The Functional Morphology, Systematics and Behavioural Ecology of Parrotfishes (Family Scaridae).

Volume 1

Morphology and Systematics

Thesis submitted by David Roy BELLWOOD BSC (Hons.) (Bath, U.K.) in December, 1985

for the degree of Doctor of Philosophy in

the Department of Zoology at

James Cook University of North Queensland,

Australia.

I, the undersigned, the author of this thesis, understand that the following restriction placed by me on access to this thesis will not extend beyond three years from the date on which the thesis is submitted to the University.

I wish to place restriction on access to this thesis as follows:

Access not to be permitted for a period of 3 years.

After this period has elapsed I understand that James Cook University of North Queensland will make it available for use within the University Library and, by microfilm or other photographic means, allow access to users in other approved libraries. All users consulting this thesis will have to sign the following statement:

"In consulting this thesis I agree not to copy or closely paraphrase it in whole or in part without the written consent of the author; and to make proper written acknowledgement for any assistance which I have obtained from it."

A. bellwood.

18.12.1985

ABSTRACT

The functional morphology and behavioural ecology of the parrotfishes (family Scaridae) were investigated to assess the degree of correlation between them.

Twenty-two species of parrotfishes from the subfamily Scarinae were examined mainly from reefs around Lizard Island, Great Barrier Reef, Australia.

Morphological studies of the osteology and myology of the heads of adult parrotfishes revealed two morphological groups within the genus Scarus, viz. the 'sordidus' and 'frenatus' groups. Species in the 'sordidus' group are characterized by: uneven cutting edges on the jaws, an entopterygoid lateral process, a tightly bound interdigitating maxilla and premaxilla, three rows of teeth on the upper pharyngeal bones, well developed adductor mandibulae sections Al, A2 and A3 and an additional unique muscle, the Awy. Species in the 'frenatus' group are characterized by: even cutting edges on the jaws, a slightly protrusible premaxilla, two rows of teeth on the upper pharyngeal bones, fusion of the abductor muscle section Ala and A2 in some species and a thin strap-like A3 section which inserts only on the articular. The morphology of the heads of species in other genera, namely, Cetoscarus bicolor, Bolbometopon muricatum and Hipposcarus longiceps was marked by their possession of a quadrato mandibularis internus muscle and the non-articulatory articular-dentary joint in C. bicolor and B. muricatum.

iii

Analyses of the two morphological groups indicated a marked difference in the functional abilities of species in the two groups. The morphological characteristics of 'sordidus' group species enable them to bite the substratum with a large powerful cracking bite. Species in this group are therefore described as 'biters'. The morphological characteristics of 'frenatus' group species are consistent with the requirements necessary for scraping the substratum with small, weak bites. Species in this group are therefore described as 'scrapers'.

Of the species examined in this study, the following species are functional 'biters': S. bleekert, S. gibbus and S. sordidus, whereas the following are functional 'scrapers': S. brevifilis, S. dimidiatus, S. flavipectoralis, S. frenatus, S. ghobban, S. globiceps, S. longipinnis, S. niger, S. oviceps, S. psittacus, S. rivulatus, S. rubroviolaceus, S. schlegelt, S. spinus, S. tricolor and Scarus sp. (cf. lunula). Because they lack some specialized morphological features associated with the biting or scraping strategies of species in the genus Scarus, Cetoscarus bicolor and Bolbometopon muricatum are considered 'proto-biters', whilst Hipposcarus longiceps is considered a 'proto-scraper'.

These differences are not apparent in juvenile specimens less than 50 mm standard length. Specimens of *Scarus* smaller than this are extremely similar. Small specimens (< 14 mm S.L.), in particular, differ markedly from the adults in the possession of caniform teeth and a simple, non-sacculated intestine.

iv

Functional interpretations of the anatomy of the head and intestine of juvenile scarids suggest that they progress from an initial carnivorous phase to a selective grazing stage before becoming functional 'scrapers' at about 50 mm S.L. 'Biting' species only possess the full complement of biting characteristics above 90 mm S.L.

