



## INTRODUCTION

# Tagging through the stages: technical and ecological challenges in observing life histories through biologging

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**ABSTRACT:** Biologging data have provided important insights into the biology of marine mammals, sea turtles, birds, fish, and some invertebrates. These techniques have primarily targeted adult organisms. As a result, the early life histories of many marine species are still poorly understood. Technological advances have enabled attachment of smaller tags to young animals, although equipment limitations, access to and capture/handling of animals, and equipment and data recovery pose additional challenges to researchers. In this Theme Section, we highlight novel uses of biologging data on juvenile animals, including reviews of tagging efforts on multiple life-history stages and the integration of oceanographic data in tagging efforts.

**KEY WORDS:** Electronic tags · Tag attachment techniques · GPS tags · Juveniles · Ocean currents · Ontogeny · Hatchling dispersal models · Satellite telemetry · Conservation

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Broad-scale anthropogenic impacts, such as climate change, affect all life stages of a species (Brander 2010, Costa et al. 2010, Evans et al. 2010, Fuentes et al. 2011). Advances in understanding the physiology, behavior, and ecology of many marine species have been made through the use of biologging techniques (Mate et al. 2007, Wilson et al. 2008, Ropert-Coudert et al. 2009, Block et al. 2011), which involve attaching electronic tags to animals (Hooker et al.

2007, Rutz & Hays 2009). These techniques have primarily targeted adult organisms, which tend to have higher survival rates than younger life stages. In addition, larger tags can hold more sensors, greater data storage, and larger batteries, resulting in longer deployments (McConnell et al. 2010). Biologging data have provided important insights into the biology of marine mammals, sea turtles, birds, fish, and some invertebrates (Godley et al. 2008, Hays et al.

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2008, Rutz & Hays 2009, Bograd et al. 2010). However, there are few tagging studies on young life stages, and the early life histories of many marine species are still poorly understood (Hazen et al. 2012).

Improvements in biologging technology have resulted in substantial tag miniaturization and enhanced data compression (Fedak et al. 2002). Although it is now possible to attach small tags to young animals (Mansfield et al. 2012), the small size of these tags limits the number of sensors and length of battery life. Thus, it is essential that careful thought goes into what research questions need to be answered and what types of, and how much, data need to be collected to answer these questions (Breed et al. 2011). Attachment mechanisms, durations, and data needs vary greatly across taxa. Young life stages tend to grow rapidly, often requiring attachment methods that are flexible enough to accommodate growth. Assessment in the field or laboratory of suitable attachment methods is necessary to ensure that any tagging effort has minimal impact on the animals and that the benefits of tagging are not outweighed by the costs (Wilson & McMahon 2006, Mansfield et al. 2012).

High mortality during early life history stages makes it difficult to recover tags, may shorten the period of data collection, and may compromise the data set when predation occurs (Hays et al. 2007, Baker 2008, Snoddy & Southwood Williard 2010). Deploying a large number of tags may therefore be necessary to ensure sufficient sample size, particularly if population processes are of interest (Lindberg & Walker 2007). Sample sizes in biologging are generally limited by available funds, so this may require the use of simpler, more inexpensive tags, and careful selection of the type of sensors they contain.

The Editors of this Theme Section convened a workshop entitled 'Tagging through the Stages: Technical and Ecological Challenges in Observing Life Histories through Biologging' on 16 March 2011 in association with the Biologging IV Symposium in Hobart, Tasmania, Australia, with these technical and ecological challenges in mind. Over 40 scientists and tag manufacturers from Australia, North America, Europe, and Japan attended the workshop. The objectives were to (1) establish the current state of knowledge and technologies for studying young life stages, (2) stimulate interdisciplinary discussion regarding ontogeny and biologging, (3) review and discuss tag design and attachment techniques, and (4) integrate a life history perspective within the field of biologging. There were 18 presentations organized in 3 categories: (1) Tag Techniques and Devel-

opment, (2) Applications, and (3) Models. The studies within this Theme Section represent a selection from the workshop and feature recent advances in our understanding of the life history of marine species and in particular young life stages.

### Tag techniques and development

Advances in tagging technology, such as tag miniaturization, increased data storage and transmission capabilities, and improved analytical methodologies, are providing researchers with important tools for understanding the biology of marine species and their environment (Fedak et al. 2002). The development of fast-acquisition GPS tags, such as Fastloc GPS, has provided increasingly accurate location estimates for species that spend little time at the surface. Shimada et al. (2012) propose a new filtering method for these data that will reduce the linear error of Fastloc GPS locations to 47 m while retaining more than 94 % of the data. This increases the accuracy of home range estimates.

