter turnover times
2524 (i.e. plasma). Therefore, while variation in niche breadth size between locations may be the
2525 result of differences in selection and sources contributions, it may also be due to the relative
2526 range of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of local sources.

2527

2528 There was also moderate geographic variation in female *R. taylori* $\delta^{15}\text{N}$ and trophic position.
2529 Most of this variability is likely due to variability in $\delta^{15}\text{N}$ at the base of the food chain as shark
2530 tissues exhibited similar geographic trends in $\delta^{15}\text{N}$ as environmental baselines. The higher $\delta^{15}\text{N}$
2531 in RE and RO may have been because these bays are adjacent to large expanses of sugarcane
2532 farms and thus exposed to high levels of nitrogen runoff (see Chapter 5). However, trophic
2533 position calculations, which accounted for variation in $\delta^{15}\text{N}$ baselines, found *R. taylori* in RE
2534 were consuming prey at lower trophic positions than in other areas. This could indicate there is

2535 a lower abundance of higher trophic level prey in RE compared to RO and CBU. It is also
2536 possible that lower trophic level prey was abundant or beneficial in RE and thus formed a
2537 larger component of local diet. However, differences in trophic level were small and warrant
2538 further investigation, potentially using stomach content analysis. Overall, the differences in diet
2539 between locations suggest prey availability likely varies between locations and that *R. taylori*
2540 may have different effects on prey structure in each area.

2541

2542 Comparisons between muscle and plasma isotope values suggested there was some temporal
2543 variation in *R. taylori* diet. The trophic position of *R. taylori* decreased in all three sample areas
2544 from muscle to plasma, suggesting a region-wide change in prey availability over time.

2545 Previous work has shown that decreases in $\delta^{15}\text{N}$ in elasmobranchs is often associated with
2546 decreased amounts of teleost consumption (Domi *et al.* 2005; MacNeil *et al.* 2005). Teleosts
2547 generally have higher $\delta^{15}\text{N}$ values and trophic levels. Therefore, it is possible a recent decrease
2548 in teleosts at high trophic levels in all areas would have forced female *R. taylori* to consume
2549 more prey at lower trophic levels than in previous years. It is also possible that lower order
2550 prey became highly abundant and thus formed a larger component of the diet.

2551

2552 Despite changes in trophic level, the relative contributions of benthic and pelagic sources to *R.*
2553 *taylori* diet were consistent over time in all areas. Niche breadth size in RO and CBU was also
2554 consistent while niche breadth in RE increased from muscle to plasma. Collectively, these
2555 results suggest that *R. taylori* in all three sample areas recently consumed prey at lower trophic
2556 levels, but maintained a large niche breadth that incorporated both food webs over
2557 approximately one year. The unique increase in niche breadth in RE could be energetic
2558 compensation for the decline in higher trophic prey or some other preferred prey. It is also

2559 possible that previously unavailable prey types became available relatively recently in the RE
2560 area, resulting in niche expansion. Although the direct cause(s) of changes in *R. taylori* diet are
2561 difficult to determine without more detail on local prey availability, the occurrence of temporal
2562 and spatial variability in the diet indicates *R. taylori* are probably highly adaptive consumers.
2563 Female *R. taylori* are likely capable of adjusting their hunting strategies to local conditions and
2564 fluctuations in prey availability.

2565

2566 Results of this chapter indicate that *R. taylori* has a low degree of dietary specialisation and is
2567 capable of opportunistic and possibly selective strategies. Therefore, the effect of *R. taylori* on
2568 nearshore food webs may change based on local environmental conditions and prey
2569 availability. Given individuals likely remain within a 100 km range of their capture location for
2570 at least a year (see Chapter 5), spatial and temporal variation in *R. taylori* diet may not only
2571 indicate differences in local prey biodiversity, but also that this species has unique effects on
2572 distinct local ecosystems. For that reason, this study emphasises the importance of examining
2573 the trophic ecology of species over multiple areas and time scales. The results from this work
2574 also suggest that female *R. taylori* are likely adaptive to changes in prey availability.

2575 Consequently, *R. taylori* may be less vulnerable to declines in prey availability of a particular
2576 species (McKinney 1997; Colles *et al.* 2009; Terraube *et al.* 2011; Curtis *et al.* 2013).

2577 *Rhizoprionodon taylori* may compensate for declines in specific prey species by expanding or
2578 shifting their dietary niche and consuming other prey that remain available. As a species with
2579 low habitat and likely dietary specialisation, *R. taylori* is probably resilient to environmental
2580 change, particularly at a local level.

2581

Chapter 7

Movement Ecology of a Nearshore, Medium-bodied Shark, the Creek Whaler *Carcharhinus fitzroyensis*

7.1 Introduction

Nearshore areas are highly productive and dynamic environments that often contain a diverse range of habitat types, such as seagrass meadows, mangroves, and rocky and/or sandy substrate (Robertson and Duke 1987; Beck *et al.* 2001). As a result of this productivity and diversity, nearshore areas function as important foraging and nursery grounds for many shark species (Heupel *et al.* 2007; Knip *et al.* 2010). However, nearshore ecosystem health and biodiversity is in a state of global decline (Suchanek 1994; Lotze *et al.* 2006) due to numerous anthropogenic influences, including coastal pollution (Shahidul Islam and Tanaka 2004), inshore fishing (Blaber *et al.* 2000), and climate change (Harley *et al.* 2006). Consequently, there is a need to determine how environmental change affects nearshore marine species.

The majority of shark research in nearshore areas has investigated the habitat use of large-bodied species that use these areas as nursery grounds (e.g. Rechisky and Wetherbee 2003; Ortega *et al.* 2009; DiGirolamo *et al.* 2012). Nearshore areas provide juvenile sharks with protection from predators and in some cases productive foraging grounds (Branstetter 1990; Castro 1993). As a result, juveniles of large-bodied species are often highly resident to nearshore areas (e.g. Knip *et al.* 2011a). In contrast, large-bodied adults use substantially larger amounts of space, spend more time offshore, and may only use specific nearshore areas

2606 for short periods of time for foraging or pupping (Feldheim *et al.* 2002; Grubbs 2010).
2607 Considerably less data are available on the nearshore habitat use of smaller coastal species,
2608 such as the small-bodied milk shark *Rhizoprionodon acutus* (maximum total length 100 cm)
2609 or the medium-bodied nervous shark *Carcharhinus cautus* (maximum total length 150 cm)
2610 (Last and Stevens 2009). Medium-bodied species (max TL < 1500 mm) in particular have
2611 received little focused study. In contrast to large-bodied sharks, small- and medium-bodied
2612 species use nearshore areas consistently throughout their lives as juveniles and adults (Knip *et*
2613 *al.* 2010).

2614

2615 Limited work suggests nearshore species exhibit a variety of movement and habitat use
2616 strategies. For example, the Australian sharpnose shark *Rhizoprionodon taylori* and Atlantic
2617 sharpnose shark *R. terraenovae* have large coastal home ranges with individuals using a wide
2618 array of habitats (Carlson *et al.* 2008; Chapter 4). In contrast, the spottail shark *Carcharhinus*
2619 *sorrah* exhibits high site fidelity to single nearshore embayments with some individuals using
2620 small home ranges and consistent habitats (Knip *et al.* 2012a). Different strategies will likely
2621 affect the fitness of sharks in nearshore ecosystems.

2622

2623 The purpose of this chapter was to define the movement and habitat use patterns of
2624 *Carcharhinus fitzroyensis* in a nearshore area, compare movement strategies to sharks of
2625 other size categories, and evaluate its vulnerability to environmental change. Passive acoustic
2626 telemetry was used to determine *C. fitzroyensis* residency, space use, habitat selection and
2627 specialisation. Results of this chapter will lead to a better understanding of how medium-
2628 bodied coastal sharks use nearshore habitats relative to other species groups.

2629 **7.2 Data Analysis**

2630

2631 **7.2.1 Residency**

2632

2633 Presence was evaluated each day. Individuals were deemed present if they were detected two
2634 or more times in the array in a given day. Residency was determined using a residency index
2635 that calculated the number of days an individual was present in the array as a proportion of
2636 the total days monitored (e.g. Abecasis and Erzini 2008; Werry *et al.* 2014). The index ranged
2637 from 1 to 0, indicating high to low residency, respectively. An ANCOVA was used to test for
2638 differences in residency between sample years with STL as a covariate. Individuals that were
2639 present in the bay for less than 14 days total were excluded from space use and habitat
2640 analysis.

2641

2642 **7.2.2. Space use**

2643

2644 Individual positions were estimated using the mean position algorithm described by
2645 Simpfendorfer *et al.* (2002) to determine individual centre of activity (COA) locations. The
2646 COA represented a weighted mean position for each 30 minute interval an individual was
2647 detected in the array. COA locations were used to calculate individual monthly activity space
2648 as 50% and 95% kernel utilisation distributions (KUDs) using the *adehabitatHR* package in R
2649 version 3.0 (Calenge 2006). To prevent overestimation of KUD size, KUD calculations
2650 incorporated an impassable boundary that represented the Cleveland Bay coastline. KUD
2651 calculations used a smoothing parameter of 0.008. A linear mixed effects model was used to
2652 determine if 50% and 95% KUD size was affected by animal size, month and/or diel period.
2653 To account for repeated measures in the data, individual was incorporated as a random factor
2654 in the resultant models. Models were computed using the *nlme* package in R (Pinheiro *et al.*

2655 2013). Models were compared using Akaike information criterion with a small sample size
2656 bias correction (AICc) where models with the lowest AICc were considered to be the most
2657 significant drivers of KUD size. Akaike weights were also calculated to facilitate model
2658 assessment (Wagenmakers and Farrell 2004).

2659

2660 **7.2.3 Habitat selection**

2661

2662 Cleveland Bay was divided into regions based on the distribution of five distinct habitat
2663 types. Receivers were assigned a habitat type based on their location in the bay. Habitat
2664 availability was determined by calculating the proportion of receivers in each habitat.
2665 Proportional habitat use by individuals was determined by comparing the amount of time
2666 spent in a habitat to the total amount of time spent in the array.

2667

2668 The proportion of time spent in each habitat by individuals was compared to habitat
2669 availability using Strauss's (1979) linear selectivity index (L_i) to determine if *C. fitzroyensis*
2670 were selecting for or avoiding habitats:

$$2671 \quad L_i = r_i - p_i \quad (7.1)$$

2672 where r_i was the proportion of habitat i used and p_i was the proportion of habitat i available in
2673 the study site. $L_i < 0$ indicated negative selection or avoidance. $L_i > 0$ indicated positive
2674 selection. $L_i = 0$ indicated the habitat was neither positively nor negatively selected for and
2675 was used opportunistically (Strauss 1979). The mean of resident individual L_i values was
2676 calculated for each year and each two week time period within each year to assess population
2677 habitat selection over time. A chi-squared goodness-of-fit test was used to determine if the

2678 proportion of time spent in each habitat by the resident population relative to habitat
2679 availability was significantly different, and hence if mean annual selection was significant.

2680

2681 **7.2.4 Habitat niche breadth**

2682

2683 Niche breadth of resident individuals was measured using Hutchinson's niche definition
2684 which only included the habitats a species used and in what proportions (Hutchinson 1957;
2685 Devictor *et al.* 2010). Based on this definition, a modified Freeman-Tukey statistic was used
2686 to calculate population niche breadth (Smith 1982):

$$FT = \sum_{i=1}^R (p_i q_i)^{1/2} \quad (7.2)$$

2687 where q_i was the proportion of habitat i available in the study site, p_i was the proportion of
2688 habitat i used, and R was the total number of habitats available. The output ranged from 1,
2689 which indicated a large niche, to 0, which indicated a narrow niche and a highly specialised
2690 species.

2691

2692 A variation of equation (7.2) was used to calculate individual niche overlap (Arlettaz 1999):

$$FT = \sum_{i=1}^R (p_{ik} q_{ik})^{1/2} \quad (7.3)$$

2693 where p_{ik} and p_{jk} were the proportions of habitat R used by individual i and j respectively. The
2694 output similarly ranged from 1 to 0. A value of 1 indicated complete overlap in habitat niche
2695 breadth between a given pair of individuals and 0 indicated no overlap in habitat niche
2696 breadth between a given pair of individuals. Where applicable, all data was checked for

2697 normality using normality and homogeneity of variance diagnostics in R, version 3.0 (R
2698 Development Core Team: www.r-project.org) and data were \log_{10} transformed if necessary.

2699

2700 **7.3 Results**

2701

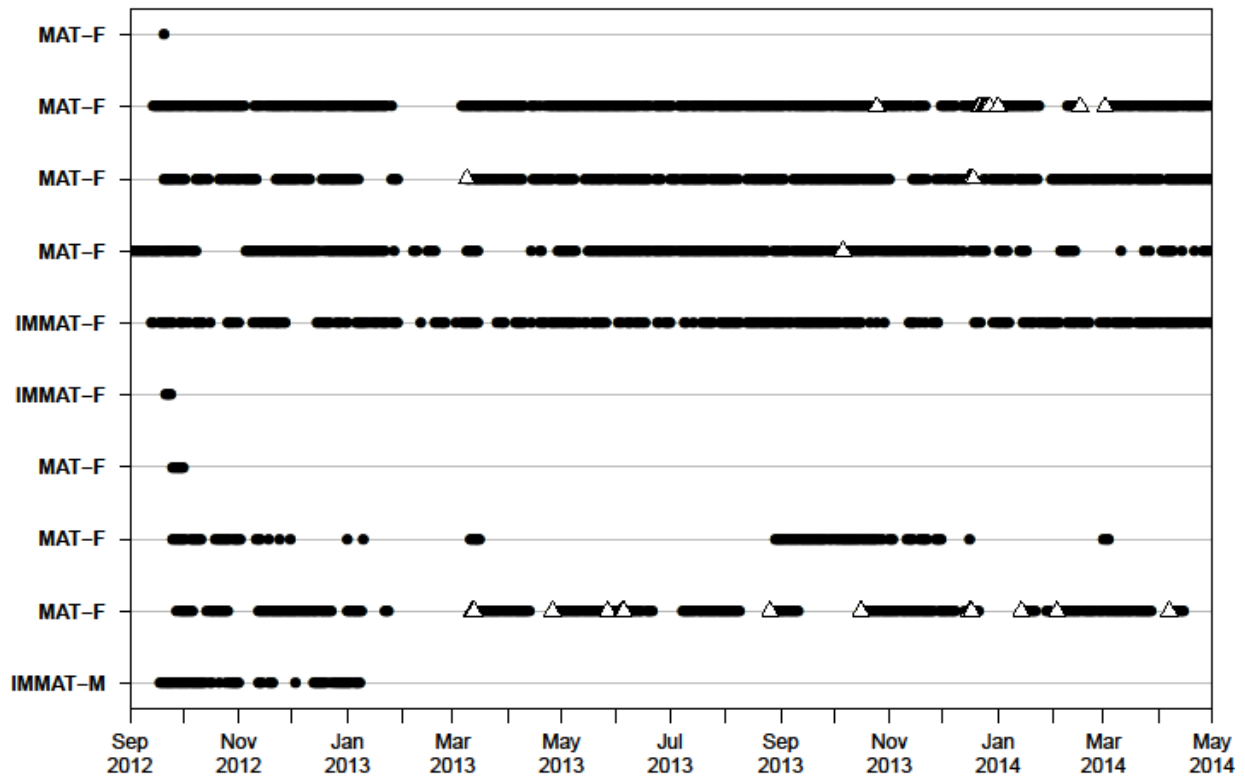
2702 Sixteen *C. fitzroyensis* (3 male, 13 female) were released with acoustic transmitters in
2703 Cleveland Bay between February and September 2012. Size ranged from 679 to 1370 mm
2704 STL (mean \pm SE = 943 \pm 48.9). All individuals were caught and released on the eastern side
2705 of Cleveland Bay. Five *C. fitzroyensis* were not detected or died following release and were
2706 excluded from analysis. One immature female was recaptured and collected by a local
2707 commercial fisherman 36 km north of the original release location approximately two weeks
2708 after release. These individuals were also excluded from analysis. The remaining 10 *C.*
2709 *fitzroyensis* (1 male, 9 female) were monitored in Cleveland Bay from September 2012 to
2710 May 2014. Size and clasper state indicated the male was immature. Size indicated seven
2711 females were mature and two were immature. Size ranged from 679 to 1109 mm (mean \pm SE
2712 = 945 \pm 54.2).

2713

2714 **7.3.1 Residency**

2715

2716 Individuals were present in Cleveland Bay for 1 to 452 days (mean \pm SE = 205 \pm 53) (Fig.
2717 7.1). Three individuals left the array within two weeks of release and did not return within the
2718 monitoring period. The residency index ranged from 0.002-0.74 (mean \pm SE = 0.34 \pm 0.09).
2719 There was no significant relationship between residency and size (ANCOVA, $F_{(1,18)} = 0.1616$,
2720 $P > 0.05$) or sample year (ANCOVA, $F_{(1,18)} = 0.1379$, $P > 0.05$). There was no clear seasonal
2721 pattern in presence except for one mature female that exhibited a seasonal pattern in presence.



2722
2723

2724 **Fig. 7.1.** Daily presence of *Carcharhinus fitzroyensis* released with acoustic transmitters in
 2725 Cleveland Bay in 2012- 2014. Individuals are identified by maturity (mature=MAT,
 2726 immature=IMMAT) and sex (male=M, female=F). Detections in Cleveland Bay are indicated
 2727 by black circles. Additional detections in Bowling Green Bay are indicated by white triangles.
 2728

2729 This individual was consistently present from September to December in 2012 and 2013, was
2730 present for several weeks in April 2013 and 2014, and was never detected between May and
2731 August during any year of the study.

2732 Four mature female *C. fitzroyensis* were detected on receivers in Bowling Green Bay. These
2733 individuals were some of the most highly resident to Cleveland Bay. Two were only detected
2734 in Bowling Green Bay for single days before returning to Cleveland Bay. However, the two
2735 other females made brief excursions lasting approximately one week into Bowling Green Bay
2736 throughout the monitoring period.