Field observations of adults revealed two feeding quilds within the genus Scarus which correspond with the two functional groups. (1) 'Biting' *i.e.* 'sordidus' group species are characterized by: i) infrequent, large bites which scar the substratum, ii) a propensity to feed upon convex substrata, and iii) aggressive interspecific interactions, when displayed, predominantly directed towards other 'sordidus' group species. (2) 'Scraping' t.e. 'frenatus' group species are characterized by: i) numerous small bites which rarely scar the substratum, ii) a tendency to feed on a range of substrata, and iii) aggressive interspecific interactions, when displayed, predominantly directed towards other 'frenatus' group species. These feeding strategies strongly influence the roles played by parrotfishes as coral predators and bioeroding agents. The major coral predator is Bolbometopon muricatum. Bioerosion is primarily the result of feeding by 'sordidus' group species, Cetoscarus bicolor and B. muricatum. It is therefore proposed that the distribution of these species within and between reefs may influence the extent and rates of bioerosion, the topography of the substratum and the distribution of various coral morphs.

Field observations of juvenile scarids revealed a wide range of behavioural traits, although these did not correspond with the 'sordidus' and 'frenatus' groupings. *S. frenatus* in particular, differed from other juvenile 'frenatus' group species in its feeding behaviour. An analysis of the gut contents of the juveniles of several species showed marked changes in the diet, from being initially predominantly carnivorous to herbivorous. These changes were strongly correlated with changes in morphology and behaviour.

νi

In addition to functional considerations, the morphological analyses in this study were used to examine the systematic relationships between genera in the subfamily Scarinae. The present generic status of *Cetoscarus*, *Bolbometopon* and *Hipposcarus* is supported. The genus *Scarus* contains two distinct groups which may be recognized at the generic level.

A descriptive account of the ontogeny of the colour patterns of the juvenile phase of 22 species is presented to facilitate the study of juvenile scarids in the field. The colour patterns of juvenile scarids are interpreted in terms of concealment colouration, including crypticism, camouflage and mimicry. In the species examined, there is a strong correlation between schooling behaviour, the range of patterns displayed, and the speed with which the patterns may be changed. TABLE OF CONTENTS

ABSTRACT	iii
TABLE OF CONTENTS	Vii
LIST OF FIGURES	ix
LIST OF TABLES	xv
ACKNOWLEDGEMENTS	xviii
GENERAL INTRODUCTION	1
PART I MORPHOLOGY (Volume 1)	
Chapter 1 THE MORPHOLOGY OF ADULT SCARIDS	5
1.1 Introduction	5
1.2 Materials, Methods and Terminology	6
A - The Muscular and Skeletal System	
 1.3 Observations 1.3.1 External appearance and integument 1.3.2 Osteology 1.3.3 Myology 1.3.4 The 'sordidus' and 'frenatus' groups 1.3.4.1 The 'sordidus' group 1.3.4.2 The 'frenatus' group 1.3.5 Cetoscarus bicolor 1.3.6 Bolbometopon muricatum 1.3.7 Hipposcarus longiceps 1.4 Functional Interpretations 4.1 'Sordidus' group species as 'biters' 4.2 'Frenatus' group species as 'scraper 4.3 C. bicolor and B. muricatus 4.4 Hipposcarus longiceps 4.5 The pharyngeal apparatus 	11 12 38 61 63 73 86 96 107 107 107 55' 115 120 123 123
1.5 Observations	132
1.6 Functional Interpretations	139
1.7 Discussion	150
Chapter 2 THE MORPHOLOGY OF JUVENILE SCARIDS	153
 2.1 Introduction 2.2 Materials and Methods 2.3 Observations 2.4 Functional Interpretations 2.5 Discussion 	153 155 155 173 178

		PART II SYSTEMATICS (Volume 1)	
Chantar 3	SYCHEM	MITCE OF MER SCAPIDAR.	
cuapter 3	PHYL	OGENY AND GENERIC CLASSIFICATION	184
Chapter 4	SYSTEM	ATICS OF THE SCARIDAE:	
	ONTO	GENY OF JUVENILE COLOUR PATTERNS	209
	4.1	Introduction	209
	4.2	Materials and Methods	213
	4.3	Results	214
		4.3.1 Meristic data	214
		4.3.2 Field observations	215
		4.3.3 Colour patterns	217
		4.3.4 Species descriptions	223
	4.4	Discussion	277
		PART III ECOLOGY (Volume 2)	
Chantor 5		WAUTOIDAL POLOCY OF ADIT SCAPIDS	200
cuapter p		RAVIOURAL ECOLOGI OF ADULI SCARIDS	290
		A - Feeding Biology	
	C 1	Tetroduction	200
	5.2	Materials and Methods	296
	5.3	Results	299
	5.4	Discussion	312
		B - General Biology	
	5.5	Introduction	318
	5.6	Materials and Methods	321
	5.7	Results	322
	5.8	Discussion	330
Chapter 6	THE BE	HAVIOURAL ECOLOGY OF JUVENILE SCARIDS	364
		Part A - Feeding Biology	
		······································	
	6.1	Introduction	364
	6.2	Materials and Methods	365
	6.3	Results	369
	6.4	Discussion	384
		Part B - General Biology	
	6.5	Introduction	392
	6.6	Materials and Methods	394
	6.7	Results	399
	6.8	Discussion	423
	CONCLU	DING DISCUSSION	434
	APPEND	IX 1	454