Ethical issues of whether the benefits of the scientific research outweigh the costs of disturbance and possible harm to the animals are a key concern when catching and tagging juvenile animals (Godley et al. 2008). In some cases, young animals may be too small or delicate to carry tags and less invasive research techniques are more appropriate. Assessment of the best techniques and practices will help to ensure that any impacts to the animals are kept to a minimum. Mansfield et al. (2012) describe the first satellite tracks of any neonate sea turtle and the first *in situ* data of the movements of neonate loggerheads *Caretta caretta*. They tested several methods for attaching small solar-powered satellite tags, first in the laboratory to determine whether there were any apparent effects on growth or body condition, and then in the field to establish retention durations and to assess the performance of antifoulants (Mansfield et al. 2012). There is increasing evidence that general guidelines and practices may not well reflect species-specific and longer-term harmful tag effects (Sherrill-Mix & James 2008, Vandenabeele et al. 2012).

Assessing effects of drag may also be possible using computer simulations, to determine how an animal's physiology or morphology may assist with creating hydrodynamic tag designs and attachment techniques (Pavlov & Rashed 2012). Abnormal swimming behavior and increased energetic demands can occur if the tag causes high drag (Hammerschlag et al. 2011, Jones et al. 2011). Designing tags and

attachments that will reduce drag is therefore of key importance for marine species that rely on swimming to capture prey or escape predators, and during flight for seabirds (Phillips et al. 2003, Wilson & McMahon 2006, Heithaus et al. 2007, Vandenabeele et al. 2012).

Accommodating the rapid rate of growth in young life stages remains a challenge for tag attachment techniques. Implantable tags may be necessary for studying animals over longer time periods and across life history stages (Horning & Hill 2005). Implantable tags have been effectively used in fish (Block et al. 2005). For example, data on depth and temperature have been used to study changes during different stages of the breeding migration in bluefin tuna *Thunnus thynnus* (Teo et al. 2007).

### Applications

Biologging has contributed significant insights that inform conservation management and population recovery efforts by identifying important habitats, risks and impacts, and helping to plan effective mitigation measures (Mate et al. 2007, Shillinger et al. 2008, McClellan et al. 2009). This has included the ability to identify life history changes and potential harvest events. Hart et al. (2012) describe the habitat use of juvenile hawksbill turtles *Eretmochelys imbricata* that had core-use areas within a national park. Two of the tagged turtles migrated from the exclusive economic zone of the USA into Cuban waters, where they may have been harvested.

Neonate, juvenile, and subadult animals tend to behave and move differently than do adults, which may be a result of reduced diving and swimming capabilities early in life, and/or may reduce competition for resources (Campagna et al. 2007). Protection by the mother can make it difficult to tag some young animals, and their unpredictable movements can limit tag recovery and thus bias estimates of survival (Bradshaw et al. 2000). Tyson et al. (2012) attached high-resolution digital acoustic recording tags (Dtags) to a mother and calf humpback whale *Megaptera novaeangliae* pair in Wilhelmina Bay (Western Antarctic Peninsula) to examine their concurrent diving and foraging behavior. The pair appeared to dive in synchrony for much of the tag duration while maintaining close proximity (Tyson et al. 2012). These results validate findings that humpback whale calves accompany their mothers following parturition, remaining within several body lengths until they separate permanently (Szabo & Duffus 2008).

Although the number of biologging studies on immature animals has been small relative to studies on adults (Hazen et al. 2012), an increasing number of recent studies, such as those on loggerhead turtles (Mansfield et al. 2009, 2012, Seney et al. 2010), green turtles *Chelonia mydas* (Hart & Fujisaki 2010), and flatback turtles *Natator depressus* (Salmon et al. 2009), have coupled advances in tag miniaturization with innovative deployment techniques to obtain critical information about the dispersal and movements of juvenile animals. Data regarding this early life history phase are essential because it represents a large proportion of the life span for many long-lived species. Moreover, experiences during these periods are often diverse as these animals undergo transition between nursery, foraging, and breeding habitats, which can exert a strong influence upon the population status.