2737

2738 **7.3.2 Space Use**

2739

2740 With the exception of one individual, all resident individuals exclusively used the eastern side
2741 of Cleveland Bay. Individual monthly activity space ranged from 2.6 to 19.8 km² (mean \pm SE
2742 = 10.6 km² \pm 0.3) for 50% KUDs and 9.1 to 81.9 km² (mean \pm SE = 47.9 km² \pm 1.0) for 95%
2743 KUDs. The best possible model to explain both 50% and 95% KUD size included month and
2744 diel period as factors (Table 7.1). Shark length appeared to have little or no effect on KUD
2745 size and the model that only included size as a factor was worse than the null model for both
2746 50% and 95% KUDs. This indicates immature and mature individuals utilized similar
2747 amounts of space within the bay. KUD size was larger during the day than at night for both
2748 50% and 95% KUDs (Fig. 7.1). The influence of month on KUD size was most prominent in
2749 May and August (Fig. 7.2). In May, there was a distinct increase in 50% and 95% KUD size
2750 compared to all other months of the year. In contrast, August 50% and 95% KUD size was
2751 considerably smaller than all other months. KUD locations also fluctuated on a monthly basis
2752 according to this pattern. In August, activity space was centralized adjacent to the south-
2753 eastern creek mouths in Cleveland Bay (Fig. 7.3). During the rest of the year, but most

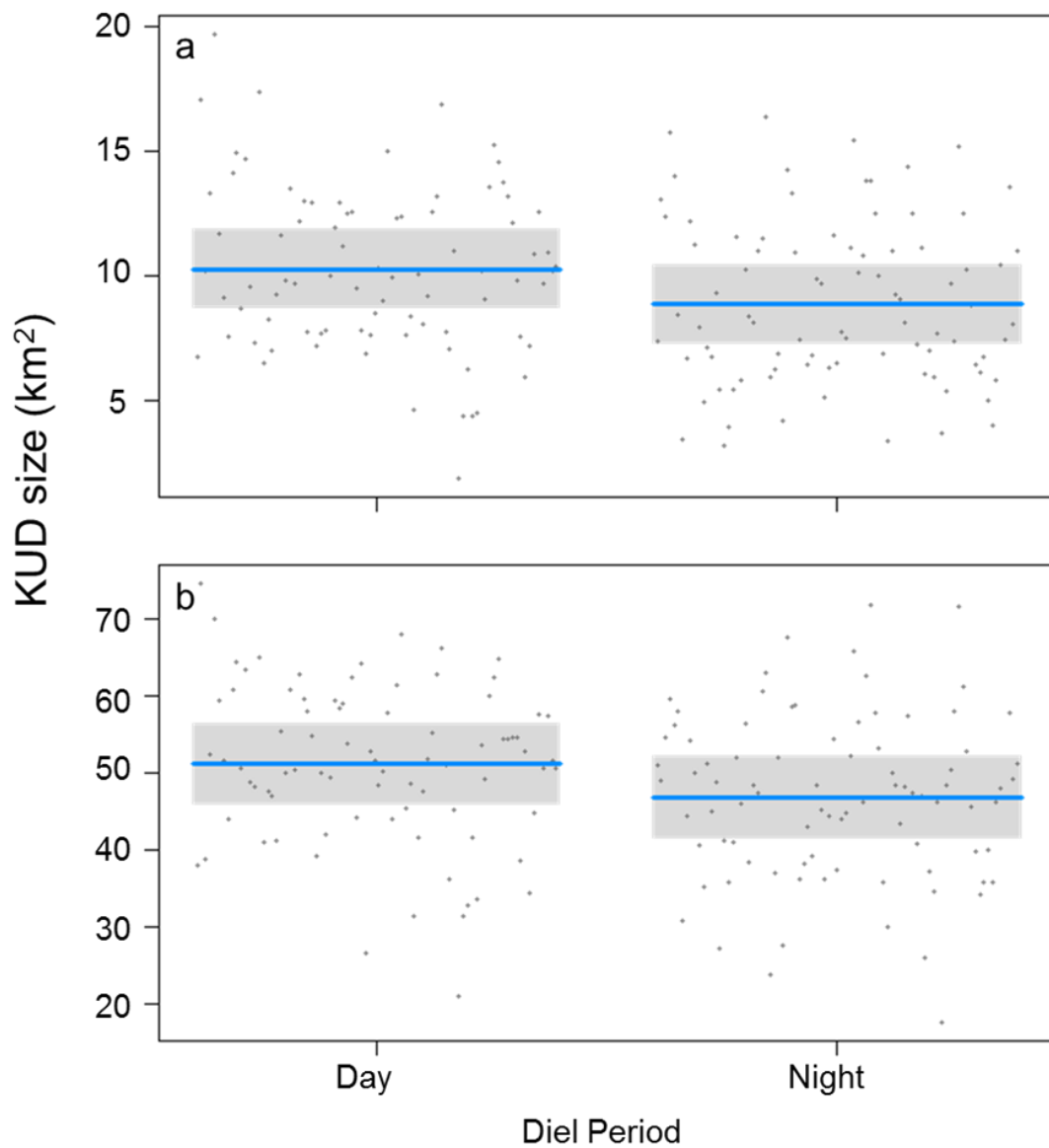
2754

2755 **Table 7.1.** Effect of animal size, month and diel period on creek whaler *Carcharhinus fitzroyensis* 50% and 95% kernel utilisation distribution
 2756 (KUD) size (km²). Degrees of Freedom (df), Akaike's information criterion correction (AICc), Δ AICc, and Akaike weights (W) values are given
 2757 for each model. AICc values are marked with an * if models were significantly different from the null model using a likelihood ratio test. The
 2758 best fit models are **bolded**.

2759

Model	df	50% KUD AICc	95% KUD AICc	50% KUD Δ AICc	95% KUD Δ AICc	50% KUD W	95% KUD W
KUD ~ 1	3	1127.2	1624.2	81.79	102.98	0.00	0.00
KUD ~ Size	4	1136.0	1631.4	90.62	109.97	0.00	0.00
KUD ~ Size+Diel	5	1131.0	1624.2	85.60	102.77	0.00	0.00
KUD ~ Size+Month	15	1064.3*	1539.3*	18.91	17.93	0.00	0.00
KUD ~ Month	14	1054.3*	1532.1*	8.87	10.72	0.012	0.05
KUD ~ Month+Diel	15	1045.4*	1521.4*	0.00	0.00	0.982	0.969
KUD ~ Diel	4	1122.2*	1617.2*	76.79	95.82	0.00	0.00
KUD ~ Size+Month+Diel	16	1055.4*	1528.6*	10.02	7.19	0.007	0.027

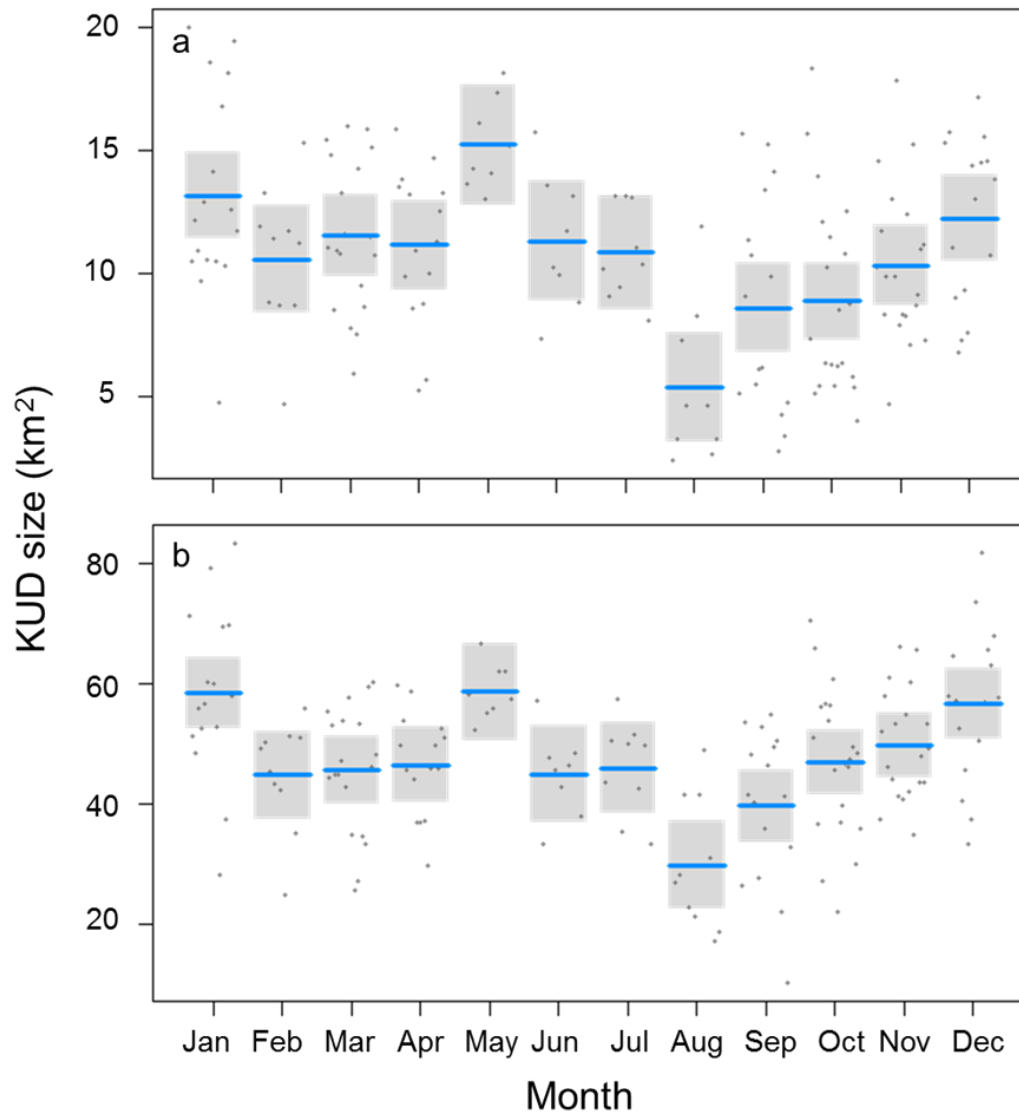
2760



2762

2763 **Fig.7.2.** Effect of Diel period on (a) 50% and (b) 95% kernel utilisation size (km^2) of
 2764 *Carcharhinus fitzroyensis*. Blue bars are mean predicted value of linear mixed effect model
 2765 output, grey bands are 95% confidence intervals of model output.

2766



2767

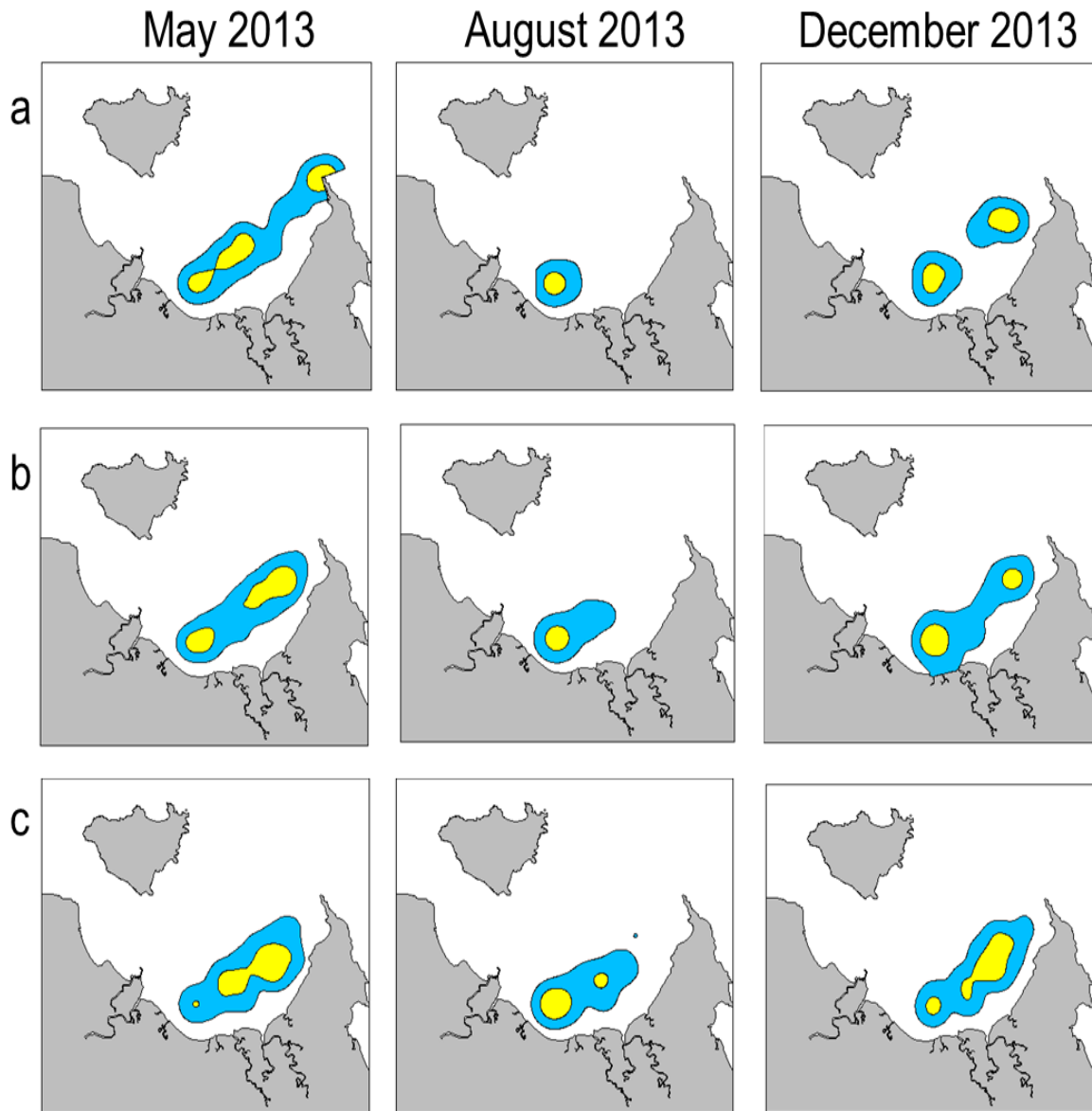
2768

2769 **Fig.7.3.** Effect of Month on 50% (a) and 95% (b) kernel utilisation size (km²) of

2770 *Carcharhinus fitzroyensis*. Blue bars are mean predicted value of linear mixed effect model

2771 output, grey bands are 95% confidence intervals of model output.

2772



2773

2774

2775 **Fig.7.4.** Monthly activity spaces of (a) one immature and (b, c) two mature *Carcharhinus*

2776 *fitzroyensis* in Cleveland Bay in May 2013, August 2013 and December 2013. Each panel

2777 shows the 95% (blue fill) and 50% (yellow fill) kernel utilisation distributions.

2778

2779 notably in May, KUD positions were more widely spread throughout the eastern half of the
2780 bay. The monthly pattern in KUDs location was observed in immature and mature
2781 individuals.

2782 **7.3.3 Habitat Selection and Specialisation**

2783
2784 Individuals were detected in all five habitat types; however, the majority of time was spent in
2785 seagrass habitat (Table 7.2). There was no significant difference in time spent in each habitat
2786 between immature and mature individuals (Chi-squared test, $X^2_{16}=20.00$, $P > 0.05$). As a
2787 result, all individuals were grouped for habitat selection and specialisation analysis. Mean
2788 individual Strauss selection values for the entire monitoring period indicated *C. fitzroyensis*
2789 selected for seagrass habitat, used outer bay mud substrate opportunistically, and avoided
2790 reef, mudflat and sand inshore habitats (Fig. 7.5). A Chi-squared goodness of fit test indicated
2791 selection was significant ($X^2_4=144.758$, $P < 0.05$). Mean fortnightly Strauss index values
2792 showed that selection for seagrass was consistently positive throughout the monitoring period
2793 (Fig. 7.6). Selection for outer bay habitat was highly variable and fluctuated between positive
2794 and negative selection. There was no apparent seasonal pattern in the selection of outer bay
2795 mud substrate; however, individuals selected for outer bay habitat less often in the second
2796 year of the study. Mudflat, reef, and sandy inshore habitats were consistently avoided over
2797 time.

2798

2799 Mean individual niche breadth for *C. fitzroyensis* was moderate and ranged from 0.70 to 0.77
2800 (mean \pm SE = 0.73 ± 0.03). Mean individual niche overlap was large and ranged from 0.91 to
2801 1 (mean \pm SE = 0.97 ± 0.03), indicating resident *C. fitzroyensis* used nearly identical
2802 proportions of the same habitats.

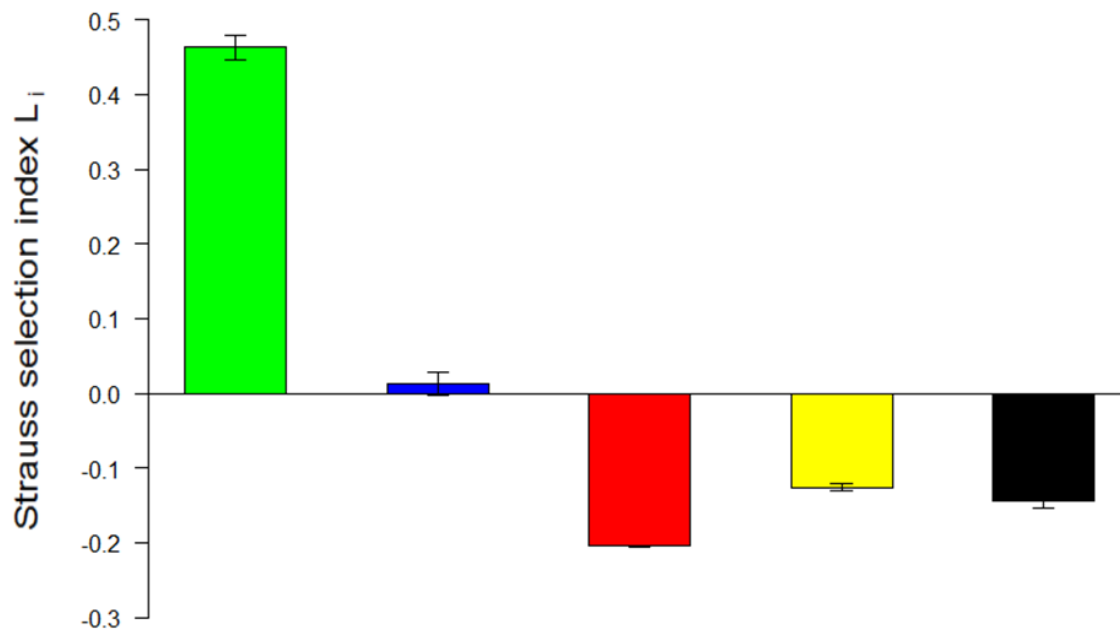
2803

2804 **Table 7.2.** Mean and standard error of time spent in each habitat by mature and immature
 2805 *Carcharhinus fitzroyensis* in Cleveland Bay, measured as a percent (%) with standard error
 2806 (SE).

Habitat Type	Time Spent \pm SE Mature	Time Spent \pm SE Immature
Seagrass	72.9 \pm 3.5	72.7 \pm 6.26
Outer bay mud substrate	20.4 \pm 4.4	20.0 \pm 2.7
Reef	0.09 \pm 0.1	0.0 \pm 0.0
Sandy inshore	1.6 \pm 1.2	0.19 \pm 0.19
Intertidal mudflat	5.0 \pm 2.5	7.11 \pm 3.69

2807

2808

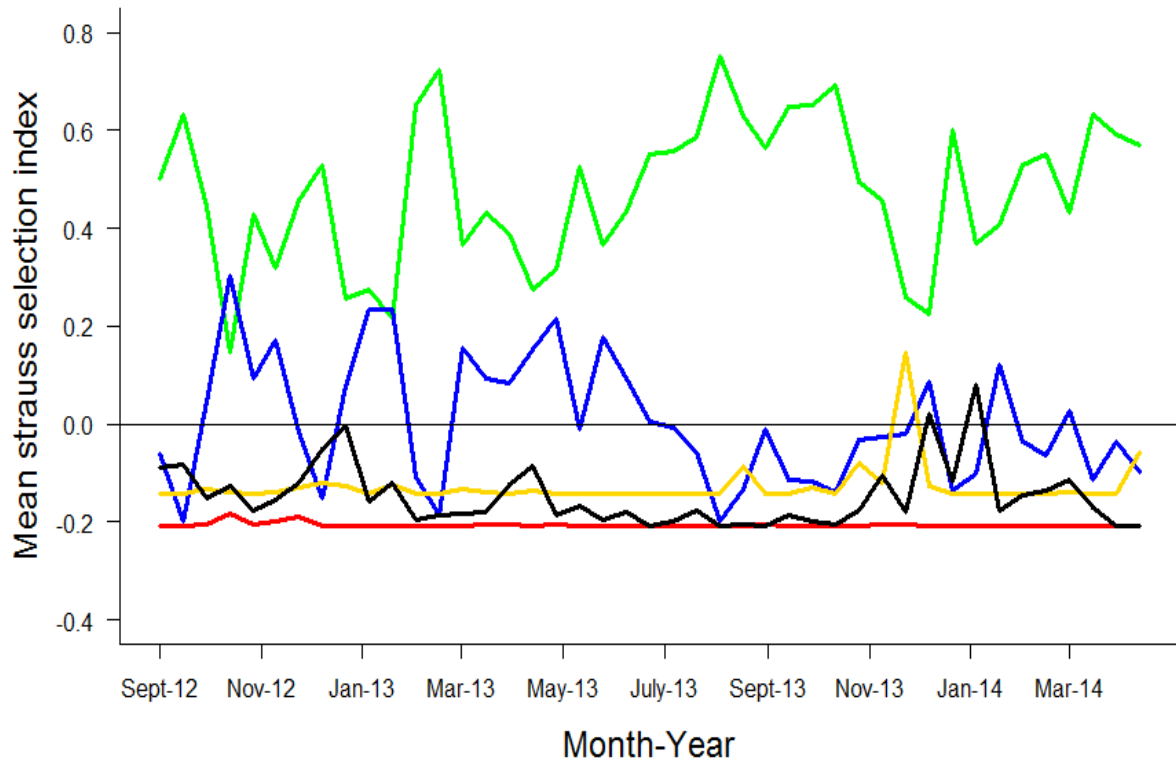


2809

2810 **Fig.7.5.** Mean Strauss linear selection index values of seagrass (green), outer bay mud
 2811 substrate (blue), reef (red), sandy inshore (yellow) and intertidal mudflat (black) habitats by
 2812 *Carcharhinus fitzroyensis* in Cleveland Bay between September 2012 to May 2014. Bars
 2813 indicate standard error.