APPENDIX 2 455

REFERENCES

LIST OF FIGURES

Figure		Page
1.1 A	The external anatomy of the head of Scarus sordidus	13
1.1 B	The external anatomy of the head of Scarus frenatus	.13
1.2	The osteology of the head of Scarus sordidus	. 15
1.3	The osteology of the head of Scarus frenatus	.16
1.4 A	A lateral view of the suspensorium of S. sordidus	.18
1.4 B	A lateral view of the suspensorium of S. frenatus	.18
1.5 A	A lateral view of the maxilla of S. sordidus	. 22
1.5 B	A medial view of the maxilla of S. sordidus	22
1.5 C	A lateral view of the maxilla of S. sordidus	. 22
1.5 D	A medial view of the maxilla of S. frenatus	. 22
1.6 A	A lateral view of the premaxilla of S. sordtaus	. 25
1.6 B	A lateral view of the premaxilla of S. frematus	. 25
1.6 C	A ventro-medial view of the premaxilla of S. sordidus.	. 25
1.6 D	A ventro-medial view of the premaxilla of S. frenatus.	. 25
1.7 A	A lateral view of the articular of S. sordidus	. 29
1.7 B	A medial view of the articular of S. frenatus	. 29
1.7 C	A lateral view of the articular of S. sordtdus	. 29
1.7 D	A medial view of the articular of S. frenatus	29
1.8 A	A lateral view of the dentary of S. sordidus	. 32
1.8 B	A lateral view of the dentary of S. frenatus	32
1.8 C	A medial view of the dentary of S. sordidus	. 32
1.8 D	A medial view of the dentary of S. frenatus	32
1.9 A	An anterior view of the dentary cutting edge of S. sordidus	33
1.9 B	An anterior view of the dentary cutting edge of S. frenatus	33

ix

1.10	The head of scarus sordidus with the integument, nasal, lacrymal, circumorbitals, Al α and part of Al β removed, showing the shading conventions used in this section39
1.11 A	The head of <i>S. sordidus</i> with the integument and circumorbitals removed41
1.11 B	The head of S. sordidus with the integument, nasal, lacrymal, circumorbitals, Al α and part of Al β removed41
1.12 A	The head of S. frenatus with the integument removed45
1.12 B	The head of S. frenatus with the integument, nasal, lacrymal, circumorbitals and parts of $Al\alpha/A2$ removed45
1.13 A	A medial view of the upper jaw musculature of S. sordidus
1.13 B	A medial view of the upper jaw musculature of S. frenatus
1.14 A	A medial view of the lower jaw musculature of S. sordidus
1.14 B	A medial view of the lower jaw musculature of S. frenatus
1.15 A	A lateral view of the pharyngeal apparatus of S. sordidus
1.15 B	A lateral view of the pharyngeal apparatus of S. frenatus
1.16	The head of a 388 mm S.L. <i>Cetoscarus bicolor</i> with the integument, nasal, lacrymal, circumorbitals and part of Al removed79
1.17 A	A lateral view of the neurocranium of Bolbometopon muricatum
1.17 B	A lateral view of the neurocranium of <i>Cetoscarus</i> bicolor
1.18 A	A ventral view of the neurocranium of <i>Cetoscarus</i> bicolor
1.18 B	A ventral view of the neurocranium of Bolbometopon muricatum
1.18 C	A ventral view of the neurocranium of <i>Hipposcarus</i> longiceps
1.18 D	A ventral view of the neurocranium of Scarus sordtdus81
1.18 E	A ventral view of the neurocranium of Scarus frenatus81