Barbour & Adams (2012) used passive integrated transponders and found that common snook *Centropomus undecimalis* had high site fidelity within specific life history stages, but changed habitat as they transitioned from juvenile to adult. In another example, Melnychuk et al. (2012) utilized acoustic tagging methods to examine the hypothesis that exposure of coho *Oncorhynchus kisutch* and sockeye salmon *O. nerka* to solar UV-B radiation during freshwater rearing of fry and parr increases mortality at the time of smoltification and ocean entry. They found that while exposure to UV-B resulted in stunted growth of juvenile coho salmon, survivorship during the early marine period was unaffected by the UV-B treatment for both populations. These results challenge one of the many hypotheses for declines in marine survival rates of salmon populations.

### Models

There are cases, such as for very young animals, where biologging may not currently be the most appropriate technique or the study design could be improved by first gaining some basic knowledge about when and where the animals are going and how they are dispersing. Ocean models provide a valuable resource for investigating potential dispersal patterns and have been of particular use in studying sea turtles (Hays & Marsh 1997, Hamann et al. 2011, Scott et al. 2011). Very little is known about the movements of hatchling turtles after they leave the beach, until they return as adults decades later. As adults, sea turtles show fidelity to their natal nesting areas and then at the end of the breeding season may migrate to distant foraging sites. Hays et al. (2010)

hypothesized a hatchling drift scenario whereby the foraging sites used by individual sea turtles reflect their previous experiences as young juveniles, when they were carried by ocean currents. The pattern of adult dispersion from the breeding area for loggerhead turtles in the Mediterranean reflects the extent of passive drift experienced by hatchlings (Hays et al. 2010). Simulations of leatherback hatchling dispersal in the western Pacific by Gaspar et al. (2012) similarly found that adults targeted favorable foraging areas inside the juveniles' drift area. However, there were drift areas where no adults have been observed (Gaspar et al. 2012). This could occur as a result of high juvenile mortality along drift trajectories towards such areas, or because of a low return rate if the return route to the natal area is difficult to traverse or navigate. Shillinger et al. (2012) investigated leatherback turtle hatchling dispersal in the eastern Pacific Ocean using passive tracer experiments within a Regional Ocean Modeling System. Tracer distribution suggested that hatchling leatherbacks entering the ocean in late winter are rapidly and efficiently transported offshore within eddies, which may provide a productive refuge for them.

These simulations of hatchling dispersal based on ocean models can further be refined through laboratory and field-derived estimates of their swimming behavior (Wynneken et al. 2008, Okuyama et al. 2009, Salmon et al. 2009). Ocean currents may be critical for determining the dispersal of hatchlings and subsequent migrations as adults. Fossette et al. (2012) reviewed various techniques for estimating current velocities, or more directly, passive drift trajectories. All methods have errors that need to be taken into account when inferring about animal behavior, and in particular, swimming activity (e.g. Jonsen et al. 2005).

### Conclusions

There are several challenges in biologging-related conservation research efforts, including: (1) equipment limitations (i.e. suitable tag attachment methods, device size, finding funding for sufficient sample sizes), (2) access to and capture/handling of animals, (3) equipment and data recovery, and (4) creating the empirical link between science and policy that encourages support by managers. The first 3 of these challenges are magnified for immature life stages, which impose stronger size, weight, and recovery constraints on tags. Some lessons can be learned from terrestrial efforts, such as those involving but-

terflies that carried extremely small, lightweight radar transponders to track their flight paths (Cant et al. 2005). Research involving birds who require small, lightweight tags can also provide insights into marine juvenile biologging techniques (Thorup et al. 2007, Egevang et al. 2010). Our knowledge of life histories can greatly benefit from combining biologging data with information from other sources, such as genetics, stable isotope analyses, modeling, and stranding and catch data (Wallace et al. 2006, Dutton et al. 2007, Godley et al. 2010, Taylor et al. 2011). Similarly, burgeoning efforts to develop high seas marine protected areas, define ecologically or biologically significant areas, and establish adaptively-managed marine reserve networks, will draw upon new tracking research that examines animal movements and behaviors across life history stages (Shillinger et al. 2008, 2010, Game et al. 2009, Dunn 2011).

*Acknowledgements.* The workshop was generously supported by funding from The Cinco Hermanos Foundation, Inter-Research, Collecte Localisation Satellites (CLS), CLS-America, The Leatherback Trust, Wildlife Computers, Desert Star Systems, and Telonics; 5 early career travel grants, funded by Inter-Research, were awarded to G. Blanco, S. Fossette, K. Mansfield, A. Stimpert, and R. Tyson. We are also grateful to the organizers of the Biologging IV Symposium for selecting our workshop to be part of this event.

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