2814

2815



2816

2817 **Fig. 7.6.** Mean fortnightly individual Strauss linear selection values of *Carcharhinus*
2818 *fitzroyensis* in Cleveland Bay for seagrass (green), outer bay mud substrate (blue), reef (red),
2819 sandy inshore (yellow) and intertidal mudflat (black).

2820

2821 **7.4 Discussion**

2822

2823 Similarity in immature and mature *C. fitzroyensis* residency, space use, and habitat use
2824 patterns suggests individuals share nearshore resources. High individual niche breadth
2825 overlap and consistent habitat selection patterns exhibited by the sample population showed
2826 that there was limited individual variability in habitat use. All individuals almost exclusively
2827 used seagrass habitats and to a lesser extent outer bay mud substrate habitats. The shared use
2828 of nearshore areas by mature and immature individuals is consistent with other small- and
2829 medium-bodied coastal species, such as *R. acutus* (Henderson *et al.* 2006; Schroeder 2011),
2830 *R. terraenovae* (Carlson *et al.* 2008), *R. taylori* (Chapter 4 and 5), *C. sorrah* (Knip *et al.*
2831 2012a), and the grey-smooth hound shark *Mustelus californicus* (Ebert 2003; Espinoza *et al.*
2832 2011). However, in contrast to *C. fitzroyensis*, many small-bodied coastal species use a wide
2833 array of habitats. For example, *R. taylori*, *R. terraenovae*, and *M. californicus* all exhibited
2834 low residency to single nearshore areas (Carlson *et al.* 2008; Espinoza *et al.* 2011; Chapter 4).
2835 *Rhizoprionodon taylori* and *R. terraenovae* also used a variety of habitats in nearshore
2836 embayments (Carlson *et al.* 2008; Chapter 4). Although *C. sorrah* displayed high residency
2837 and site fidelity to single nearshore habitats, individuals also demonstrated a high degree of
2838 spatial segregation and individual variability in habitat use (Knip *et al.* 2012a; Knip *et al.*
2839 2012c). Unlike *C. fitzroyensis*, *C. sorrah* demonstrated no habitat preference at the population
2840 level. Instead, the residency and habitat use patterns exhibited by *C. fitzroyensis* are more
2841 similar to those exhibited by juveniles of large-bodied species. Juveniles of large-bodied
2842 species, such as the pigeye shark *Carcharhinus amboinensis*, the bull shark *Carcharhinus*
2843 *leucas*, and the lemon shark *Negaprion brevirostris*, are highly resident to nearshore areas and
2844 often use the same habitats and home ranges for long periods of time (Heupel *et al.* 2010;
2845 Murchie *et al.* 2010; Knip *et al.* 2011a). Therefore, the results of this study demonstrate that

2846 *C. fitzroyensis* used a combination of residency and movement strategies previously reported
2847 in small- and large-bodied species.

2848 The long term use of seagrass exhibited by the *C. fitzroyensis* population suggests this habitat
2849 had sufficient resources to support the majority of the population over long periods of time.

2850 Consistent use of nursery areas by large-bodied sharks is in part motivated by high nearshore
2851 productivity and prey availability (Castro 1993), Seagrass habitats are usually highly productive
2852 and contain relatively large populations of small fish and bottom dwelling prey (Orth *et al.*
2853 1984; Edgar and Shaw 1995; Jackson *et al.* 2001). Therefore, seagrass habitat may be excellent
2854 foraging grounds for *C. fitzroyensis*. Moreover, consistent use of seagrass habitat on the eastern
2855 side of Cleveland Bay could result in greater familiarity with the distribution of resources in
2856 that area (Kuba *et al.* 2010; Schluessel 2014). Increased resource availability and better
2857 knowledge of resource distribution could make it highly beneficial for immature and mature *C.*
2858 *fitzroyensis* to remain in the eastern side of the bay over long periods of time. It should be noted
2859 that some of the highly resident *C. fitzroyensis* individuals made occasional excursions into
2860 Bowling Green Bay and the habitats used during those excursions could not be precisely
2861 determined. However, Bowling Green Bay is dominated by mud substrate and seagrass habitats
2862 (Furnas 2003; GBRMPA 2011) Therefore, when individuals were in Bowling Green Bay it is
2863 likely they were utilizing habitats similar to those used in

2864 It is possible that the close association with seagrass habitat and the eastern half of the bay
2865 demonstrated by *C. fitzroyensis* was due to bias in catch distribution. Fishing effort was
2866 concentrated in the eastern portion of the bay. Therefore the high residency to that area may
2867 have been the by-product of where individuals were captured (Knip *et al.* 2012). However,
2868 fishery independent and dependent sampling throughout Cleveland Bay revealed that *C.*
2869 *fitzroyensis* has been almost exclusively captured on the eastern side of the bay (Simpfendorfer,

2870 unpublished data). Therefore, tracking and activity space data are likely representative of how
2871 this species uses space within Cleveland Bay and are not the result of sampling bias. Fishery
2872 independent catch data from northern Australia also showed this species is often caught in mud
2873 substrate habitats and/or in areas with high potential for benthic growth (Simpfendorfer,
2874 unpublished data; GBRMPA 2011). Therefore, results from coastal catch data supports the
2875 conclusion that seagrass is one of the primary habitats used by *C. fitzroyensis*.

2876

2877 Long-term residency by immature *C. fitzroyensis* may also have been influenced by life
2878 history (Heupel *et al.* 2007). As previously discussed, juveniles of large-bodied species
2879 exhibit high site fidelity to nearshore areas because these areas can provide protection from
2880 predators (Castro 1993). Although *C. fitzroyensis* is potentially a productive species, it
2881 reaches sexual maturity five to six years later than small-bodied species and generally gives
2882 birth to smaller litters (Last and Stevens 2009; Smart *et al.* 2013). Therefore, high residency
2883 to shallow nearshore areas may provide some beneficial protection for immature *C.*
2884 *fitzroyensis* from large-bodied predators (Heithaus 2007). This may also explain why, similar
2885 to large-bodied species, neonate *C. fitzroyensis* have been observed in shallow habitats and
2886 expand their range as they grow (Harry *et al.* 2011).

2887

2888 While seagrass habitat was the principal habitat used by *C. fitzroyensis*, moderate habitat
2889 specialisation values and selection analysis indicate seagrass habitat was used in conjunction
2890 with outer bay mud substrate. Use of outer bay mud substrate was highly variable over time and
2891 lack of seasonality in selection suggests this habitat was not used in response to seasonal abiotic
2892 changes in the environment. Selection of outer bay habitat could represent occasional foraging
2893 excursions in this habitat. Although outer bay habitats were available on both sides of

2894 Cleveland Bay, *C. fitzroyensis* rarely use outer bay habitats on the western side of the bay. This
2895 would suggest that, regardless of why outer bay habitat was used, individuals preferred to
2896 remain in close proximity to seagrass habitat. Consistent avoidance of sandy inshore, reef and
2897 mudflat habitat suggests these habitats were not suitable for *C. fitzroyensis*.

2898 This study also revealed there was individual variability in presence. A few individuals spent
2899 less than two weeks in the bay, suggesting broader movements occur in a portion of the
2900 population. Further evidence of broad movement came from an individual that was recaptured
2901 36 km from Cleveland Bay a few weeks after release. Individual variability in presence has
2902 been reported in other elasmobranchs, such as the medium-bodied *C. sorrah* (Knip *et al.*
2903 2012a) and the cownose ray *Rhinoptera bonasus* (Collins *et al.* 2007). The low residency
2904 exhibited by some *C. fitzroyensis* are more similar to the movement patterns exhibited by
2905 small-bodied species such as *R. terraenovae* (Carlson *et al.* 2008). Therefore, some *C.*
2906 *fitzroyensis* individuals may have used more transitory strategies to gain greater access to a
2907 wider range of resources, such as prey and potential mates.

2908

2909 The space use patterns exhibited by *C. fitzroyensis* may have also been motivated by prey
2910 availability. The short-term change in activity space and location observed in August and May
2911 suggests a biotic or behavioural stimulus that caused changes in distribution to occur over a
2912 short period of time. If *C. fitzroyensis* were responding to changes in abiotic factors, changes in
2913 space use would likely mimic the more gradual pattern of changes in bay temperature and
2914 salinity. Therefore, the short term change in space use may have been a response to a short-
2915 lived biotic factor, such as a change in prey location and abundance. Diel patterns in space use
2916 may also be linked to foraging behaviour. Larger day time activity spaces suggest *C.*
2917 *fitzroyensis* were more active during the day and may be primarily diurnal feeders. This

2918 contrasts with what has been found in other elasmobranchs that typically utilize small core
2919 areas during the day and move more widely at night for feeding (Sundström *et al.* 2001;
2920 Cartamil *et al.* 2003; Garla *et al.* 2006b).

2921

2922 The results of this study have shown that *C. fitzroyensis* exhibited resource use and movement
2923 strategies that are often associated with both small- and large-bodied species. While immature
2924 and mature age classes shared space and resources, the population was highly resident to the
2925 bay. As previously discuss, the high residency displayed by some individuals may be highly
2926 beneficial.. However, the habitat use strategies of *C. fitzroyensis* could make them more
2927 vulnerable to environmental change. The regular movement of small-bodied sharks between
2928 different bays and/or habitats often means they are resilient to local environmental change
2929 (Knip *et al.* 2010; Yates *et al.* 2012). However, the high residency and strong association with
2930 seagrass habitat makes *C. fitzroyensis* vulnerable to local nearshore environmental change,
2931 specifically deterioration of seagrass habitat. Seagrass habitat abundance and biodiversity is
2932 under threat of decline from numerous human activities, including dredging (Walker and
2933 McComb 1992; Erftemeijer and Lewis III 2006), excessive nutrient deposition (McGlathery
2934 2001; Ralph *et al.* 2006), chemical contamination such as herbicides (Haynes *et al.* 2000), and
2935 the effects of climate change (Orth *et al.* 2006; Hughes *et al.* 2008; Waycott *et al.* 2009). As
2936 immature and mature individuals share space and resources, declines in seagrass availability
2937 would likely affect all age classes within the population. This chapter has demonstrated that
2938 medium-bodied coastal species use nearshore habitats differently compared to other size classes
2939 of sharks. Moreover, the movement patterns of medium-bodied species could increase their
2940 vulnerability to environmental alterations. Therefore, future research should strive to increase
2941 understanding of medium-bodied species and the evolutionary causes, costs, and benefits of
2942 their behaviours.

Chapter 8

General Discussion

8.1 Conclusions and Implications

Nearshore areas are highly dynamic, productive environments that contain key habitats for many marine species, including sharks (Beck *et al.* 2001; Knip *et al.* 2010). However, nearshore areas are exposed to a variety of destructive anthropogenic influences, such as human development, pollution, and inshore fishing (Lotze *et al.* 2006; Bulleri and Chapman 2010; Breen *et al.* 2014). Consequently, nearshore shark species may be susceptible to population decline as a result of environmental change. Therefore, data on the residency, space use, resource selection, and specialisation of nearshore sharks are critical to species management. However, varying definitions of shark resource use, specifically resource specialisation, have limited research in this area. Without a consistent, widely applicable definition to describe the resource use of shark species, researchers are unable to compare results and may misinterpret findings. The definition for shark resource specialisation proposed in Chapter 2 provides a conceptual context for measuring and interpreting shark resource specialisation in a variety of circumstances. The “requirements only” (Peterson *et al.* 2011) definition of niche breadth proposed by Hutchinson (1957) best suits the current technological constraints of marine research and most directly addresses the primary questions of most resource use investigations. Moreover, the definition provided uses a continuum model for species comparison that more accurately reflects the role of species in nature. The definition is also flexible enough to incorporate unique environmental circumstances and scales that vary between studies and species. As a result, specialisation and vulnerability can be considered at a scale and context appropriate to the focal species and

2968 environment. By applying a consistent but widely applicable definition for shark resource
2969 specialisation it was possible to assess and compare the diet, habitat use, and level of
2970 specialisation of *R. taylori* and *C. fitzroyensis*.

2971

2972 Based on the collective findings of Chapters 4, 5, and 6, *R. taylori* is best described as a
2973 species with a low degree of resource specialisation that is adaptive to local environmental
2974 change. Results showed that *R. taylori* used different habitats and embayments, which is
2975 consistent with other small-bodied species. The diet of *R. taylori* included prey from seagrass
2976 and plankton food chains. Geographic variation in *R. taylori* diet also suggested that
2977 populations were foraging opportunistically. Thus, if a particular bay or habitat was in
2978 decline, it is likely that *R. taylori* could move into another bay or habitat and successfully use
2979 the resources there. Previous work has also shown that *R. taylori* is a highly productive
2980 species (Simpfendorfer 1992b). The combination of *R. taylori* movement patterns, diet, and
2981 productivity mean this species would be resilient to nearshore environmental change.
2982 However, large-scale decreases in seagrass availability may affect *R. taylori* behaviour.
2983 Stable isotope analysis revealed *R. taylori* had a smaller coastal range than was expected
2984 based on the movement of similar small-bodied species (e.g. *Rhizoprionodon terraenovae*)
2985 (Carlson *et al.* 2008; Kohler *et al.* 1998). Dietary analysis also suggested seagrass habitat was
2986 important for *R. taylori* throughout its range. Therefore, although small-bodied coastal
2987 species are often considered less vulnerable to environmental change, loss of seagrass over a
2988 broad spatial scale could be problematic for *R. taylori*. If there was a decline in seagrass
2989 availability and/or benthic productivity across multiple bays, *R. taylori* may have to use
2990 different habitats (e.g. sandy inshore) more often and/or expand its range to ensure sufficient
2991 access to resources. Prey abundance in seagrass would decline with loss of habitat, and as a
2992 result *R. taylori* may need to increase consumption of prey from plankton food chains. A

2993 reduction in seagrass availability and/or benthic productivity may also affect population
2994 fitness, but this would be dependent on other factors such as competition for reduced
2995 resources.

2996

2997 In contrast to *R. taylori*, the resource use patterns of *C. fitzroyensis* suggested this species has
2998 a moderately high degree of habitat specialisation. Immature and mature *C. fitzroyensis*
2999 demonstrated consistent selection for seagrass habitat and high residency to Cleveland Bay.
3000 Although there was some individual variability in presence and evidence for long range
3001 movement, the majority of individuals exhibited high residency. Coastal catch records also
3002 suggest that *C. fitzroyensis* prefers mud substrate and seagrass habitats throughout its range
3003 (Simpfendorfer unpublished data). Therefore, unlike *R. taylori*, there is little evidence to
3004 suggest that *C. fitzroyensis* would be able to efficiently use other habitats if seagrass habitats
3005 were no longer available. These results indicate that *C. fitzroyensis* may be vulnerable to
3006 population decline as a result of nearshore change, in particular the destruction of seagrass
3007 habitat. Therefore, although both species occupy nearshore areas throughout their lives and
3008 exhibit a preference for seagrass habitat, the differences in habitat use and movement indicate
3009 that *C. fitzroyensis* is more vulnerable to environmental change than *R. taylori*.

3010

3011 As discussed in Chapters 4 and 7, differences in movement strategies may be a result of
3012 contrasting life histories. The small-bodied *R. taylori* is a highly productive, fast-growing
3013 species (Simpfendorfer 1992b; Last and Stevens 2009). Due to its small size, *R. taylori* is
3014 vulnerable to predation inside and outside of nearshore areas (see Chapter 4), and thus may
3015 ultimately benefit from moving between embayments to gain access to more resources. In
3016 comparison, *C. fitzroyensis* is slower-growing and less productive (Lyle 1987; Smart *et al.*

3017 2013), therefore it may derive greater benefit from residency in highly productive habitats,
3018 similar to juveniles of large-bodied sharks (e.g. Conrath and Musick 2010). However,
3019 differences in movement patterns observed between these species are likely based on a
3020 variety of complex biological factors that require further investigation.

3021

3022 Research in Cleveland Bay has shown that nearshore sharks can exhibit a diverse range of
3023 movement and resource use strategies (Knip *et al.* 2011; Knip,*et al.* 2012; Chin *et al.* 2013).

3024 These strategies will affect species exposure and vulnerability to environmental change.

3025 Movement and resource use data can also be used to identify key habitats and locations for
3026 different populations and species. Improved knowledge of the resource selection and

3027 specialisation of nearshore shark species will result in a better understanding of species and

3028 enhance management strategies. For example, both *R. taylori* and *C. fitzroyensis* preferred

3029 seagrass habitat. As such, protection of seagrass will likely be an important consideration in

3030 spatial management of these species if required. This dissertation also demonstrated that the

3031 resource use patterns of immature and mature individuals may affect the population's overall

3032 response to change. For example, consistent use of seagrass by immature and mature *C.*

3033 *fitzroyensis* suggests that both groups will be negatively affected by declines in seagrass

3034 availability. Therefore it is important to consider movement and habitat use across multiple

3035 age classes to gain a population level understanding of resource requirements.

3036

3037 This dissertation has demonstrated the importance of defining the movement and resource use

3038 patterns of sharks in nearshore areas. Species resource use patterns are highly diverse and

3039 dependent on life history and environmental circumstance. Species that are more specialised

3040 (i.e. *C. fitzroyensis*) will likely prove more vulnerable to environmental change or degradation

3041 than species that have a low degree of resource specialisation (i.e. *R. taylori*). However, as
3042 demonstrated by *R. taylori*, even species that have a low degree of specialisation may be
3043 negatively affected by changes to the environment. As human development in coastal areas
3044 increases, the level of resource specialisation exhibited by species will be a critical
3045 component of habitat and species management plans.

3046

3047 **8.2 Future Research**

3048

3049 Although the work described in this dissertation has increased understanding of nearshore
3050 sharks, it has also highlighted topics that require further study. It is still unclear how changes
3051 in abiotic factors such as water temperature, oxygen content, or salinity affect *R. taylori* and
3052 *C. fitzroyensis* habitat use and movement. The use of sandy inshore habitat by *R. taylori* was
3053 correlated with changes in freshwater input into Cleveland Bay, however this correlation was
3054 inconsistent among individuals. It is possible that movement into sandy habitats was not a
3055 physiological response to changes in freshwater input but the result of another driver. For
3056 example, some *R. taylori* may have moved into sandy inshore areas to give birth. Therefore,
3057 having more detailed knowledge of how species respond to changes in environmental factors
3058 and what role biological drivers play is important to understanding population level
3059 movements and vulnerabilities.

3060

3061 As discussed in Chapter 3, isotope analysis is not commonly used to study the movement of
3062 sharks. This is in part due to limiting factors such as long turnover times in tissues (Logan
3063 and Lutcavage 2010). Studies that have used isotopes to examine the movement of sharks
3064 have primarily investigated movement at broad or coarse spatial scales (e.g. Abrantes and

3065 Barnett 2011; Carlisle *et al.* 2012). However, the techniques used in Chapter 3 to assess the
3066 annual range of *R. taylori* demonstrated that isotopes can be used to define movement at
3067 regional scales in relatively high detail. In future, isotope analysis could be used to gain a
3068 better understanding of shark movement as well as diet.