x

1.19	The head of a 530 mm S.L. Bolbometopon muricatum with the integument, nasal, lacrymal, circumorbitals and part of Al removed92
1.20	The head of a 329 mm S.L. <i>Hipposcarus longiceps</i> with the integument, nasal, lacrymal and circumorbitals removed101
1.21 A	A lateral view of the neurocranium of <i>Htpposcarus</i> longiceps102
1.21 B	A lateral view of the neurocranium of S. sordtaus102
1.21 C	A lateral view of the neurocranium of S. frenatus102
1.22 A	The dentary of <i>S. sordidus</i> showing the theoretical forces acting upon the dentary during muscle
	contraction
1.22 B	A diagramatic figure describing the relationship between the angle of a muscle and its mechanical
	efficiency114
1.23	A diagramatic figure describing the properties of
	muscles with weakly pinnate or strongly pinnate fibres
1.24	The three types of intestinal pattern found in <i>Scarus</i> species136
2.1 A	A lateral view of the anterior edge of the premaxilla of a 9.0 mm S.L. S. sordidus
2.1 B	A lateral view of the anterior edge of the premaxilla of a 8.7 mm S.L. S. frenatus
2.2	The head of <i>S. sordidus</i> at 25.0 mm S.L. with the integument, nasal, lacrymal and circumorbitals
	removed161
2.3	The head of <i>S. frenatus</i> at 26.5 mm S.L. with the integument, nasal, lacrymal and circumorbitals
	removed165
3.1	A cladogram showing the proposed phylogeny of the genera in the subfamily Scarinae
3.2	The phylogeny of the genera in the family Scaridae as proposed by Schultz (1958)
4.1	A series of diagrams showing the ontogenetic changes in the colour pattern of juvenile <i>Scarus</i> species leading to the formation of the four primary stripes shown in Figure 4.2218

4.2	The 'typical' colour pattern of a juvenile <i>Scarus</i> species showing the position of the four primary dark longitudinal stripes and the six vertical bands219
4.3 A	S. bleekeri : 33 mm S.L242
4.3 B	5. bleekeri : 46 mm S.L242
4.3 C	S. bleekert : 54 mm S.L242
4.4 A	S. gibbus : 24 mm S.L243
4.4 B	S. gibbus : 28.5 mm S.L243
4.4 C	S. gtbbus : 40 mm S.L243
4.5 A	S. sorātāus : 25 mm S.L244
4.5 B	S. sordidus : 40 mm S.L244
4.5 C	S. brevifilis : 26 mm S.L244
4.6 A	S. brevifilis : 28.5 mm S.L
4.6 B	S. brevifilis : 54 mm S.L245
4.6 C	S. brevifilis : 55 mm S.L245
4.7 A	S. frenatus : 10.5 mm S.L246
4.7 B	S. frenatus : 22 mm S.L246
4.7 C	S. frenatus : 31 mm S.L246
4.8 A	S. frenatus : 55 mm S.L247
4.8 B	S. flavipectoralis : 33 mm S.L
4.8 C	S. flavipectoralis : 44 mm S.L247
4.9 A	S. dimidiatus : 30.5 mm S.L248
4.9 B	S. oviceps : 32 mm S.L
4.9 C	S. oviceps : 73 mm S.L
4.10 A	S. ghobban : 53 mm S.L249
4.10 B	S.longipinnis : 39 mm S.L249
4.10 C	S. longipinnis : 75 mm S.L249
4.11 A	S. niger : 9.5 mm S.L250
4.11 B	S. niger : 24 mm S.L

4.11 C	S. niger : 34 mm S.L
4.12 A	S. niger : 57 mm S.L251
4.12 B	S. psittacus : 29.5 mm S.L
4.12 C	S. tricolor(cf. S. forsteni) : 61 mm S.L251
4.13 A	S. rivulatus : 49 mm S.L252
4.13 B	S. schlegelt : 37 mm S.L252
4.13 C	S. schlegelt : 38 mm S.L252
4.14 A	S. spinus : 20 mm S.L253
4.14 B	S. spinus : 22 mm S.L253
4.14 C	S. spinus : 27 mm S.L253
4.15 A	S. rubroviolaceus : 18 mm S.L
4.15 B	S. rubroviolaceus : 22.5 mm S.L
4.15 C	S. rubruviolaceus : 38 mm S.L
4.16 A	S. sp. (cf. lunula)? : 10.3 mm S.L
4.16 B	S. sp. (cf. lunula)? : 33 mm S.L
4.16 C	Cetoscarus bicolor : 11 mm S.L
4.17 A	Hipposcarus longiceps : 48 mm S.L256
4.17 B	Calotomus carolinus : 43 mm S.L256
4.17 C	Calotomus carolinus : 43 mm S.L256
4.18 A	A ventral view of the upper pharyngeal tooth rows of a 128 mm S.L. S. globiceps
4.18 B	A ventral view of the upper pharyngeal tooth rows of a 128 mm S.L. <i>S. rivulatus</i> 260
5.1	The location of the major study sites in the lagoon and at North Reef, Lizard Island
5.2 A	A diagramatic figure of the three parameters measured when quantifying scarid feeding scars298
5.2 B	A diagramatic figure showing the disproportionate effect of an increase in the bite length on the ratio of H/L (bite height/bite length)298
5.3	The foray sizes of two Scarus species

xiii

6.1 A	The mean feeding rates of three juvenile <i>Scarus</i> species between 20 and 120 mm T.L
6.1 B	The mean foray size of three juvenile <i>Scarus</i> species between 20 and 120 mm T.L
6.2 A	The intestinal contents of juvenile scarids; algae and sand
6.2 B	The intestinal contents of juvenile scarids; crustacea
6.3	A summary of the ontogenetic changes in the behaviour, diet and morphology of juvenile scarids385
6.4	The relative abundance of juvenile scarids at the North Reef and lagoon study sites during the 1982-3 and 1983-4 November-January recruitment periods400
6.5	The relative abundance of juvenile scarids at three depths in the North Reef study site and at two depths in the lagoon study site, during the 1982-3 November- January recruitment period401
6.6	The relationship between home range/territory area and total length of juvenile scarids406
6.7	The home ranges of two juvenile <i>S. sordidus</i> in the lagoon study site407
6.8	A vertical profile of the primary study area at the North Reef study site, showing the distribution of home ranges/territories of juvenile scarids in the study area412
6.9 A	The relative rates of aggression by pomacentrids towards juvenile scarids in the lagoon study site419
6.9 B	The relative rates of aggression by pomacentrids and Acanthurus lineatus towards juvenile S. frenatus in the lagoon study site419
6.10	The feeding preferences of juvenile scarids: The mean number of juvenile scarids feeding on six experimental algal-covered substratum types422
7.1 A	The proposed distribution of early scarids during the late Eocene period448
7.1 B	The proposed distribution of the major scarid groups approximately 4 million years ago
7.1 C	The present distribution of the major scarid groups 448

LIST OF TABLES

Table	Page
1.1	A partial synonomy of the Scaridae of the Great Barrier Reef7
1.2	The morphology of adult scarids: Material examined8
1.3	Abbreviations used in the figures
1.4	A comparative list of relative liver weights134
1.5 A	The liver weights of two Scarus species
1.5 B	The intestinal length vs. standard length of two Scarus species
1.5 C	The intestinal length vs. body weight of two Scarus species
1.6	The distribution of intestinal types in <i>Scarus</i> species
1.7	Gut lengths of herbivorous reef fishes143
2.1	The morphology of juvenile scarids: material examined
2.2	Ontogenetic changes in the morphology of the intestinal tract of juvenile scarids
2.3	The functional morphology of juvenile scarids: a summary
3.1	The distribution of morphological characters among the scarid genera
4.1	Variability in the colour patterns of some juvenile <i>Scarus</i> species
5.1	The feeding rates of six Scarus species
5.2	Observations of the feeding rates and the extent of scarring of the substratum by scarids
5.3	The size of substratum scars which result from the grazing activity of scarids
5.4	Patterns of substratum utilization by S. sordidus and S. frenatus
5.5 A	An analysis of the curvature of the substrata grazed by <i>S. sordidus</i> and <i>S. frenatus</i> 308
5.5 B	A comparison of the bite-scar lengths of S. sordidus and S. frenatus