3069

3070 Due to its relative rarity, it was not possible to sample sufficient *C. fitzroyensis* for stable
3071 isotope movement and diet analysis. However, knowledge of *C. fitzroyensis* broader scale
3072 movements and diet would greatly enhance our current understanding of this species. For
3073 example, isotope analysis could be used to determine if seagrass or benthic food chains are
3074 the primary contributor to *C. fitzroyensis* diet, as the telemetry data suggests. Previous isotope
3075 analysis in Cleveland Bay suggests that *C. fitzroyensis* has higher $\delta^{13}\text{C}$ values than *R. taylori*
3076 and several other species of sharks and teleosts (Kinney *et al.* 2011). This could indicate *C.*
3077 *fitzroyensis* feeds from benthic or seagrass food chains more often than *R. taylori*. This
3078 interpretation is supported by the telemetry results in Chapters 4 and 7, which demonstrated
3079 *C. fitzroyensis* used seagrass habitats more consistently than *R. taylori*. However, the sample
3080 size of *C. fitzroyensis* in the previous study was small (n=9; Kinney *et al.* 2011), therefore
3081 further study is needed to comprehensively assess *C. fitzroyensis* diet. Given that *C.*
3082 *fitzroyensis* was highly resident to Cleveland Bay, the inclusion of additional isotopes in
3083 analysis, such as sulphur, may improve assessment of coastal movement and habitat use
3084 (Connolly *et al.* 2004). The potential for more distinct coastal patterns in $\delta^{34}\text{S}$ compared to
3085 $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ would make it easier to detect *C. fitzroyensis* movement between embayments,
3086 even over long periods of time. Including tissues that have relatively fast isotope turnover
3087 rates, such as liver (MacNeil *et al.* 2006), would also make it possible to detect seasonal
3088 changes in *C. fitzroyensis* diet. Plasma and muscle turnover rates in medium-bodied sharks

3089 would likely be too slow to detect changes in diet between seasons (Logan and Lutcavage
3090 2010; Kim *et al.* 2012).

3091

3092 Movement and residency patterns exhibited by *R. taylori* were mostly consistent with those
3093 exhibited by other small-bodied sharks (Carlson *et al.* 2008; Espinoza *et al.* 2011).

3094 Collectively, these results suggest that small-bodied sharks are generally highly mobile and
3095 use a wide array of habitats. However, the results of Chapter 7 suggest that medium-bodied
3096 species exhibit unique nearshore movement and habitat use strategies compared to small- and
3097 large-bodied sharks. Medium-bodied species may exhibit resource use and movement
3098 patterns better suited to their biological traits. Unfortunately, there has been little research on
3099 the movement of medium-bodied nearshore sharks. The results of this dissertation indicate
3100 that more focused study on medium-bodied species is required as these species likely have a
3101 distinct relationship with the environment and specific vulnerabilities to change. Future work
3102 should also focus on developing theoretical explanations for medium-bodied shark behaviour.

3103

References

- 3104
3105
- 3106 Aalbers, S.A., Bernal, D., and Sepulveda, C.A. (2010). The functional role of the caudal fin in
3107 the feeding ecology of the common thresher shark *Alopias vulpinus*. *Journal of Fish*
3108 *Biology* **76**,1863-1868. doi:10.1111/j.1095-8649.2010.02616.x
- 3109 Abecasis, D., and Erzini, K. (2008). Site fidelity and movements of gilthead sea bream
3110 (*Sparus aurata*) in a coastal lagoon (Ria Formosa, Portugal). *Estuarine, Coastal and*
3111 *Shelf Science* **79**, 758-763.
- 3112 Abrantes, K.G., and Barnett, A. (2011). Intrapopulation variations in diet and habitat use in a
3113 marine apex predator, the broadnose sevengill shark *Notorynchus cepedianus*. *Marine*
3114 *Ecology Progress Series* **442**, 133-148. doi 10.3354/meps09395
- 3115
- 3116 Alerstam, T., Hedenström, A., and Åkesson, S. (2003). Long-distance migration: evolution
3117 and determinants. *Oikos* **103**, 247-260. doi 10.1034/j.1600-0706.2003.12559.x
- 3118
- 3119 Alonso, M.K., Crespo, E.A., García, N.A., Pedraza, S.N., Mariotti, P.A., and Mora, J.N.
3120 (2002). Fishery and ontogenetic driven changes in the diet of the spiny dogfish,
3121 *Squalus acanthias*, in Patagonian waters, Argentina. *Environmental Biology of Fishes*
3122 **63**, 193-202. doi:10.1023/a:1014229432375
- 3123
- 3124 Araújo, M.B., Thuiller, W., and Pearson, R.G. (2006). Climate warming and the decline of
3125 amphibians and reptiles in Europe. *Journal of Biogeography* **33**, 1712-1728. doi
3126 10.1111/j.1365-2699.2006.01482.x
- 3127
- 3128 Arlettaz, R. (1999). Habitat selection as a major resource partitioning mechanism between the
3129 two sympatric sibling bat species *Myotis myotis* and *Myotis blythii*. *Journal of Animal*
3130 *Ecology* **68**, 460-471.
- 3131
- 3132 Avsar, D. (2001). Age, growth, reproduction and feeding of the spurdog (*Squalus acanthias*
3133 Linnaeus, 1758) in the south-eastern Black Sea. *Estuarine, Coastal, and Shelf Science*
3134 **52**, 269-278. doi:http://dx.doi.org/10.1006/ecss.2000.0749
- 3135
- 3136 Ba, B.A., Diop, M.S., Diatta, Y., Justine, D., and Ba, C.T. (2013). Diet of the milk shark,
3137 *Rhizoprionodon acutus* (Chondrichthyes: Carcharhinidae), from the Senegalese coast.
3138 *Journal of Applied Ichthyology* **2013**, 1-7. doi:10.1111/jai.12156
- 3139
- 3140 Backwell, P.R.Y., O'Hara, P.D., and Christy, J.H. (1998). Prey availability and selective
3141 foraging in shorebirds. *Animal Behaviour* **55**,1659-1667. doi:10.1006/anbe.1997.0713
- 3142
- 3143 Baremore, I.E., Murie, D.J., and Carlson, J.K. (2008). Prey selection by the Atlantic angel
3144 shark *Squatina dumeril* in the Northeastern Gulf of Mexico. *Bulletin of Marine*
3145 *Science* **82**, 297-313.
- 3146
- 3147 Basille, M., Calenge, C., Marboutin, É., Andersen, R., and Gaillard, J-M. (2008). Assessing
3148 habitat selection using multivariate statistics: Some refinements of the ecological-
3149 niche factor analysis. *Ecology Modelling* **211**,233-240.
3150 doi:http://dx.doi.org/10.1016/j.ecolmodel.2007.09.006

- 3151 Beck, M.W., Heck, K.L., Able, K.W., Childers, D.L., Eggleston, D.B., Gillanders, B.M.,
3152 Halpern, B., Hays, C.G., Hoshino, K., Minello, T.J., Orth, R.J., Sheridan, P.F., and
3153 Weinstein, M.P. (2001). The identification, conservation, and management of
3154 estuarine and marine nurseries for fish and invertebrates. *BioScience* **51**, 633-641.
3155
- 3156 Belleggia, M., Figueroa, D., Sánchez, F., and Bremec, C. (2012). The feeding ecology of
3157 *Mustelus schmitti* in the southwestern Atlantic: geographic variations and dietary
3158 shifts. *Environmental Biology of Fishes* **95**, 99-114.
3159
- 3160 Ben-David, M., Flynn, R.W., and Schell, D.M. (1997). Annual and seasonal changes in diets
3161 of martens: evidence from stable isotope analysis. *Oecologia* **111**, 280-291.
3162
- 3163 Berenbaum, M.R. (1996). Introduction to the symposium: on the evolution of specialization.
3164 *The American Naturalists*, **148**, 78-83. doi:10.2307/2463049
3165
- 3166 Berger, J.O. (2006). The case for objective Bayesian analysis. *Bayesian Analysis* **1**, 385-402.
3167
- 3168 Bernardo, J.M., and Smith A.F.M. (1994). 'Bayesian Theory'. (John Wiley & Sons: New
3169 York, NY.)
3170
- 3171 Bernays, E.A., and Funk, D.J. (1999). Specialists make faster decisions than generalists:
3172 experiments with aphids. *Proceedings of the Royal Society B: Biological Sciences*
3173 **266**, 151-156. doi:10.1098/rspb.1999.0615
3174
- 3175 Bernays, E.A., and Wcislo, W.T. (1994). Sensory capabilities, information processing, and
3176 resource specialization. *Quarterly Review of Biology* **69**, 187-204.
3177
- 3178 Bethea, D., Hale, L., Carlson, J., Cortés, E., Manire, C., and Gelsleichter, J. (2007).
3179 Geographic and ontogenetic variation in the diet and daily ration of the bonnethead
3180 shark, *Sphyrna tiburo*, from the eastern Gulf of Mexico. *Marine Biology* **152**, 1009-
3181 1020. doi:10.1007/s00227-007-0728-7
3182
- 3183 Bethea, D.M., Carlson, J.K., Buckel, J.A., and Satterwhite, M. (2006). Ontogenetic and site-
3184 related trends in the diet of the Atlantic sharpnose shark *Rhizoprionodon terraenovae*
3185 from the northeast Gulf of Mexico. *Bulletin of Marine Science* **78**, 287-307.
3186
- 3187 Blaber, S.J.M., Cyrus, D.P., Albaret, J.-J., Ching, C.V., Day, J.W., Elliott, M., Fonseca, M.S.,
3188 Hoss, D.E., Orensanz, J., Potter, I.C., and Silvert, W. (2000). Effects of fishing on the
3189 structure and functioning of estuarine and nearshore ecosystems. *ICES Journal of*
3190 *Marine Science: Journal du Conseil* **57**, 590-602.
3191
- 3192 Bligh, E.G., and Dyer, W.J. (1959). A rapid method of total lipid extraction and purification.
3193 *Canadian Journal Biochemistry Physiology* **37**, 911-917.
3194
- 3195 Bolnick, D.I., Svanbäck, R., Fordyce, J.A., Yang, L.H., Davis, J.M., Hulsey, C.D., and
3196 Forister, M.L. (2003). The ecology of individuals: incidence and implications of
3197 individual specialization. *The American Naturalist* **161**, 1-28.
3198

- 3199 Bolnick, D.I., Yang, L.H., Fordyce, J.A., Davis, J.M., and Svanback, R. (2002). Measuring
3200 individual-level resource specialisation. *Ecology* **10**, 2936-2941.
3201
- 3202 Bonfil, R., Mejer, M., Scholl, M.C., Johnson, R., O'Brien, S., Oosthuizen, H., Swanson, S.,
3203 Kotze, D., and Paterson, M. (2005). Transoceanic migration, spatial dynamics, and
3204 population linkages of white sharks. *Science* **310**, 100-103.
3205 doi:10.1126/science.1114898
3206
- 3207 Bornatowski, H., Heithaus, M.R., Abilhoa, V., and Corrêa, M.F.M. (2012). Feeding of the
3208 Brazilian sharpnose shark *Rhizoprionodon lalandii* (Müller & Henle, 1839) from
3209 southern Brazil. *Journal of Applied Ichthyology* **28**, 623-627.
3210
- 3211 Boustany, A.M., Davis, S.F., Anderson, S.D., Le Boeuf, B.J., and Block, B.A. (2002).
3212 Satellite tagging: expanded niche for white sharks. *Nature* **415**, 35-36.
3213
- 3214 Boutton, T. (1991). Stable carbon isotope ratios of natural materials: II. Atmospheric,
3215 terrestrial, marine, and freshwater environments. In 'Carbon Isotope Techniques'. (Eds
3216 D.C. Coleman and B. Fry.), pp 173-176. (Academic Press Inc: San Diego, CA.)
3217
- 3218 Branstetter, S. (1990). Early life-history implications of selected carcharhinoid and lamnoid
3219 sharks of the northwest Atlantic. *NOAA Tech. Rep. NMFS* **90**, 17-28.
3220
- 3221 Breitburg, D.L. (1990). Near-shore hypoxia in the Chesapeake Bay: Patterns and relationships
3222 among physical factors. *Estuarine, Coastal and Shelf Science* **30**, 593-609.
3223
3224
- 3225 Brischoux, F., Bonnet, X., Cherel, Y., and Shine, R. (2011). Isotopic signatures, foraging
3226 habitats and trophic relationships between fish and seasnakes on the coral reefs of
3227 New Caledonia. *Coral Reefs* **30**, 155-165. doi:10.1007/s00338-010-0680-8
3228
- 3229 Britt, E.J., Hicks, J.W., and Bennett, A.F. (2006). The energetic consequences of dietary
3230 specialization in populations of the garter snake, *Thamnophis elegans*. *Journal of*
3231 *Experimental Biology* **209**, 3164-3169. doi:10.1242/jeb.02366
3232
- 3233 Brodeur, R.D., Fleming, I.A., Bennett, J.M., and Campbell, M.A. (2009). Summer
3234 distribution and feeding of spiny dogfish off the Washington and Oregon coasts. In:
3235 'Biology and management of dogfish sharks'. (Eds V.F. Gallucci, G.A. McFarland,
3236 and G.G. Bargmann.) pp. 39-51. (American Fisheries Society: Bethesda.)
3237
- 3238 Bruce, B.D. (2008). The biology and ecology of the white shark, *Carcharodon carcharias*. In
3239 'Sharks of the open ocean: biology, fisheries and conservation'. (Eds M.D. Camhi,
3240 E.K. Pikitch, and E.A. Babcock.) pp.69-76. (Blackwell Publishing Ltd: Oxford, U.K.)
3241
- 3242 Buchheister, A., and Latour, R.J. (2010). Turnover and fractionation of carbon and nitrogen
3243 stable isotopes in tissues of a migratory coastal predator, summer flounder
3244 (*Paralichthys dentatus*). *Canadian Journal Fisheries and Aquaculture Science* **67**,
3245 445-461. doi 10.1139/f09-196
3246
- 3247 Bulleri, F., and Chapman, M.G. (2010). The introduction of coastal infrastructure as a driver
3248 of change in marine environments. *Journal of Applied Ecology* **47**, 26-35.

3249
3250
3251 Buskirk, S.W., and Millsaugh, J.J. (2006). Metrics for studies of resource selection. *Journal*
3252 *of Wildlife Management* **70**, 358-366.
3253
3254 Cabrera-Chávez-Costa, A.A., Galván-Magaña, F., and Escobar-Sánchez, O. (2010). Food
3255 habits of the silky shark *Carcharhinus falciformis* (Müller & Henle, 1839) off the
3256 western coast of Baja California Sur, Mexico. *Journal of Applied Ichthyology* **26**, 499-
3257 503. doi:10.1111/j.1439-0426.2010.01482.x
3258
3259 Calenge, C. (2006). The package adehabitat for the R software: a tool for the analysis of space
3260 and habitat use by animals. *Ecological Modelling* **197**, 516-519.
3261
3262 Campana, S.E., and Joyce, W.N. (2004). Temperature and depth associations of porbeagle
3263 shark (*Lamna nasus*) in the northwest Atlantic. *Fisheries Oceanography* **13**, 52-64.
3264 doi:10.1111/j.1365-2419.2004.00236.x
3265
3266 Carleton, S.A., and Del Rio, C.M. (2010). Growth and catabolism in isotopic incorporation: a
3267 new formulation and experimental data. *Functional Ecology* **24**, 805-812. doi
3268 10.1111/j.1365-2435.2010.01700.x
3269
3270 Carlisle, A.B., and Starr, R.M. (2009). Habitat use, residency, and seasonal distribution of
3271 female leopard sharks *Triakis semifasciata* in Elkhorn Slough, California. *Marine*
3272 *Ecology Progress Series* **380**, 213-228. doi:10.3354/meps07907
3273
3274 Carlisle, A.B., Kim, S.L., Semmens, B.X., Madigan, D.J., Jorgensen, S.J., Perle, C.R.,
3275 Anderson, S.D., Chapple, T.K., Kanive, P.E., and Block, B.A. (2012). Using stable
3276 isotope analysis to understand the migration and trophic ecology of northeastern
3277 Pacific white sharks (*Carcharodon carcharias*). *PLoS ONE* **7**:e30492.
3278 doi:10.1371/journal.pone.0030492
3279
3280 Carlson, J., Heupel, M., Bethea, D., and Hollensead, L. (2008). Coastal habitat use and
3281 residency of juvenile Atlantic sharpnose sharks (*Rhizoprionodon terraenovae*).
3282 *Estuaries and Coasts* **31**, 931-940.
3283
3284 Carlton, J.T., Vermeij, G.J., Lindberg, D.R., Carlton, D.A., and Dubley, E.C. (1991). The first
3285 historical extinction of a marine invertebrate in an ocean basin: The demise of the
3286 eelgrass limpet *Lottia alveus*. *The Biological Bulletin* **180**, 72-80.
3287
3288 Cartamil, D., Wegner, N.C., Aalbers, S., Sepulveda, C.A., Baquero, A., and Graham, J.B.
3289 (2010). Diel movement patterns and habitat preferences of the common thresher shark
3290 (*Alopias vulpinus*) in the Southern California Bight. *Marine and Freshwater Research*
3291 **61**, 596-604. doi:http://dx.doi.org/10.1071/MF09153
3292
3293 Cartamil, D.P., Vaudo, J.J., Lowe, C.G., Wetherbee, B.M., and Holland, K.N. (2003). Diel
3294 movement patterns of the Hawaiian stingray, *Dasyatis lata*: implications for
3295 ecological interactions between sympatric elasmobranch species. *Marine Biology* **142**,
3296 841-847.
3297
3298

- 3299 Castro, J.I. (1993) The shark nursery of Bulls Bay, South Carolina, with a review of the shark
 3300 nurseries of the southeastern coast of the United States. *Environmental Biology of*
 3301 *Fishes* **38**, 37-48.
 3302
- 3303 Chapman, C.A., and Mackay, W.C. (1984). Versatility in habitat use by a top aquatic
 3304 predator, *Esox lucius* L. *Journal of Fish Biology* **25**,109-115. doi:10.1111/j.1095-
 3305 8649.1984.tb04855.x
 3306
- 3307 Chapman, D.D., Pikitch, E.K., Babcock, E.A., and Shivji, M.S. (2007). Deep-diving and diel
 3308 changes in vertical habitat use by Caribbean reef sharks *Carcharhinus perezii*. *Marine*
 3309 *Ecology Progress Series* **344**, 271-275.
 3310
- 3311 Chase, J.M., and Leibold, M.A. (2009). ' Ecological niches: linking classic and contemporary
 3312 approaches'. (University of Chicago Press: Chicago, IL).
 3313
- 3314 Chin, A., Heupel, M.R., Simpfendorfer, C.A., and Tobin, A.J. (2013). Ontogenetic
 3315 movements of juvenile blacktip reef sharks: evidence of dispersal and connectivity
 3316 between coastal habitats and coral reefs. *Aquatic Conservation* **23**, 468-474. doi
 3317 10.1002/aqc.2349
 3318
- 3319 Chin, A., Kyne, P.M., Walker, T.I., and McAuley, R.B. (2010). An integrated risk assessment
 3320 for climate change: analysing the vulnerability of sharks and rays on Australia's Great
 3321 Barrier Reef. *Global Change Biology* **16**, 1936-1953.
 3322
- 3323 Chin, A., Tobin, A., Simpfendorfer, C., and Heupel, M. (2012). Reef sharks and inshore
 3324 habitats: patterns of occurrence and implications for vulnerability. *Marine Ecology*
 3325 *Progress Series* **460**, 115-125.
 3326
- 3327 Clarke, A., and Leakey, R.J. (1996) The seasonal cycle of phytoplankton, macronutrients, and
 3328 the microbial community in a nearshore Antarctic marine ecosystem. *Limnology and*
 3329 *Oceanography* **41**(6), 1281-1294.
 3330
- 3331 Clarke, E., and Nelson, D.R. (1997). Young whale sharks, *Rhincodon typus*, feeding on a
 3332 copepod bloom near La Paz, Mexico. *Environmental Biology of Fishes* **50**, 63-73.
 3333
- 3334 Clarke, C., Lea, J.S.E., and Ormond, R.F.G. (2011). Reef-use and residency patterns of a
 3335 baited population of silky sharks, *Carcharhinus falciformis*, in the Red Sea. *Marine*
 3336 *and Freshwater Research* **62**, 668-675.
 3337
- 3338 Clavel, J., Julliard, R., and Devictor, V. (2010). Worldwide decline of specialist species:
 3339 toward a global functional homogenization? *Frontiers in Ecology and the*
 3340 *Environment* **9**, 222-228.
 3341
- 3342 Colles, A., Liow, L.H., and Prinzing, A. (2009). Are specialists at risk under environmental
 3343 change? Neocological, paleoecological and phylogenetic approaches. *Ecology*
 3344 *Letters* **12**, 849-863.
 3345
- 3346 Collins, A.B., Heupel, M.R., and Motta, P.J. (2007). Residence and movement patterns of
 3347 cownose rays *Rhinoptera bonasus* within a south-west Florida estuary. *Journal of*
 3348 *Fish Biology* **71**, 1159-1178.

- 3349
3350 Colman, J.G. (1997). A review of the biology and ecology of the whale shark. *Journal of Fish*
3351 *Biology* **51**, 1219-1234.
3352
- 3353 Colwell, R.K., and Futuyma, D.J. (1971). On the measurement of niche breadth and overlap.
3354 *Ecology* **52**, 567-576. doi:10.2307/1934144
- 3355 Compagno, L.J.V. (1984). Vol.4. Sharks of the world. An annotated and illustrated catalogue of
3356 sharks species known to date. Part 2, Carcharhiniformes. *FAO Fisheries Synopsis* **125**,
3357 251-655.
3358
- 3359 Compagno LV (1990). Alternative life-history styles of cartilaginous fishes in time and space.
3360 *Environmental Biology of Fishes* **28**, 33-75. doi:10.1007/bf00751027
- 3361 Compagno LJV (2001). Sharks of the world. An annotated and illustrated catalogue of shark
3362 species known to date. vol. 2. Bullhead, mackerel and carpet sharks (Heterodontiformes,
3363 Lamniformes and Orectolobiformes). pp. 269 (FAO Species Catalogue for Fisheries
3364 Purposes: Rome.)
3365
- 3366 Connolly, R.M. (1994). Removal of seagrass canopy: effects on small fish and their prey.
3367 *Journal of Experimental Marine Biology and Ecology* **184**, 99-110.
3368
- 3369 Connolly, R.M., Guest, M.A., Melville, A.J., and Oakes, J.M. (2004). Sulfer stable istopes
3370 separate produces in marine food-web analysis. *Oecologica* **138**, 161-167.
3371
- 3372 Conrath, C.L., and Musick, J.A. (2010). Residency, space use and movement patterns of
3373 juvenile sandbar sharks (*Carcharhinus plumbeus*) within a Virginia summer nursery
3374 area. *Marine and Freshwater Research* **61**, 223-235.
3375
- 3376 Cortés, E., and Gruber, S.H. (1990). Diet, feeding habits and estimates of daily ration of
3377 young lemon Sharks, *Negaprion brevirostris* (Poey). *Copeia* **1990**, 204-218.
3378
3379
- 3380 Curtis, T.H., Parkyn, D.C., and Burgess, G.H. (2013). Use of human-altered habitats by bull
3381 sharks in a Florida nursery area. *Marine and Coastal Fisheries* **5**, 28-38.
3382
- 3383 Dale, J.J., Wallsgrove, N.J., Popp, B.N., and Holland, K.N. (2011). Nursery habitat use and
3384 foraging ecology of the brown stingray *Dasyatis lata* determined from stomach
3385 contents, bulk and amino acid stable isotopes. *Marine Ecology Progress Series* **433**,
3386 221-236. doi 10.3354/meps09171
3387
- 3388 Dalerum, F., and Angerbjörn, A. (2005). Resolving temporal variation in vertebrate diets
3389 using naturally occurring stable isotopes. *Oecologia* **144**, 647-658. doi
3390 10.1007/s00442-005-0118-0
3391
- 3392 Dean, M.N., Wilga, C.D., and Summers, A.P. (2005). Eating without hands or tongue:
3393 specialization, elaboration and the evolution of prey processing mechanisms in
3394 cartilaginous fishes. *Biology Letters* **1**, 357-361. doi:10.1098/rsbl.2005.0319
3395
- 3396 DeNiro, M.J., and Epstein, S. (1978). Influence of diet on the distribution of carbon isotopes
3397 in animals. *Geochimica et Cosmochimica Acta* **42**, 495-506

3398
3399 DeNiro, M.J., and Epstein, S. (1981). Influence of diet on the distribution of nitrogen isotopes
3400 in animals. *Geochimica et Cosmochimica* **45**, 341-351.
3401
3402 Devictor, V., Clavel, J., Julliard, R., Lavergne, S., Mouillot, D., Thuiller, W., Venail, P.,
3403 Villéger, S., and Mouquet, N. (2010). Defining and measuring ecological
3404 specialization. *Journal of Applied Ecology* **47**, 15-25.
3405
3406 Devlin, M., Brodie, J., Wenger, A., de Silva, E., Alvarez-Romero, J.G., Waterhouse, J., and
3407 McKenzie, L. (2012). Extreme weather conditions in the Great Barrier Reef: Drivers
3408 of change In 'Proceedings of the 12th International Coral Reef Symposium'. (Eds D.
3409 Yellowlees and T. P. Hughes.) pp. ICRS2012_21A_2. (ARC Center for Excellence
3410 for Coral Reef Studies: Carins, Australia.)
3411
3412 DeWitt, T.J. (1998). Costs and limits of phenotypic plasticity: tests with predator-induced
3413 morphology and life history in a freshwater snail. *Journal of Evolutionary Biology* **11**,
3414 465-480. doi:10.1046/j.1420-9101.1998.11040465.x
3415
3416 DeWitt, T.J., Sih, A., and Wilson, D.S. (1998). Costs and limits of phenotypic plasticity.
3417 *Trends in Ecology and Evolution* **13**, 77-81. doi:10.1016/s0169-5347(97)01274-3
3418
3419 Dicken, M.L., Booth, A.J., and Samle, M.J. (2008). Estimates of juvenile and adult
3420 raggedtooth shark (*Carcharias taurus*) abundance along the east coast of South
3421 Africa. *Canadian Journal of Fisheries and Aquaculture Sciences* **65**, 621-632.
3422
3423 DiGirolamo, A.L., Gruber, S.H., Pomory, C., and Bennett, W.A. (2012). Diel temperature
3424 patterns of juvenile lemon sharks *Negaprion brevirostris*, in a shallow-water nursery.
3425 *Journal of Fish Biology* **80**, 1436-1448.
3426
3427 Domi, N., Bouquegneau, J.M., and Das, K. (2005). Feeding ecology of five commercial shark
3428 species of the Cletic Sea through stable isotope and trace metal analysis. *Marine*
3429 *Environmental Research* **60**, 551-569.
3430 Drymon, J.M., Powers, S., and Carmichael, R. (2012). Trophic plasticity in the Atlantic
3431 sharpnose shark (*Rhizoprionodon terraenovae*) from the north central Gulf of Mexico.
3432 *Environmental Biology of Fishes* **95**:21-35.
3433
3434 Duarte, C.M. (2002). The future of seagrass meadows. *Environmental Conservation* **29**, 192-
3435 206.
3436
3437 Dulvy, N.K., Baum, J.K., Clarke, S., Compagno, L.J.V., Cortés, E., Domingo, A., Fordham,
3438 S., Fowler, S., Francis, M.P., Gibson, C., Martínez, J., Musick, J.A., Soldo, A.,
3439 Stevens, J.D., and Valenti, S. (2008). You can swim but you can't hide: the global
3440 status and conservation of oceanic pelagic sharks and rays. *Aquatic Conservation:*
3441 *Marine and Freshwater Ecosystems* **18**, 459-482. doi:10.1002/aqc.975
3442
3443 Dulvy, N.K., Ellis, J.R., Goodwin, N.B., Grant, A., Reynolds, J.D., and Jennings, S. (2004).
3444 Methods of assessing extinction risk in marine fishes. *Fish and Fisheries* **5**, 255-276.
3445

- 3446 Duncan, K.M., and Holland K.N. (2006). Habitat use, growth rates, and dispersal patterns of
 3447 juvenile scalloped hammerhead sharks (*Sphyrna lewini*) in a nursery habitat. *Marine*
 3448 *Ecology Progress Series* **312**, 211-221.
- 3449
- 3450 Ebert, D.A. (2003). 'Sharks, rays and chimaeras of California.' (University California Press:
 3451 Berkeley, California.)
- 3452
- 3453 Economakis, A., and Lobel, P. (1998). Aggregation behavior of the grey reef shark,
 3454 *Carcharhinus amblyrhynchos*, at Johnston Atoll, Central Pacific ocean.
 3455 *Environmental Biology of Fishes* **51**, 129-139. doi:10.1023/a:1007416813214
- 3456
- 3457 Edwards, M., Derocher, A., Hobson, K., Branigan, M., and Nagy, J. (2011). Fast carnivores
 3458 and slow herbivores: differential foraging strategies among grizzly bears in the
 3459 Canadian Arctic. *Oecologia* **165**, 877-889. doi:10.1007/s00442-010-1869-9
- 3460
- 3461 Edgar, G.J., and Shaw, C. (1995). The production and trophic ecology of shallow-water fish
 3462 assemblages in southern Australia III. General relationships between sediments,
 3463 seagrasses, invertebrates and fishes. *Journal of Experimental Marine Biology and*
 3464 *Ecology* **194**, 107-131.
- 3465
- 3466 Eide, N.E., Eid, P.M., Prestrud, P., and Swenson, J.E. (2005). Dietary responses of arctic
 3467 foxes *Alopex lagopus* to changing prey availability across an Arctic landscape.
 3468 *Wildlife Biology* **11**, 109-121.
- 3469
- 3470 Ellis, J.R., Pawson, M.G., Shackley, S.E. (1996). The comparative feeding ecology of six
 3471 species of shark and four species of ray (Elasmobranchii) in the north-east Atlantic.
 3472 *Journal of Marine Biological Association of the United Kingdom* **76**, 89-106.
- 3473
- 3474 Elton, C. (1927). Animal ecology In 'Niche: theory and application' (Eds R.Whittaker and S
 3475 Levin.) pp. 20-26 (Dowden, Hutchinson and Ross, Inc: Stroudsburg, Pennsylvania.)
- 3476
- 3477 Erftemeijer, P.L.A., and Lewis III, R.R. (2006). Environmental impacts of dredging on
 3478 seagrasses: A review. *Marine Pollution Bulletin* **52**, 1553-1572.
- 3479
- 3480 Espinoza, M., Farrugia, T.J., and Lowe, C.G. (2011). Habitat use, movements and site fidelity
 3481 of the gray smooth-hound shark (*Mustelus californicus* Gill 1863) in a newly restored
 3482 southern California estuary. *Journal of Experimental Marine Biology and Ecology*
 3483 **401**, 63-74. doi:10.1016/j.jembe.2011.03.001
- 3484
- 3485 Feinsinger, P., Spears, E.E., and Poole, R.W. (1981). A simple measure of niche breadth.
 3486 *Ecology* **62**, 27-32. doi:10.2307/1936664
- 3487
- 3488 Feldheim, K.A., Gruber, S.H., and Ashley, M.V. (2002). The breeding biology of lemon
 3489 sharks at a tropical nursery lagoon. *Proceedings of the Royal Society of London.*
 3490 *Series B: Biological Sciences* **269**, 1655-1661.
- 3491
- 3492 Fridley, J.D., Vandermast, D.B., Kuppinger, D.M., Manthey, M., and Peet, R.K. (2007). Co-
 3493 occurrence based assessment of habitat generalists and specialists: a new approach for
 3494 the measurement of niche width. *Journal of Ecology* **95**, 707-722. doi:10.1111/j.1365-
 3495 2745.2007.01236.x

3496
3497 Furnas, M.M. (2003). Catchments and corals: terrestrial runoff to the Great Barrier Reef.
3498 Australian Institute of Marine Science & CRC Reef Research Centre.
3499

3500 Futuyma, D.J., and Moreno, G. (1988). The evolution of ecological specialization. *Annual*
3501 *Review of Ecology and Systematics* **19**, 207-233.
3502

3503 Garla, R., Chapman, D., Wetherbee, B., and Shivji, M. (2006a). Movement patterns of young
3504 Caribbean reef sharks, *Carcharhinus perezi*, at Fernando de Noronha Archipelago,
3505 Brazil: the potential of marine protected areas for conservation of a nursery ground.
3506 *Marine Biology* **149**, 189-199.
3507

3508 Garla, R.C., Chapman, D.D., Shivji, M.S., Wetherbee, B.M., and Amorim, A.F. (2006b).
3509 Habitat of juvenile Caribbean reef sharks, *Carcharhinus perezi*, at two oceanic insular
3510 marine protected areas in the southwestern Atlantic Ocean: Fernando de Noronha
3511 Archipelago and Atol das Rocas, Brazil. *Fisheries Research* **81**, 236-241.
3512 doi:10.1016/j.fishres.2006.07.003
3513

3514 Gelman, A., and Hill, J. (2007). 'Data analysis using regression and multilevel/hierarchical
3515 models.' (Cambridge University Press: Cambridge UK.)
3516

3517 Gelsleichter, J., Musick, J.A., and Nichols, S. (1999). Food habits of the smooth dogfish,
3518 *Mustelus canis*, dusky shark, *Carcharhinus obscurus*, Atlantic sharpnose shark,
3519 *Rhizoprionodon terraenovae*, and the sand tiger, *Carcharias taurus*, from the
3520 northwest Atlantic Ocean. *Environmental Biology Fishes* **54**, 205-217.
3521

3522 Goldman, K., and Anderson, S. (1999). Space utilization and swimming depth of white
3523 sharks, *Carcharodon carcharias*, at the South Farallon Islands, Central California.
3524 *Environmental Biology of Fishes* **56**, 351-364.
3525

3526 Gill, A.B. (2003). The dynamics of prey choice in fish: the importance of prey size and
3527 satiation. *Journal of Fish Biology* **63**, 105-116. doi:10.1111/j.1095-8649.2003.00214.x
3528

3529 Gillanders, B.M. (2006). Seagrasses, fish, and fisheries. In 'Seagrass: biology, ecology, and
3530 conservation' (Eds A. Larkum, R.J. Orth, and C.M., Duarte) pp. 503-505. (Springer:
3531 Netherlands.)
3532

3533 Graham, B., Koch, P., Newsome, S., McMahon, K., and Aurioles, D. (2010). Using isoscapes
3534 to trace the movements and foraging behavior of top predators in oceanic ecosystems.
3535 In 'Isoscapes' (Eds J.B. West, G.J. Bowen, T.E. Dawson and K.P. Tu.) pp. 299-318.
3536 (Springer: Netherlands.)
3537

3538 Graham, N.J. (2007). Ecological versatility and the decline of coral feeding fishes following
3539 climate driven coral mortality. *Marine Biology* **153**, 119-127.
3540

3541 Graham, N.A.J., Wilson, S.K., Pratchett, M.S., Polunin, N.V.C., and Spalding, M.D. (2009).
3542 Coral mortality versus structural collapse as drivers of corallivorous butterflyfish
3543 decline. *Biodiversity Conservation* **18**, 3325-3336.
3544

- 3545 Great Barrier Reef Marine Park Authority (2011). 'A Vulnerability Assessment for the Great
3546 Barrier Reef: Seagrass.'
- 3547
- 3548 Grinnell, J. (1917). The niche-relationships of the california thrasher. In 'Niche: theory and
3549 application' (Eds R. Whittaker. and A. Simon) pp. 12-18. (Dowden, Hutchinson and
3550 Ross, Inc: Stroudsburg, Pennsylvania.)
- 3551
- 3552 Grubbs, R.D. (2010). Ontogenetic shifts in movements and habitat use. In 'Sharks and their
3553 relatives II: Biodiversity, adaptive physiology, and conservation'. (Eds J.C. Carrier,
3554 J.A. Musick, and M.R. Heithaus) pp. 319-342. (CRC Press: Boca Raton, Florida.)
- 3555
- 3556 Gruber, S.H., Nelson, D.R., and Morrissey, J.F. (1988). Patterns of activity and space
3557 utilization of lemon sharks, *Negaprion brevirostris*, in a shallow Bahamian lagoon.
3558 *Bulletin of Marine Science* **43**, 61-76.
- 3559
- 3560 Gruber, S.H., de Marignac, J.R.C., and Hoenig, J.M. (2001). Survival of juvenile lemon
3561 sharks at Bimini, Bahamas, estimated by mark-depletion experiments. *Transaction of*
3562 *the American Fisheries Society* **130**, 376-384.
- 3563
- 3564 Gutteridge, A.N., Bennett, M.B., Huveneers, C., and Tibbetts, I.R. (2011). Assessing the
3565 overlap between the diet of a coastal shark and the surrounding prey communities in a
3566 sub-tropical embayment. *Journal of Fish Biology* **78**, 1405-1422.
- 3567
- 3568 Hallacher, L.E. (1977). On the feeding behavior of the Basking shark, *Cetorhinus maximus*.
3569 *Environmental Biology of Fishes* **2**, 297-298. doi:10.1007/bf00005996
- 3570
- 3571 Halpern, B.S., Walbridge, S., Selkoe, K.A., Kappel, C.V., Micheli, F., D'Agrosa, C., Bruno,
3572 J.F., Casey, K.S., Ebert, C., Fox, H.E., Fujita, R., Heinemann, D., Lenihan, H.S.,
3573 Madin, E.M.P., Perry, M.T., Selig, E.R., Spalding, M., Steneck, R., and Watson, R.
3574 (2008). A global map of human impact on marine ecosystems. *Science* **319**, 948-952.
- 3575
- 3576 Hammerschlag, N., and Sulikowski, J. (2011). Killing for conservation: the need for
3577 alternatives to lethal sampling of apex predatory sharks. *Endangered Species*
3578 *Research* **14**, 135-140.
- 3579
- 3580
- 3581 Hanchet, S. (1991). Diet of spiny dogfish, *Squalus acanthias* Linnaeus, on the east coast,
3582 South Island, New Zealand. *Journal of Fish Biology* **39**, 313-323. doi:10.1111/j.1095-
3583 8649.1991.tb04365.x
- 3584
- 3585 Harley, C.D.G., Randall Hughes, A., Hultgren, K.M., Miner, B.G., Sorte, C.J.B., Thornber,
3586 C.S., Rodriguez, L.F., Tomanek, L., and Williams, S.L. (2006). The impacts of
3587 climate change in coastal marine systems. *Ecology Letters* **9**, 228-241.
- 3588
- 3589 Harry, A.V., Tobin, A.J., Simpfendorfer, C.A., Welch, D.J., Mapleston, A., White, J.,
3590 Williams, A.J., and Stapley, J. (2011). Evaluating catch and mitigating risk in a
3591 multispecies, tropical, inshore shark fishery within the Great Barrier Reef World
3592 Heritage Area. *Marine and Freshwater Research* **62**, 710-721.
- 3593