xv

5.6	Coral species eaten by scarids
5.7	A summary of the morphological and behavioural characteristics of 'sordidus' and 'frenatus' group species
5.8 A	Duration and size of home ranges/territories of scarids: A - known individuals, at North Reef323
5.8 B	Duration and size of home ranges/territories of scarids: B - permanent sites, unknown individuals, at North Reef
5.9	Aggressive interactions between scarids
5.10	Differences in the extent of interspecific aggression by terminal and initial phase S. frenatus
5.11 A	Estimated scarid bioerosion rates at North Reef, Lizard Island
5.11 B	Scarid abundances: a comparison between Caribbean and Indo-Pacific reefs
5.12 A	Estimates of bioerosion by scarids: a comparison between the Caribbean and the Great Barrier Reef
5.12 B	Rates of bioerosion: a summary
6.1	Functional interpretations of the morphology of juvenile scarids
6.2	Size-related differences in the feeding rates of three juvenile <i>Scarus</i> species between 20 and 120 mm T.L
6.3 A	Changes in the bite rate and foray size of <i>S. sordidus</i> between 10 and 19.9 mm T.L
6,3 B	The feeding rates of <i>S. sordidus</i> and <i>S. frenatus</i> between 20 and 120 mm T.L
6.3 C	The mean number of bites per foray of <i>S. sordidus</i> and <i>S. frenatus</i> between 20 and 120 mm T.L
6.4	Qualitative observations of the feeding rates of juvenile scarids between 40 and 110 mm T.L373
6.5	Size-related differences in the mean foray size of three juvenile <i>Scarus</i> species between 20 and 120 mm T.L
6,6	The intestinal contents of juvenile scarids - A: Algae, sand and crustacea

6.7	The intestinal contents of juvenile scarids - B: Crustacea
6.8	The intestinal contents of some juvenile herbivorous reef fishes other than scarids
6.9	Relative rates of recruitment at the lagoon and North Reef study sites during three settlement seasons
6.10	A list of the scarid speies present as new recruits and juveniles in the vicinity of the lagoon and North Reef study sites
6.11	The duration and nature of site attachment in juvenile scarids410
6.12	The size of early post-recruit scarids over various substratum types in the North Reef and lagoon study sites415
6.13	Aggressive interactions of three Scarus species417
6.14	Aggressive interactions of juvenile scarids418
6.15	The relative frequency of attacks upon juvenile scarids by various damselfish species at the North Reef and lagoon study sites420
7.1	A summary of the biological factors influencing the three post-larval life stages of scarids443

ACKNOWLEDGEMENTS

Facilities for this study were provided by the School of Biological Sciences, James Cook University, and I am grateful to the former Head of the School, Professor C. Burdon-Jones, the present Head of Zoology, Professor R.E. Jones, and other staff members for their assistance.

I would like to thank my supervisor, Dr. N.E. Milward for his support and guidance.

Most of the field work in this study was undertaken at Lizard Island Research Station, a facility of the Australian Museum. My time at the Lizard Island was both pleasant and rewarding as a result of the kindness, humour and fellowship of the people there. In particular I wish to extend my gratitude to Hugh Sweatman, Bill Gladstone, Jeff Leis, Jack Randall, Vicki Harriott, Lyle Vail, Randy Olson, Patrick Filmar-Sankey, John Chisolm, Geoff Smith and Roger am particularly endebted to Dr. J.H. Choat. Steene. Ι His encouragement and infectious enthusiasm was a source of much My visits to Lizard Island were both comfortable and inspiration. productive, primarily as a result of the thoughtfulness and painstaking efforts of Lois and Barry Goldman.

Specimens were kindly donated by: Drs G.R. Russ and (Australian D.McB. Williams Institute of Marine Science); Dr. J.H. Choat (University of Auckland); Dr. P.D. Doherty (Griffith University); Dr. J.E. Randall (Bernice P. Bishop Museum); A. Cabanban (Silliman University); Dr. G. Denton, O.S. Perez, L.L. Dolar, R.S. Smith and J. Chisolm (J.C.U.).

I wish to thank the following people for their helpful discussions and/or comments on various drafts of this thesis: Drs C. Alaxander, P. Arnold, P. Doherty, D.G. Reid, G.R. Russ, D.McB. Williams, Associate Professor R. Kenny, Professor R.E. Jones and my collegues at J.C.U. and Lizard Island.

Some publications were kindly translated by Dirk Zeller and Tomoko Yoda.

This project was supported by a scholarship from the Drapers' Company London, to whom I am extremely grateful. Additional financial support was kindly provided by the Great Barrier Reef Marine Park Authority. The completion of this thesis would not have been possibly without the timely financial assistance of my parents who supported me during the latter part of my studies.

I wish to thank Orpha S. Perez for her friendship and invaluable assistance in the preparation of this thesis.

Finally, I would like to thank the two people whose thoughts and guidance throughout my studies have been so valuable. To June and Roy, my parents, this thesis is as much yours as it is mine.

xix

DECLARATION

I declare that this thesis is my own work and has not been submitted in any form for another degree or diploma at any university or other institution of tertiary education. Information derived from the published or unpublished work of others has been acknowledged in the text and a list of references is given.