- 3594 Haynes, D., Müller, J., and Carter, S. (2000). Pesticide and herbicide residues in sediments
3595 and seagrasses from the Great Barrier Reef World Heritage Area and Queensland
3596 coast. *Marine Pollution Bulletin* **41**, 279-287.
3597
- 3598 Haynes, D., Ralph, P., Prange, J., and Dennison, B. (2000). The impact of the herbicide
3599 diuron on photosynthesis in three species of tropical seagrass. *Marine Pollution*
3600 *Bulletin* **41**, 288-293.
3601
- 3602 Heithaus, M. (2001). The biology of tiger sharks, *Galeocerdo cuvier*, in Shark Bay, Western
3603 Australia: sex ratio, size distribution, diet, and seasonal changes in catch rates.
3604 *Environmental Biology of Fishes* **61**, 25-36. doi:10.1023/a:1011021210685
3605
- 3606 Heithaus, M.R. (2005). Habitat use and group size of pied cormorants (*Phalacrocorax varius*)
3607 in a seagrass ecosystem: possible effects of food abundance and predation risk.
3608 *Marine Biology* **147**:27-35
3609
- 3610 Heithaus, M.R. (2007). Nursery areas as essential shark habitats: a theoretical perspective. In
3611 'American Fisheries Society Symposium 2007'. pp. 3-13. American Fisheries Society.
3612
- 3613 Heithaus, M.R., and Dill, L.M. (2002). Food availability and tiger shark predation risk
3614 influence bottlenose dolphin habitat use. *Ecology* **83**, 480-491.
3615
- 3616 Heithaus, M., Dill, L., Marshall, G., and Buhleier, B. (2002). Habitat use and foraging
3617 behavior of tiger sharks (*Galeocerdo cuvier*) in a seagrass ecosystem. *Marine Biology*
3618 **140**, 237-248.
3619
- 3620 Heithaus, M.R., Frid, A., Wirsing, A.J., and Worm, B. (2008). Predicting ecological
3621 consequences of marine top predator declines. *Trends in Ecology and Evolution* **23**,
3622 202-210.
3623
- 3624 Heithaus, M.R., Hamilton, I.M., Wirsing, A.J., and Dill, L.M. (2006). Validation of a
3625 randomization procedure to assess animal habitat preferences: microhabitat use of
3626 tiger sharks in a seagrass ecosystem. *Journal of Animal Ecology* **75**, 666-676.
3627
- 3628 Heithaus, M., Wirsing, A., Dill, L., and Heithaus, L. (2007). Long-term movements of tiger
3629 sharks satellite-tagged in Shark Bay, Western Australia. *Marine Biology* **151**, 1455-
3630 1461. doi:10.1007/s00227-006-0583-y
3631
- 3632 Herbert, C.E., Shutt, J.L., Hobson, K.A., and Weseloh, D.V.C. (2011). Spatial and temporal
3633 differences in the diet of Great Lakes gerring gulls (*Larus argentatus*): evidence from
3634 stable isotope analysis. *Canadian Journal of Fisheries and Aquatic Science* **56**, 323-
3635 338.
3636
- 3637 Heupel, M.R., and Bennett, M.B. (1998). Observations on the diet and feeding habits of the
3638 epaulette shark, *Hemiscyllium ocellatum* (Bonnaterre), on Heron Island Reef, Great
3639 Barrier Reef, Australia. *Marine and Freshwater Research* **49**, 753-756.
3640 doi:http://dx.doi.org/10.1071/MF97026
3641

- 3642 Heupel, M.R., Knip, D.M., Simpfendorfer, C.A., and Dulvy, N.K. (2014). Sizing up the
3643 ecological role of sharks as predators. *Marine Ecology Progress Series* **495**, 291-298.
3644 doi: 10.3354/meps10597.
3645
- 3646 Heupel, M.R., and Simpfendorfer, C.A.(2014). Importance of environmental and biological
3647 drivers in the presence and space use of a reef-associated shark. *Marine Ecology*
3648 *Progress Series* **496**, 47-57. doi: 10.3354/meps10529
3649
- 3650 Heupel, M., and Simpfendorfer, C. (2008). Movement and distribution of young bull sharks
3651 *Carcharhinus leucas* in a variable estuarine environment. *Aquatic Biology* **1**, 277-289.
3652
- 3653 Heupel, M.R., Carlson, J.K., and Simpfendorfer, C.A. (2007). Shark nursery areas: concepts,
3654 definition, characterization and assumptions. *Marine Ecology Progress Series* **337**,
3655 287-297.
3656
- 3657 Heupel, M.R., Simpfendorfer, C.A., and Fitzpatrick, R. (2010). Large-scale movement and
3658 reef fidelity of grey reef sharks. *PLoS ONE* **5**(3):e9650.
3659 doi:10.1371/journal.pone.0009650
3660
- 3661 Heupel, M., Simpfendorfer, C., Collins, A., and Tyminski, J. (2006). Residency and
3662 movement patterns of bonnethead sharks, *Sphyrna tiburo*, in a large Florida estuary.
3663 *Environmental Biology of Fishes* **76**, 47-67.
3664
- 3665 Heupel, M., Simpfendorfer, C., and Hueter, R. (2004). Estimation of shark home ranges using
3666 passive monitoring techniques. *Environmental Biology of Fishes* **71**, 135-142.
3667
- 3668 Heupel, M.R., Yeiser, B.G., Collins, A.B., Ortega, L., and Simpfendorfer, C.A. (2010). Long-
3669 term presence and movement patterns of juvenile bull sharks, *Carcharhinus leucas*, in
3670 an estuarine river system. *Marine and Freshwater Research* **61**, 1-10.
3671
- 3672 Hobson, K.A. (1999). Tracing origins and migration of wildlife using stable isotopes: a
3673 review. *Oecologia* **120**, 314-326. doi 10.1007/s004420050865
3674
- 3675 Hobson, K.A. (2008). Applying isotopic methods to tracking animal movements. In 'Tracking
3676 animal migration with stable isotopes' (Eds K.A. Hobson and L.I. Wassenaar.) pp.
3677 45-70. (Academic Press: San Diego.)
3678
- 3679 Hobson, K.A., and Clark, R.G. (1992). Assessing avian diets using stable isotopes I: turnover
3680 of ¹³C in tissues. *The Condor* **94**, 181-188. doi 10.2307/1368807
3681
- 3682 Holland, K.N., Wetherbee, B.M., Lowe, C.G., and Meyer, C.G. (1999). Movements of tiger
3683 sharks (*Galeocerdo cuvier*) in coastal Hawaiian waters. *Marine Biology* **134**, 665-673.
3684 doi:10.1007/s002270050582
3685
- 3686 Holland, K.N., Wetherbee, B.M., Peterson, J.D., and Lowe, C.G. (1993). Movements and
3687 distribution of hammerhead shark pups on their natal grounds. *Copeia* **1993**, 495-502.
3688
- 3689 Holts, D., and Bedford, D. (1993). Horizontal and vertical movements of the shortfin mako
3690 shark, *Isurus oxyrinchus*, in the Southern California Bight. *Marine and Freshwater*
3691 *Research* **44**, 901-909. doi:http://dx.doi.org/10.1071/MF9930901

3692
3693 Hughes, A.R., Williams, S.L., Duarte, C.M., Heck, K.L., and Waycott, M. (2008).
3694 Associations of concern: declining seagrasses and threatened dependant species.
3695 *Frontiers in Ecology and the Environment* **7**, 242-246.
3696
3697 Hussey, N.E., Dudley, S.F.J., McCarthy, I.D., Cliff, G., and Fisk, A.T. (2011). Stable isotope
3698 profiles of large marine predators: viable indicators of trophic position, diet, and
3699 movement in sharks? *Canadian Journal of Fisheries and Aquaculture Science* **68**,
3700 2029-2045.
3701
3702 Hussey, N.E., MacNeil, M.A., Olin, J.A., McMeans, B.C., Kinney, M.J., Chapman, D.D., and
3703 Fisk, A.T. (2012a). Stable isotopes and elasmobranchs: tissue types, methods,
3704 applications and assumptions. *Journal of Fish Biology* **80**, 1449-1484.
3705 doi:10.1111/j.1095-8649.2012.03251.x
3706
3707 Hussey, N.E., Olin, J.A., Kinney, M.J., McMeans, B.C., and Fisk, A.T. (2012b). Lipid
3708 extraction effects on stable isotope values ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) of elasmobranch muscle
3709 tissue. *Journal of Experimental Marine Biology and Ecology* **434-435**, 7-15.
3710
3711 Hutchinson, G. (1957) Concluding remarks. In 'Niche: Thoery and Application'. (Eds R.
3712 Whittaker, and S. Levin). pp. 387-399. (Dowden, Hutchinson and Ross, Inc:
3713 Stroudsburg, Pennsylvania.)
3714
3715 Irschick, D., Dyer, L., and Sherry, T.W. (2005). Phylogenetic methodologies for studying
3716 specialization. *Oikos* **110**, 404-408. doi:10.1111/j.0030-1299.2005.13927.x
3717
3718 Jackson, A.L., Inger, R., Parnell, A.C., and Bearhop, S. (2011). Comparing isotopic niche
3719 widths among and within communities: SIBER - Stable Isotope Bayesian Ellipses in
3720 R. *Journal of Animal Ecology* **80**, 595-602.
3721
3722 Jackson, J.B.C., Kirby, M.X., Berger, W.H., Bjorndal, K.A., Botsford, L.W., Bourque, B.J.,
3723 Bradbury, R.H., Cooke, R., Erlandson, J., Estes, J.A., Hughes, T.P., Kidwell, S.,
3724 Lange, C.B., Lenihan, H.S., Pandolfi, J.M., Peterson, C.H., Steneck, R.S., Tegner,
3725 M.J., and Warner, R.R. (2001). Historical overfishing and the recent collapse of
3726 coastal ecosystems. *Science* **293**, 629-637.
3727
3728 Jackson, E.L., Rowden, A.A., Attrill, M.J., Bossley, S.J., and Jones, M.B. (2001). The
3729 importance of seagrass beds as a habitat for fisheries species. *Oceanography and*
3730 *Marine Biology, an Annual Review* **39**, 269-303.
3731
3732 Jarman, S.N., and Wilson, S.G. (2004). DNA-based species identification of krill consumed
3733 by whale sharks. *Journal of Fish Biology* **65**, 586-591.
3734
3735 Johnson, D.H. (1980). The comparison of usage and availability measurements for evaluating
3736 resource preference. *Ecology* **61**, 65-71. doi:10.2307/1937156
3737
3738 Jones, B.C., and Geen, G.H. (1977). Food and feeding of spiny dogfish (*Squalus acanthias*)
3739 in British Columbia waters. *Journal of the Fisheries Research Board of Canada* **34**,
3740 2056-2066. doi:10.1139/f77-274
3741

- 3742 Jorgensen, S.J., Reeb, C.A., Chapple, T.K., Anderson, S., Perle, C., Van Sommeran, S.R.,
3743 Fritz-Cope, C., Brown, A.C., Klimley, A.P., and Block, B.A. (2010). Philopatry and
3744 migration of Pacific white sharks. *Proceedings of the Royal Society B: Biological*
3745 *Sciences* **277**, 679-688. doi:10.1098/rspb.2009.1155
3746
- 3747 Julliard, R., Clavel, J., Devictor, V., Jiguet, F., and Couvet, D. (2006). Spatial segregation of
3748 specialists and generalists in bird communities. *Ecology Letters* **9**, 1237-1244.
3749 doi:10.1111/j.1461-0248.2006.00977.x
3750
- 3751 Kawecki, T.J. (1994). Accumulation of deleterious mutations and the evolutionary cost of
3752 being a generalist. *The American Naturalist* **144**, 833-838.
3753
- 3754 Kennish, M. (1998). 'Pollution impacts on marine biotic communities.' (CRC Press: Boca
3755 Raton, Florida.)
3756
- 3757 Kerr, L.A., Andrews, A.H., Cailliet, G.M., Brown, T.A., & Coale, K.H. (2006). Investigations
3758 of $\delta^{14}\text{C}$, $\delta^{13}\text{C}$, and $\delta^{15}\text{N}$ in vertebrae of white shark (*Carcharodon carcharias*) from
3759 the eastern North Pacific Ocean. In 'Special issue: age and growth of chondrichthyan
3760 fishes: new methods, techniques and analysis' (Eds J.K. Carlson and K.J. Goldman.)
3761 pp. 337-353. (Springer: Netherlands.)
3762
- 3763 Kessel, S.T., Cooke, S.J., Heupel, M.R., Hussey, N.E., Simpfendorfer, C.A., Vagle, S., and
3764 Fisk, A.T. (2013). A review of detection range testing in aquatic passive acoustic
3765 telemetry studies. *Reviews in Fish Biology and Fisheries* **24**, 1-20.
3766 doi:10.1007/s11160-013-9328-4.
3767
- 3768 Kim, S.L., del Rio, C.M., Casper, D., and Koch, P.L. (2012). Isotopic incorporation rates for
3769 shark tissues from a long-term captive feeding study. *Journal of Experimental Biology*
3770 **215**, 2495-2500. doi 10.1242/jeb.070656
3771
- 3772 Kinney, M., Hussey, N., Fisk, A., Tobin, A., and Simpfendorfer, C. (2011). Communal or
3773 competitive? Stable isotope analysis provides evidence of resource partitioning within
3774 a communal shark nursery. *Marine Ecology Progress Series* **439**, 263-276. doi
3775 10.3354/meps09327
3776
- 3777 Klages, J., Broad, A., Kelaher, B.P., and Davis, A.R. (2014). The influence of gummy sharks,
3778 *Mustelus antarcticus*, on observed fish assemblage structure. *Environmental Biology*
3779 *of Fishes* **97**, 215-222.
3780
- 3781 Knip, D.M., Heupel, M.R., and Simpfendorfer, C.A. (2010). Sharks in nearshore
3782 environments: models, importance, and consequences. *Marine Ecology Progress*
3783 *Series* **402**, 1-11.
3784
- 3785 Knip, D.M., Heupel, M.R., and Simpfendorfer, C.A. (2012a). Evaluating marine protected
3786 areas for the conservation of tropical coastal sharks. *Biological Conservation* **148**,
3787 200-209.
3788
- 3789 Knip, D.M., Heupel, M.R., and Simpfendorfer, C.A. (2012b). Habitat use and spatial
3790 segregation of adult spottail sharks *Carcharhinus sorrah* in tropical nearshore waters.
3791 *Journal of Fish Biology* **80**, 767-784.

3792
3793 Knip, D.M., Heupel, M.R., and Simpfendorfer, C.A. (2012c) To roam or to home: site fidelity
3794 in a tropical coastal shark. *Marine Biology* **159**, 1647-1657
3795
3796 Knip, D.M., Heupel, M.R., Simpfendorfer, C.A., Tobin, A.J., and Moloney, J. (2011a).
3797 Ontogenetic shifts in movement and habitat use of juvenile pigeye sharks
3798 *Carcharhinus amboinensis* in a tropical nearshore region. *Marine Ecology Progress*
3799 *Series* **425**, 233-246.
3800
3801 Knip, D.M., Heupel, M.R., Simpfendorfer, C.A., Tobin, A.J., and Moloney, J. (2011b). Wet-
3802 season effects on the distribution of juvenile pigeye sharks, *Carcharhinus*
3803 *amboinensis*, in tropical nearshore waters. *Marine and Freshwater Research* **62**, 658-
3804 667.
3805
3806 Kohler, N., Casey, J., and Turner, P. (1998). NMFS cooperative shark tagging program,
3807 1962-93: An atlas of shark tag and recapture data. *Marine Fisheries Review* **60**, 1-87.
3808
3809 Kohler, N.E., and Turner, P.A. (2001). Shark tagging: A review of conventional methods and
3810 studies. *Environmental Biology of Fishes* **60**, 191-224.
3811
3812 Kuba, M., Byrne, R., and Burghardt, G. (2010). A new method for studying problem solving
3813 and tool use in stingrays (*Potamotrygon castexi*). *Animal Cognition* **13**, 507-513.
3814
3815 Last, P.R., and Stevens, J.D. (2009). ‘Sharks and rays of Australia.’ 2nd Edn.(CSIRO
3816 Publishing Collingwood: Victoria, Australia.)
3817
3818 Layman. C.A., Arrington, D.A., Montaña, C.G., and Post, D.M. (2007). Can stable isotope
3819 ratios provide for community-wide measures of trophic structure? *Ecology* **88**, 42-48.
3820
3821 Layman, C.A., Araujo, M.S., Boucek, R., Hammerschlag-Peyer, C.M., Harrison, E., Jud,
3822 Z.R., Matich, P., Rosenblatt, A.E., Vaudo, J.J., Yeager, L.A., Post, D.M., and
3823 Bearhop, S. (2012). Applying stable isotopes to examine food-web structure: an
3824 overview of analytical tools. *Biological Reviews* **87**, 545-562.
3825
3826 Leibold, M.A. (1995). The niche concept revisited: mechanistic models and community
3827 context. *Ecology* **76**, 1371-1382. doi:10.2307/1938141
3828
3829 Levins. R. (1968). 'Evolution in changing environments.' (Princeton University Press:
3830 Princeton, New Jersey, USA.)
3831
3832 Lima, S.L., and Dill, L.M. (1990). Behavioral decisions made under the risk of predation: a
3833 review and prospectus. *Canadian Journal of Zoology* **68**, 619-640.
3834
3835 Link, J.S., and Ford, M.D. (2006). Widespread and persistent increase of Ctenophora in the
3836 continental shelf ecosystem off NE USA. *Marine Ecology Progress Series* **320**, 153-
3837 159. doi:10.3354/meps320153
3838
3839 Loefer, J.K., and Sedberry, G.R. (2003). Life history of the Atlantic sharpnose shark
3840 (*Rhizoprionodon terraenovae*) (Richardson, 1836) off the southern United States.
3841 *Fisheries Bulletin* **101**, 75-88.

3842
3843 Logan, J., and Lutcavage, M. (2010). Stable isotope dynamics in elasmobranch fishes.
3844 *Hydrobiologia* **644**, 231-244. doi 10.1007/s10750-010-0120-3
3845

3846 Lotze, H.K., Lenihan, H.S., Bourque, B.J., Bradbury, R.H., Cooke, R.G., Kay, M.C., Kidwell,
3847 S.M., Kirby, M.X., Peterson, C.H., and Jackson, J.B.C. (2006). Depletion,
3848 degradation, and recovery potential of estuaries and coastal seas. *Science* **312**, 1806-
3849 1809.
3850

3851 Lundberg, J., and Moberg, F. (2003). Mobile link organisms and ecosystem functioning:
3852 implications for ecosystem resilience and management. *Ecosystems* **6**, 0087-0098. doi
3853 10.1007/s10021-002-0150-4.
3854

3855 Lowe, C., Wetherbee, B., Crow, G., and Tester, A. (1996). Ontogenetic dietary shifts and
3856 feeding behavior of the tiger shark, *Galeocerdo cuvier*, in Hawaiian waters.
3857 *Environmental Biology of Fishes* **47**, 203-211. doi:10.1007/bf00005044
3858

3859 Lucifora, L.O., Garcia, V.B., Menni, R.C., and Escalante, A.H. (2006). Food habits,
3860 selectivity, and foraging modes of the school shark *Galeorhinus galeus*. *Marine*
3861 *Ecology Progress Series* **315**, 259-270. doi:10.3354/meps315259
3862

3863 Lundberg, J., and Moberg, F. (2003). Mobile link organisms and ecosystem functioning:
3864 implications for ecosystem resilience and management. *Ecosystems* **6**, 0087-0098. doi
3865 10.1007/s10021-002-0150-4
3866

3867 Lyle, J. (1987). Observations on the biology of *Carcharhinus cautus* (Whitley), *C.*
3868 *melanopterus* (Quoy & Gaimard) and *C. fitzroyensis* (Whitley) from Northern
3869 Australia. *Marine and Freshwater Research* **38**, 701-710.
3870

3871 MacNeil, M.A., Drouillard, K.G., and Fisk, A.T. (2006). Variable uptake and elimination of
3872 stable nitrogen isotopes between tissues in fish. *Canadian Journal of Fisheries and*
3873 *Aquatic Sciences* **63**, 345-353.
3874

3875 MacNeill, M.A., Skomal, G.B., and Fisk, A.T. (2005). Stable isotopes from multiple tissues
3876 reveal diet switching in sharks. *Marine Ecology Progress Series* **302**, 199-206. doi
3877 10.3354/meps302199
3878

3879 Malpica-Cruz, L., Herzka Sharon, Z., Sosa-Nishizaki, O., and Lazo, J.P. (2012). Tissue-
3880 specific isotope trophic discrimination factors and turnover rates in a marine
3881 elasmobranch: empirical and modeling results. *Canadian Journal of Fisheries and*
3882 *Aquatic Science* **69**, 551-564. doi 10.1139/f2011-172
3883

3884 Manly, B.F.J., McDonald, L.L., Thomas, D.L., McDonald, T.L., and Erickson, W.P. (2002).
3885 'Resource selection by animals: statistical design and analysis for field studies.'
3886 (Kluwer Academic Publishers: Netherlands, Dordrecht.)
3887

3888 Marshall, P., Beeden, R., Dryden, J., and Goldberg, J. (2011). Impacts of tropical cyclone
3889 Yasi on the Great Barrier Reef: a report on the findings of a rapid ecological impact

3890 assessment, July 2011, GBRMPA, Townsville. Great Barrier Reef Marine Park
3891 Authority.

3892

3893 Martin, R.A. (2007). A review of the behavioural ecology of whale sharks (*Rhincodon typus*).
3894 *Fisheries Research* **84**, 10-16.

3895

3896 Marvier, M., Kareiva, P., and Neubert, M.G. (2004). Habitat destruction, fragmentation, and
3897 disturbance promote invasion by habitat generalists in a multispecies metapopulation.
3898 *Risk Analysis* **24**, 869-878.

3899

3900 Matich, P., Heithaus, M.R., and Layman, C.A. (2011). Contrasting patterns of individual
3901 specialisation and trophic coupling in two marine apex predators. *Journal of Animal*
3902 *Ecology* **80**, 294-305.

3903

3904 McCarthy, M.A. (2007). 'Bayesian Methods for Ecology.' (New York:Cambridge University
3905 Press.)

3906

3907 McCauley, D.J., DeSalles, P.A., Young, H.S., Papastamatiou, Y.P., Caselle, J.E., Deakos
3908 M.H., Gardner, J.P.A., Garton, D.W., Collen, J.D., and Micheli, F. (2014). Reliance
3909 of mobile species on sensitive habitats: a case study of manta rays (*Manta alfredi*)
3910 and lagoons. *Marine Biology* **9**, 1987-1998.

3911

3912 McElroy, W.D., Wetherbee, B., Mostello, C., Lowe, C., Crow, G., and Wass, R. (2006). Food
3913 habits and ontogenetic changes in the diet of the sandbar shark, *Carcharhinus*
3914 *plumbeus*, in Hawaii. *Environmental Biology of Fishes* **76**, 81-92.

3915

3916 McGlathery, K.J. (2001). Macroalgal blooms contribute to the decline of seagrass in nutrient-
3917 enriched coastal waters. *Journal of Phycology* **37**, 453-456.

3918

3919 McKenzie, L., Collier, C., Waycott, M., Unsworth, R., Yoshida, R., and Smith, N. (2012).
3920 Monitoring inshore seagrasses of the GBR and responses to water quality In
3921 'Proceedings of the 12th International Coral Reef Symposium' (Eds D. Yellowlees
3922 and T. P. Hughes.) pp. ICRS2012_15B_4. (ARC Center for Excellence for Coral Reef
3923 Studies: Cairns, Australia.)

3924

3925 McKibben, J.N., and Nelson, D.R. (1986). Patterns of movement and grouping of gray reef
3926 sharks, *Carcharhinus amblyrhynchos*, at Enewetak, Marshall Islands. *Bulletin of*
3927 *Marine Science* **38**, 89-110.

3928

3929 McKinney, M.L. (1997). Extinction vulnerability and selectivity: combining ecological and
3930 paleontological views. *Annual Review Ecology and Systematics*. **28**, 495-516.

3931

3932 McMeans, B.C., Svavarsson, J., Dennard, S., and Fisk, A.T. (2010). Diet and resource use
3933 among Greenland sharks (*Somniosus microcephalus*) and teleosts sampled in
3934 Icelandic waters, using $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, and mercury. *Canadian Journal of Fisheries and*
3935 *Aquatic Science* **67**, 1428-1438. doi 10.1139/f10-072

3936

3937 Meekan, M.G., Jarman, S.N., McLean, C., and Schultz, M.B. (2009). DNA evidence of whale
3938 sharks (*Rhincodon typus*) feeding on red crab (*Gecarcoidea natalis*) larvae at
3939 Christmas Island, Australia. *Marine and Freshwater Research* **60**, 607-609.

3940
3941 Meyer, C., Papastamatiou, Y., and Holland, K. (2010). A multiple instrument approach to
3942 quantifying the movement patterns and habitat use of tiger (*Galeocerdo cuvier*) and
3943 Galapagos sharks (*Carcharhinus galapagensis*) at French Frigate Shoals, Hawaii.
3944 *Marine Biology* **157**, 1857-1868. doi:10.1007/s00227-010-1457-x
3945
3946 Meyer, C.G., Clark, T.B., Papastamatiou, Y.P., Whitney, N.M., and Holland, K.N. (2009).
3947 Long-term movement patterns of tiger sharks *Galeocerdo cuvier* in Hawaii. *Marine*
3948 *Ecology Progress Series* **381**, 223-235. doi:10.3354/meps07951
3949
3950 Meynecke, J.O., and Lee, S.Y. (2011), Climate-coastal fisheries relationships and their spatial
3951 variation in Queensland, Australia. *Fisheries Research* **110**, 365-376.
3952
3953 Mittelbach, G., Osenberg, C., and Wainwright, P. (1992). Variation in resource abundance
3954 affects diet and feeding morphology in the pumpkinseed sunfish (*Lepomis gibbosus*).
3955 *Oecologia* **90**, 8-13.
3956
3957 Montoya, J.P. (2007) Natural abundance of ¹⁵N in marine planktonic ecosystems. In ‘Stable
3958 Isotopes in Ecology and Environmental Science, Second Edition’. (Eds R. Michener
3959 and K Lajtha.) pp 176-200. (Blackwell Publishing: Oxford UK.)
3960
3961 Morrissey, J.F., and Gruber, S.H. (1993). Habitat selection by juvenile lemon sharks,
3962 *Negaprion brevirostris*. *Environmental Biology of Fishes* **38**, 311-319.
3963
3964 Motta, P.J., Maslanka, M., Hueter, R.E., Davis, R.L., de la Parra, R., Mulvany, S.L.,
3965 Habegger, M.L., Strother, J.A., Mara, K.R., Gardiner, J.M., Tyminski, J.P., and
3966 Zeigler, L.D. (2010). Feeding anatomy, filter-feeding rate, and diet of whale sharks
3967 *Rhincodon typus* during surface ram filter feeding off the Yucatan Peninsula, Mexico.
3968 *Zoology* **113**, 199-212. doi:10.1016/j.zool.2009.12.001
3969
3970 Munday, P.L. (2004), Habitat loss, resource specialization, and extinction on coral reefs.
3971 *Global Change Biology* **10**, 1642-1647. doi:10.1111/j.1365-2486.2004.00839.x
3972
3973 Murchie, K., Schwager, E., Cooke, S., Danylchuk, A., Danylchuk, S., Goldberg, T., Suski, C.,
3974 and Philipp, D. (2010). Spatial ecology of juvenile lemon sharks (*Negaprion*
3975 *brevirostris*) in tidal creeks and coastal waters of Eleuthera, The Bahamas.
3976 *Environmental Biology of Fishes* **89**, 95-104.
3977
3978 Newman, S.P., Handy, R.D., and Gruber, S.H. (2010). Diet and prey preference of juvenile
3979 lemon sharks *Negaprion brevirostris*. *Marine Ecology Progress Series* **398**, 221-234.
3980 doi:10.3354/meps08334
3981
3982 Newsome, S.D., Tinker, M.T., Monson, D.H., Oftedal, O.T., Ralls, K., Staedler, M.M., Fogel,
3983 M.L., and Estes, J.A. (2009). Using stable isotopes to investigate individual diet
3984 specialization in California sea otters (*Enhydra lutris nereis*). *Ecology* **90**, 961-974.
3985 doi 10.1890/07-1812.1
3986
3987 Nixon, S.W., Oviatt, C.A., Frithsen, J., and Sullivan, B. (1986). Nutrients and the productivity
3988 of estuarine and coastal marine ecosystems. *Journal of the Limnological Society of*
3989 *Southern Africa* **12**, 43-71.

3990
3991 Nøttestad, L., Giske, J., Holst, J.C., and Huse, G. (1999). A length-based hypothesis for
3992 feeding migrations in pelagic fish. *Canadian Journal of Fisheries and Aquatic Science*
3993 **56**, 26-34. doi 10.1139/f99-222
3994
3995 Olin, J.A., Hussey, N.E., Fritts, M., Heupel, M.R., Simpfendorfer, C.A., Poulakis, G.R., and
3996 Fisk, A.T. (2011). Maternal meddling in neonatal sharks: implications for interpreting
3997 stable isotopes in young animals. *Rapid Communications in Mass Spectrometry* **25**,
3998 1008-1016. doi 10.1002/rcm.4946
3999
4000 Oro, D., Genovart, X., Ruiz, X., Jimenez, J., and Garcia-Gans, J. (1996). Differences in diet,
4001 population size, and reproductive performance between two colonies of Audouin's
4002 Gull *Larus audouinii* affed by a trawling moratorium. *Journal of Avian Biology* **27**,
4003 245-251.
4004
4005 Ortega, L.A., Heupel, M.R., Van Beynen, P., and Motta, P.J. (2009). Movement patterns and
4006 water quality preferences of juvenile bull sharks (*Carcharhinus leucas*) in a Florida
4007 estuary. *Environmental Biology of Fisheries* **84**, 361-373. doi:10.1007/s10641-009-
4008 9442-2
4009
4010 Orth, R., Heck, K., and van Montfrans, J. (1984). Faunal communities in seagrass beds: A
4011 review of the influence of plant structure and prey characteristics on predator-prey
4012 relationships. *Estuaries* **7**, 339-350.
4013
4014 Orth, R.J., Carruthers, T.J.B., Dennison W.C., Duarte, C.M., Fourqurean J.W., Heck, K.L.Jr.,
4015 Hughes, A.R., Kendrick G.A., Kenworthy, W.J., Olyarnik S, Short, F.T., Waycott, M.,
4016 and Williams, S.L. (2006). A global crisis for seagrass ecosystems. *BioScience* **56**,
4017 987-996.
4018
4019 Ovenden, J., Morgan, J., Street, R., Tobin, A., Simpfendorfer, C., Macbeth, W., and Welch,
4020 D. (2011). Negligible evidence for regional genetic population structure for two shark
4021 species *Rhizoprionodon acutus* (Rüppell, 1837) and *Sphyrna lewini* (Griffith & Smith,
4022 1834) with contrasting biology. *Marine Biology* **158**: 1497-1509. doi 10.1007/s00227-
4023 011-1666-y
4024
4025 Papastamatiou, Y.P., Friedlander, A.M., Caselle, J.E., and Lowe, C.G. (2010). Long-term
4026 movement patterns and trophic ecology of blacktip reef sharks (*Carcharhinus*
4027 *melanopterus*) at Palmyra Atoll. *Journal of Experimental Marine Biology and*
4028 *Ecology* **386**, 94-102. doi 10.1016/j.jembe.2010.02.009
4029
4030 Papastamatiou, Y., Wetherbee, B., O'Sullivan, J., Goodmanlowe, G., and Lowe, C. (2010).
4031 Foraging ecology of cookiecutter sharks (*Isistius brasiliensis*) on pelagic fishes in
4032 Hawaii, inferred from prey bite wounds. *Environmental Biology of Fishes* **88**, 361-
4033 368. doi:10.1007/s10641-010-9649-2
4034
4035 Papastamatiou, Y.P., Wetherbee, B.M., Lowe, C.G., and Crow, G.L. (2006). Distribution and
4036 diet of four species of carcharhinid shark in the Hawaiian Islands: evidence for
4037 resource partitioning and competitive exclusion. *Marine Ecology Progress Series* **320**,
4038 239-251. doi:10.3354/meps320239
4039

- 4040 Pardini, A.T., Jones, C.S., Noble, L.R., Kreiser, B., Malcolm, H., Bruce, B.D., Stevens, J.D.,
 4041 Cliff, G., Scholl, M.C., and Martin, A.P. (2001). Sex-biased dispersal of great white
 4042 sharks. *Nature* **412**,139-140.
 4043
- 4044 Parnell, A., and Jackson, A. (2011). SIAR: stable isotope analysis in R. R package version
 4045 4.1.3.
 4046
- 4047 Parnell, A.C., Inger, R., Bearhop, S., and Jackson, A.L. (2010). Source partitioning using
 4048 stable isotopes: coping with too much variation. *PLoS ONE* **5**:e9672.
 4049
- 4050 Parsons, G. (1993). Geographic variation in reproduction between two populations of the
 4051 bonnethead shark, *Sphyrna tiburo*. *Environmental Biology of Fishes* **38**, 25-35.
 4052
- 4053 Peers, M.J.L., Thornton, D.H., and Murray, D.L. (2012). Reconsidering the specialist-
 4054 generalist paradigm in niche breadth dynamics: resource gradient selection by Canada
 4055 lynx and bobcat. *PLoS ONE* **7**:e51488. doi:10.1371/journal.pone.0051488
 4056
- 4057 Peterson, A.T., Soberón, J., Pearson, R.G., Anderson, R.P., Martínez-Meyer, E., Nakamura,
 4058 M., and Araújo, M.B. (2011). 'Ecological niches and geographic distributions'
 4059 (Princeton University Press: Princeton.)
 4060
- 4061 Peterson, B.J., and Fry, B. (1987). Stable isotopes in ecosystem studies. *Annual Review*
 4062 *Ecology and Systematics* **18**, 293-320.
 4063
- 4064 Pikitch, E.K., Chapman, D.D., Babcock, E.A., and Shivji, M.S. (2005). Habitat use and
 4065 demographic population structure of elasmobranchs at a Caribbean atoll (Glover's
 4066 Reef, Belize). *Marine Ecology Progress Series* **302**, 187-197.
 4067 doi:10.3354/meps302187
 4068
- 4069 Pillans, R.D., Stevens, J.D., Kyne, P.M., and Salinil, J. (2010). Observations on the
 4070 distribution, biology, short-term movements and habitat requirements of river sharks
 4071 *Glyphis spp.* in northern Australia. *Endangered Species Research* **10**, 321-332.
 4072
- 4073 Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D. (2013). nlme: Linear and nonlinear mixed
 4074 effects models. R package version 3.1-109.
 4075
- 4076 Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using
 4077 gibbs sampling. In 'Proceedings of the 3rd International Workshop on Distributed
 4078 Statistical Computing, Vienna, Austria March 20–22 ' (Eds K. Hornik, F. Leisch, and
 4079 A. Zeileis.) March 20–22, Vienna, Austria.
 4080
- 4081 Poisot, T., Canard, E., Mouquet, N., and Hochberg, M.E. (2012). A comparative study of
 4082 ecological specialization estimators. *Methods in Ecology and Evolution* **3**, 537-544.
 4083 doi:10.1111/j.2041-210X.2011.00174.x
 4084
- 4085 Post, D., Layman, C.A., Arrington, D., Takimoto, G., Quattrochi, J., and Montana, C. (2007).
 4086 Getting to the fat of the matter: models, methods and assumptions for dealing with
 4087 lipids in stable isotope analyses *Oecologia* **152**: 179-189.
 4088

- 4089 Post, D.M. (2002). Using stable isotopes to estimate trophic position: models, methods, and
4090 assumptions. *Ecology* **83**, 703-718. doi 10.1890/0012-
4091 9658(2002)083[0703:usitet]2.0.co;2
4092
- 4093 de Queiroz, A., Henke, C., Smith, H.M., and Guyer, C. (2001). Geographic variation and
4094 ontogenetic change in the diet of the Mexican Pacific lowlands garter snake,
4095 *Thamnophis validus*. *Copeia* **2001**, 1034-1042.
4096
- 4097 Ralph, P., Tomasko, D., Moore, K., Seddon, S., and Macinnis-Ng, C.O. (2006). Human
4098 impacts on seagrasses: eutrophication, sedimentation, and contamination. In ‘
4099 Seagrasses: biology, ecology, and conservation.’ pp. 567-593. (Springer:
4100 Netherlands.)
4101
- 4102 Randall, J.E. (1977). Contribution to biology of whitetip reef shark (*Triaenodon obesus*).
4103 *Pacific Science* **31**, 143-164.
4104
- 4105 Randall, J.E. (1992). Review of the biology of the tiger shark (*Galeocerdo cuvier*). *Australian*
4106 *Journal of Marine and Freshwater Research* **43**, 21-31.
4107
- 4108 Rechisky, E., and Wetherbee, B. (2003). Short-term movements of juvenile and neonate
4109 sandbar sharks, *Carcharhinus plumbeus*, on their nursery grounds in Delaware Bay.
4110 *Environmental Biology of Fishes* **68**, 113-128.
4111
- 4112 Reeve, A., Handy, R.D., and Gruber, S.H. (2009). Prey selection and functional response of
4113 juvenile lemon sharks *Negaprion brevirostris*. *Journal of Fish Biology* **75**, 276-281.
4114 doi:10.1111/j.1095-8649.2009.02265.x
4115
- 4116 Reum, J.C.P., and Essington, T.E. (2013). Spatial and seasonal variation in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$
4117 values in a mesopredator shark, *Squalus suckleyi*, revealed through multitissue
4118 analysis. *Marine Biology* **160**, 399-411.
4119
- 4120 Richmond, C.E., Breitburg, D.L., and Rose, K.A. (2005). The role of environmental
4121 generalist species in ecosystem function. *Ecological Modelling* **188**, 279-295.
4122
- 4123 Robertson, A.I., and Duke, N.C. (1987). Mangroves as nursery sites: comparisons of the
4124 abundance and species composition of fish and crustaceans in mangroves and other
4125 nearshore habitats in tropical Australia. *Marine Biology* **96**, 193-205.
4126
- 4127 Roff, D.A. (1988). The evolution of migration and some life history parameters in marine
4128 fishes. *Environmental Biology of Fishes* **22**, 133-146. doi 10.1007/bf00001543
4129
- 4130 Rubenstein, D.R., and Hobson, K.A. (2004). From birds to butterflies: animal movement
4131 patterns and stable isotopes. *Trends in Ecology and Evolution* **19**, 256-263. doi
4132 10.1016/j.tree.2004.03.017
4133
- 4134 Salini, J., Blaber, S., and Brewer, D. (1992). Diets of sharks from estuaries and adjacent
4135 waters of the North-eastern Gulf of Carpentaria, Australia. *Marine and Freshwater*
4136 *Research* **43**, 87-96.
4137

4138 Schluessel, V. (2014). Who would have thought that ‘Jaws’ also has brains? Cognitive
4139 functions in elasmobranchs. *Animal Cognition*, 1-19. DOI 10.1007/s10071-014-0762-
4140 z

4141

4142 Seagrass Watch (2013) ‘Seagrass Watch:Townsville’ Available at
4143 <http://www.seagrasswatch.org/townsville.html> [accessed 30 July 2013].
4144

4145 Secor, D.H., Kerr, L.A., and Cadrin, S.X. (2009). Connectivity effects on productivity,
4146 stability, and persistence in a herring metapopulation model. *ICES Journal of Marine*
4147 *Science* **66**, 1726-1732. doi 10.1093/icesjms/fsp154
4148

4149 Schlaff, A., Heupel, M., and Simpfendorfer, C. (2014). Influence of environmental factors on
4150 shark and ray movement, behaviour and habitat use: a review. *Reviews in Fish*
4151 *Biology and Fisheries*. DOI 10.1007/s11160-014-9364-8
4152

4153 Schroeder, R. (2011). Utilisation of vertebral microchemistry techniques to determine
4154 population structure of two inshore shark species along the east coast of Queensland,
4155 Australia. PhD dissertation, James Cook University.
4156

4157 Shahidul Islam, M., and Tanaka, M. (2004). Impacts of pollution on coastal and marine
4158 ecosystems including coastal and marine fisheries and approach for management: a
4159 review and synthesis. *Marine Pollution Bulletin* **48**, 624-649.
4160

4161 Shea, K., and Chesson, P. (2002). Community ecology theory as a framework for biological
4162 invasions. *Trends in Ecology and Evolution* **17**, 170-176.
4163 doi:[http://dx.doi.org/10.1016/S0169-5347\(02\)02495-3](http://dx.doi.org/10.1016/S0169-5347(02)02495-3)
4164

4165 Shiffman, D.S., Gallagher, A.J., Boyle, M.D., Hammerschlag-Peyer, C.M., and
4166 Hammerschlag, N. (2012). Stable isotope analysis as a tool for elasmobranch
4167 conservation research: a primer for non-specialists. *Marine Freshwater Research* **63**,
4168 635-643.
4169

4170 Simpfendorfer, C. (1992a). Biology of tiger sharks (*Galeocerdo cuvier*) caught by the
4171 Queensland shark meshing program off Townsville, Australia. *Marine and*
4172 *Freshwater Research* **43**, 33-43. doi:<http://dx.doi.org/10.1071/MF9920033>
4173

4174 Simpfendorfer, C. (1992b). Reproductive strategy of the Australian Sharpnose Shark,
4175 *Rhizoprionodon taylori* (Elasmobranchii: Carcharhinidae), from Cleveland Bay,
4176 Northern Queensland. *Marine and Freshwater Research* **43**, 67-75.
4177

4178 Simpfendorfer, C.A. (1993). Age and growth of the Australian sharpnose shark,
4179 *Rhizoprionodon taylori*, from north Queensland, Australia. *Environmental Biology of*
4180 *Fishes* **36**, 233-241.
4181

4182 Simpfendorfer, C.A. (1998). Diet of the Australian sharpnose shark, *Rhizoprionodon taylori*,
4183 from northern Queensland. *Marine and Freshwater Research* **49**, 757-761.
4184

- 4185 Simpfendorfer, C., Chidlow, J., McAuley, R., and Unsworth, P. (2000). Age and Growth of
4186 the Whiskery Shark, *Furgaleus macki*, from Southwestern Australia. *Environmental*
4187 *Biology of Fishes* **58**, 335-343.
- 4188
- 4189 Simpfendorfer, C., Goodreid, A., and McAuley, R. (2001a). Size, sex and geographic
4190 variation in the diet of the tiger shark, *Galeocerdo cuvier*, from Western Australian
4191 waters. *Environmental Biology of Fishes* **61**, 37-46. doi:10.1023/a:1011021710183
4192
- 4193 Simpfendorfer, C.A., Goodreid, A., and McAuley, R.B. (2001b). Diet of three commercially
4194 important shark species from Western Australian waters. *Marine and Freshwater*
4195 *Research* **52**, 975-985. doi:http://dx.doi.org/10.1071/MF01017
4196
- 4197 Simpfendorfer, C.A., Heupel, M.R., and Hueter, R.E. (2002). Estimation of short-term centers
4198 of activity from an array of omnidirectional hydrophones and its use in studying
4199 animal movements. *Canadian Journal of Fisheries and Aquatic Sciences* **59**, 23-32.
4200
- 4201 Simpfendorfer, C.A., Heupel, M.R., White, W.T., and Dulvy, N.K. (2011). The importance of
4202 research and public opinion to conservation management of sharks and rays: a
4203 synthesis. *Marine and Freshwater Research* **62**, 518-527.
4204 doi:http://dx.doi.org/10.1071/MF11086
4205
- 4206 Simpfendorfer, C.A., and Milward, N.E. (1993). Utilization of a tropical bay as a nursery area
4207 by sharks of the families carcharhinidae and sphyrnidae. *Environmental Biology of*
4208 *Fishes* **37**, 337-345.
4209
- 4210 Sims, D., Nash, J., and Morritt, D. (2001). Movements and activity of male and female
4211 dogfish in a tidal sea lough: alternative behavioural strategies and apparent sexual
4212 segregation. *Marine Biology* **139**, 1165-1175.
4213
- 4214 Sims, D.W., Witt, M.J., Richardson, A.J., Southall, E.J., and Metcalfe, J.D. (2006). Encounter
4215 success of free-ranging marine predator movements across a dynamic prey landscape.
4216 *Proceedings of the Royal Society B: Biological Sciences* **273**, 1195-1201.
4217
- 4218 Sims, D.W., and Quayle, V.A. (1998). Selective foraging behaviour of basking sharks on
4219 zooplankton in a small-scale front. *Nature* **393**, 460-464.
4220
- 4221 Smart, J.J., Harry, A.V., Tobin, A.J., and Simpfendorfer, C.A. (2013). Overcoming the
4222 constraints of low sample sizes to produce age and growth data for rare or threatened
4223 sharks. *Aquatic Conservation: Marine and Freshwater Ecosystems* **23**, 124-134.
4224
- 4225 Smith, E.P. (1982). Niche breadth, resource availability, and inference. *Ecology* **63**, 1675-
4226 1681.
4227
- 4228 Springer, S. (1967). Social organization of shark populations. In 'Sharks, skates and rays.'
4229 (Eds P.W. Gilbert, R.F. Matheson and D.P. Rall.) pp. 149-174. (John Hopkins Press:
4230 Baltimore, USA.)
4231
- 4232 Sommerville, E., Platell, M.E., White, W.T., Jones, A.A., and Potter, I.C. (2011). Partitioning
4233 of food resources by four abundant, co-occurring elasmobranch species: relationships

4234 between diet and both body size and season. *Marine and Freshwater Research* **62**, 54-
4235 65.

4236

4237 Spalding, M.D., Fox, H.E., Allen, G.R., Davidson, N., Ferdaña, Z.A., Finlayson, M., Halpern,
4238 B.S., Jorge, M.A., Lombana, A., Lourie, S.A., Martin, K.D., McManus, E., Molnar, J.,
4239 Recchia, C.A., and Robertson, J. (2007). Marine ecoregions of the world: A
4240 bioregionalization of coastal and shelf areas. *BioScience* **57**, 573-583.

4241

4242 Stephens, D.W., and Krebs, J.R. (1986). 'Foraging theory.' (Princeton University Press:
4243 Princeton, New Jersey.)

4244

4245 Stevens, J.D. (2007). Whale shark (*Rhincodon typus*) biology and ecology: A review of the
4246 primary literature. *Fisheries Research* **84**, 4-9.
4247 doi:<http://dx.doi.org/10.1016/j.fishres.2006.11.008>

4248

4249 Stevens, J.D., Bonfil, R., Dulvy, N.K., and Walker, P.A. (2000). The effects of fishing on
4250 sharks, rays, and chimaeras (chondrichthyans), and the implications for marine
4251 ecosystems. *ICES Journal of Marine Science: Journal du Conseil* **57**, 476-494.
4252 doi:10.1006/jmsc.2000.0724

4253

4254 Stevens, J., and McLoughlin, K. (1991). Distribution, size and sex composition, reproductive
4255 biology and diet of sharks from Northern Australia. *Marine and Freshwater Research*
4256 **42**, 151-199.

4257

4258 Stevens, J.D., West, G.J., and McLoughlin, K.J. (2000). Movements, recapture patterns, and
4259 factors affecting the return rate of carcharhinid and other sharks tagged off northern
4260 Australia. *Marine and Freshwater Research* **51**, 127-141. doi
4261 <http://dx.doi.org/10.1071/MF98158>

4262

4263 Strauss, R.E. (1979). Reliability estimates for Ivelv electivity index, the forge ratio, and a
4264 proposed linear index of food selection *Transactions of the American Fisheries*
4265 *Society* **108**, 344-352.

4266

4267 Su, Y., and Yajima, M. (2013). R2jags: a package for running jags from R. R package version
4268 0.03-11.

4269

4270 Suarez, A.V., and Case, T.J. (2002). Bottom-up effects on persistence of a specialist predator:
4271 Ant invasions and horned lizards. *Ecological Applications* **12**, 291-298.

4272

4273 Suárez-Moo, P.D.J., Rocha-Olivares, A., Zapata-Pérez, O., Quiroz-Moreno, A., and Sánchez-
4274 Teyer, L.F. (2013). High genetic connectivity in the Atlantic sharpnose shark,
4275 *Rhizoprionodon terraenovae*, from the southeast Gulf of Mexico inferred from AFLP
4276 fingerprinting. *Fisheries Research* **147**, 338-343. doi
4277 <http://dx.doi.org/10.1016/j.fishres.2013.07.003>

4278

4279 Suchanek, T. (1994). Temperate coastal marine communities: biodiversity and threats.
4280 *American Zoologist* **34**, 100-114.

4281

4282 Sundström, L.F., Gruber, S.H., Clermont, S.M., Correia, J.P.S., de Marniac, J.R.C.,
4283 Morrissey, J.F., Lowrance, C.R., Thomassen, L., and Oliveira, M.T. (2001). Review

4284 of elasmobranch behavioral studies using ultrasonic telemetry with special reference
4285 to the lemon shark, *Negaprion brevirostris*, around Bimini Islands, Bahamas.
4286 *Environmental Biology of Fishes* **60**, 225-250.
4287

4288 Tanasichuk, R.W., Ware, D.M., Shaw, W., and McFarlane, G.A. (1991). Variations in diet,
4289 daily ration, and feeding periodicity of Pacific hake (*Merluccius productus*) and spiny
4290 dogfish (*Squalus acanthias*) off the lower West Coast of Vancouver Island. *Canadian*
4291 *Journal of Fisheries and Aquatic Sciences* **48**, 2118-2128. doi:10.1139/f91-251
4292

4293 Tarboush, R.A., MacAvoy, S.E., Macko, S.A., and Connaughton, V. (2006). Contribution of
4294 catabolic tissue replacement to the turnover of stable isotopes in *Danio rerio*.
4295 *Canadian Journal Zoology* **84**, 1453-1460. doi 10.1139/z06-136
4296

4297 Taylor, S.M., and Bennett, M.B. (2008). Cephalopod dietary specialization and ontogenetic
4298 partitioning of the Australian weasel shark *Hemigaleus australiensis* White, Last &
4299 Compagno. *Journal of Fish Biology* **72**, 912-936. doi: 10.1111/j.1095-
4300 8649.2007.01771.x
4301

4302 Terraube, J., Arroyo, B., Madders, M., and Mougeot, F. (2011). Diet specialisation and
4303 foraging efficiency under fluctuating vole abundance: a comparison between
4304 generalist and specialist avian predators. *Oikos* **120**, 234-244.
4305

4306 Thomas, C.D., Cameron, A., Green, R.E., Bakkenes, M., Beaumont, L.J., Collingham, Y.C.,
4307 Erasmus, B.F.N., de Siqueira, M.F., Grainger, A., Hannah, L., Hughes, L., Huntley,
4308 B., van Jaarsveld, A.S., Midgley, G.F., Miles, L., Ortega-Huerta, M.A., Peterson,
4309 A.T., Phillips, O.L., and Williams, S.E. (2004). Extinction risk from climate change.
4310 *Nature* **427**, 145-148.
4311

4312 Thorburn, P.J., Biggs, J.S., Attard, S.J., and Kemei, J. (2011). Environmental impacts of
4313 irrigated sugarcane production: Nitrogen lost through runoff and leaching.
4314 *Agriculture, Ecosystems & Environment* **144**, 1-12. doi.
4315 <http://dx.doi.org/10.1016/j.agee.2011.08.003>
4316

4317 Tieszen, L.L., Boutton, T.W., Tesdahl, K.G., and Slade, N.A. (1983). Fractionation and
4318 turnover of stable carbon isotopes in animal tissues: Implications for $\delta^{13}\text{C}$ analysis of
4319 diet. *Oecologia* **57**, 32-37.
4320

4321 Tilley, A., Lopez-Angarita, J., and Turner, J.R. (2013). Diet reconstruction and resource
4322 partitioning of a Caribbean marine mesopredator using stable isotope bayesian
4323 modelling. *Plos One* ,**e79560** DOI: 10.1371/journal.pone.0079560
4324

4325 Townsend, C.R., and Winfield, I.J. (1985). The application of optimal foraging theory to
4326 feeding behaviour in fish. In 'Fish energetics: new perspectives.' (Eds P. Tytler, and P.
4327 Calow.) pp. 67-98 (John Hopkins University Press: Baltimore.)
4328

4329 Trueman, C.N., McGill, R.A.R., and Guyard, P.H. (2005). The effect of growth rate on tissue-
4330 diet isotopic spacing in rapidly growing animals. An experimental study with Atlantic
4331 salmon (*Salmo salar*). *Rapid Communications Mass Spectrometry* **19**, 3239-3247. doi
4332 10.1002/rcm.2199

- 4333 Ubeda, A., Simpfendorfer, C.A., and Heupel, M.R. (2009). Movements of bonnetheads,
 4334 *Sphyrna tiburo*, as a response to salinity change in a Florida estuary. *Environmental*
 4335 *Biology of Fishes* **84**, 293-303.
- 4336
- 4337 Van Tienderen, P.H. (1991). Evolution of generalists and specialists in spatially
 4338 heterogeneous environments. *Evolution* **45**, 1317-1331.
- 4339
- 4340 Vandermeer, J.H. (1972). Niche theory. *Annual review of ecology and systematics* **3**, 107-
 4341 132. doi:10.2307/2096844
- 4342
- 4343 Verberk, W.C.E.P., Van Der Velde, G., Esselink, H. (2010). Explaining abundance–
 4344 occupancy relationships in specialists and generalists: a case study on aquatic
 4345 macroinvertebrates in standing waters. *Journal of Animal Ecology* **79**, 589-601.
 4346 doi:10.1111/j.1365-2656.2010.01660.x
- 4347
- 4348 Wagenmakers, E.J., and Farrell, S. (2004). AIC model selection using Akaike weights.
 4349 *Psychonomic Bulletin & Review* **11**, 192-196.
- 4350
- 4351 Walker, T. (1981). Seasonal salinity variations in Cleveland Bay, northern Queensland.
 4352 *Marine and Freshwater Research* **32**, 143-149.
- 4353
- 4354 Walker, D.I., and McComb, A.J. (1992). Seagrass degradation in Australian coastal waters.
 4355 *Marine Pollution Bulletin* **25**, 191-195.
- 4356
- 4357 Waltrick, D., Awruch, C., and Simpfendorfer, C (2012). Embryonic diapause in the
 4358 elasmobranchs. *Reviews in Fish Biology and Fisheries* **22**, 849-859.
- 4359
- 4360 Waycott, M., Duarte, C.M., Carruther, T.J.B., Orth R.J., Denison, W.C., Olyarnik, S.,
 4361 Calladine, A., Fourqurean, J.W., Heck, K.L. Jr., Hughes, A.R., Kendrick, G.A.,
 4362 Kenworthy, W.J., Short, F.T., and Williams, S.L. (2009). Accelerating loss of seagrass
 4363 across the globe threaten coastal ecosystems. *PNAS* **106**, 12377-12381.
- 4364
- 4365 Weidel, B.C., Carpenter, S.R., Kitchell, J.F., and Vander Zanden, M.J. (2011). Rates and
 4366 components of carbon turnover in fish muscle: insights from bioenergetics models and
 4367 a whole-lake ¹³C addition. *Canadian Journal Fisheries and Aquatic Sciences* **68**, 387-
 4368 399. doi 10.1139/f10-158
- 4369
- 4370 Weng, K., Boustany, A., Pyle, P., Anderson, S., Brown, A., and Block, B. (2007). Migration
 4371 and habitat of white sharks (*Carcharodon carcharias*) in the eastern Pacific Ocean.
 4372 *Marine Biology* **152**, 877-894. doi:10.1007/s00227-007-0739-4
- 4373
- 4374 Weng, K.C., Castilho, P.C., Morrissette, J.M., Landeira-Fernandez, A.M., Holts, D.B.,
 4375 Schallert, R.J., Goldman, K.J., and Block, B.A. (2005). Satellite tagging and cardiac
 4376 physiology reveal niche expansion in salmon sharks. *Science* **310**, 104-106.
 4377 doi:10.1126/science.1114616
- 4378
- 4379 Weng, K.C., Foley, D.G., Ganong, J.E., Perle, C., Shillinger, G.L., and Block, B.A. (2008).
 4380 Migration of an upper trophic level predator, the salmon shark *Lamna ditropis*,
 4381 between distant ecoregions. *Marine Ecology Progress Series* **372**, 253-264. doi
 4382 10.3354/meps07706

4383
4384 Werry, J.M., Lee, S.Y., Otway, N.M., Hu, Y., and Sumpton, W. (2011). A multi-faceted
4385 approach for quantifying the estuarine-nearshore transition in the life cycle of the bull
4386 shark, *Carcharhinus leucas*. *Marine and Freshwater Research* **62**, 1421-1431.
4387
4388 Werry, J.M., Planes, S., Berumen, M.L., Lee, K.A., Braun, C.D., and Clua, E. (2014). Reef-
4389 fidelity and migration of tiger sharks, *Galeocerdo cuvier*, across the Coral Sea. *PLoS*
4390 *ONE* **9**, e83249. doi:10.1371/journal.pone.0083249.
4391
4392 West, J.B., Bowen, G.J., Cerling, T.E., and Ehleringer, J.R. (2006). Stable isotopes as one of
4393 nature's ecological recorders. *Trends in Ecology and Evolution* **21**, 408-414.
4394
4395 Wetherbee, B.M., Gruber, S.H., and Cortés E (1990). Diet, feeding habits, digestion and
4396 consumption in sharks with special reference to the lemon shark *Negaprion*
4397 *brevirostris*. In 'Elasmobranchs as living resources: advances in the biology, ecology,
4398 systematics, and the status of the fisheries.' (Eds H.L. Pratt Jr, S.H. Gruber, and T.
4399 Taniuchi.) NOAA Technical Report NMFS 90.
4400
4401 Whitney, N., Pyle, R., Holland, K., and Barcz, J. (2012). Movements, reproductive
4402 seasonality, and fisheries interactions in the whitetip reef shark (*Triaenodon obesus*)
4403 from community-contributed photographs. *Environmental Biology of Fishes* **93**, 121-
4404 136. doi:10.1007/s10641-011-9897-9
4405
4406 Wilga, C.A.D., Stoehr, A.A., Duquette, D.C., and Allen, R.M. (2012). Functional ecology of
4407 feeding in elasmobranchs. *Environmental Biology of Fishes* **95**, 155-167.
4408 doi:10.1007/s10641-011-9781-7
4409
4410 Williams, S.E., Shoo, L.P., Isaac, J.L., Hoffmann, A.A., and Langham, G. (2008). Towards
4411 an integrated framework for assessing the vulnerability of species to climate change.
4412 *PLoS Biol* **6**, e325. doi 10.1371/journal.pbio.0060325
4413
4414 Wilson, S.K., Burgess, S.C., Cheal, A.J., Emslie, M., Fisher, R., Miller, I., Polunin, N.V.C.,
4415 and Sweatman, H.P.A. (2008) Habitat utilization by coral reef fish: implications for
4416 specialists vs. generalists in a changing environment. *Journal of Animal Ecology* **77**,
4417 220-228.
4418
4419 Wilson, D.S., and Yoshimura, J. (1994). On the coexistence of specialists and generalists.
4420 *American Naturalist* **144**, 692-707.
4421
4422 Wilson, S.K., Burgess, S.C., Cheal, A.J., Emslie, M., Fisher, R., Miller, I., Polunin, N.V.C.,
4423 and Sweatman, H.P.A. (2008). Habitat utilization by coral reef fish: implications for
4424 specialists vs. generalists in a changing environment. *Journal of Animal Ecology* **77**,
4425 220-228.
4426
4427 Yamaguchi, A., and Taniuchi, T. (2000). Food variations and ontogenetic dietary shift of the
4428 starspotted-dogfish *Mustelus manazo* at five locations in Japan and Taiwan. *Fisheries*
4429 *Science* **66**, 1039-1048.
4430

4431 Yates, P., Heupel, M., Tobin, A., and Simpfendorfer, C. (2012). Diversity in young shark
4432 habitats provides the potential for portfolio effects. *Marine Ecology Progress Series*
4433 **458**, 269-281. doi 10.3354/meps09759
4434
4435