D.R. Bellwood

D.R. Bellwood December, 1985

GENERAL INTRODUCTION

The Scaridae[†] is a distinctive group of perciform teleosts, characterized by the possession of a pharyngeal mill and fused teeth, which form dental plates. Scarid species have a tropical and sub-tropical distribution and are typically associated with coral reefs. They are predominantly herbivorous and form a conspicuous part of the herbivorous reef fish community. At present, there are 69 recognised species in 12 genera (Schultz, 1969 and Randall & Bruce, 1983).

Members of the Scaridae are predominantly protogynous hermaphrodites, with two main colour phases, a drab initial phase (IP) and a colourful terminal phase (TP) (Robertson & Warner, 1978). Initial phase individuals may be either male or female, whilst terminal phase individuals are invariably male. Some individuals are male throughout their life, during both initial and terminal phases; these are referred to as primary males. Most males, however, are secondary males and, as such, spend the initial phase period as females before developing testes and exhibiting the terminal phase colouration (Choat, 1966, Choat and Robertson, 1975, Robertson and Warner, 1978 and Bruce, 1979). The common name for the family, the parrotfishes, has arisen from the striking colours of the terminal phase and the close resemblance of the dental plates to a parrot's beak.

† The conventional nomenclature is utilized. The recently proposed familial synonomies are discussed in Chapter 3.

Parrotfishes have many unusual characteristics. These include the secretion of a mucous cocoon at night (Winn, 1955, Winn & Bardach, 1959, 1960 and Casimir, 1971), a diet that includes live coral (Hiatt & Strasburg, 1960, Glynn et al., 1972 and Randall & Bruce, 1983), a grinding pharyngeal mill (Al Hussaini, 1945 and Board, 1956), an additional articulation point in the lower jaw (Lubosch, 1923 and Gregory, 1933), an unusual sacculated intestine (Al Hussaini, 1945, 1947 and Gohar & Latif, 1959) and a range of physiological adaptations to the exceptionally high carbonate content of the diet (Fontaine et al., 1973 and Smith & Paulson, 1975). Parrotfishes have been implicated as important agents in bioerosion (Gygi, 1975 and Frydl & Stearn, 1978), sediment transport (Bardach, 1961) and in the regulation of benthic invertebrates (Glynn et al., 1972, Kaufman, 1977, Brock, 1979 and Wellington, 1982), as well as being an important component of the grazing reef fish community (Stephenson & Searles, 1960, Randall, 1965, Day, 1977, Ogden & Lobel, 1978, Bouchon-Navaro & Harmelin-Vivien, 1981, Hatcher, 1981, Miller, 1982 and Russ, 1984 a, b). The parrotfishes form a significant proportion of the food fishes caught in many tropical regions and include at least one highly prized species (Alcala & Luchavez, 1981 and Johannes, 1981).

Despite the recent increase in the number of studies on coral reef fishes (Ehrlich, 1975), parrotfishes have received relatively little attention, particularly in the Indo-Pacific region. One aspect that appears to have deterred many prospective investigators is that of identification. The identification of scarids in the field and in preserved collections is difficult owing to the lack of diagnostic morphological characters. Because of this, species

descriptions are frequently based on colour patterns. This has resulted in considerable confusion as colour patterns change with size, sex, reproductive status and a variety of environmental and behavioural factors. The colours also change during fixation. On the Great Barrier Reef, there are about 26 scarid species with a total of approximately 125 colour phases or patterns. Of these, seven are shared by at least two species.

Recent studies (including Randall & Ormond, 1978, Randall & Choat, 1980, Randall, 1983, Randall & Bruce, 1983 and Choat & Randall, pers. comm.) are contributing greatly to a more stable scarid nomenclature. Detailed morphological analyses have revealed useful characters that were previously overlooked. The initial and terminal phase colour patterns of most species have now been linked and the problems of synonomy, as a result of colour phases being identified as different species, have been largely resolved.

The aim of this study was to investigate the relationship between the functional morphology and behavioural ecology of parrotfishes. Parrotfishes are ideally suited for such comparative studies as they have highly specialized morphological characters and distinctive behavioural traits. Parrotfishes are also suitable study species in that they are diurnally active, strongly reef associated and numerous, and have a limited mobility.

This study includes analyses of both adults and juveniles. The biology of adults and juveniles are clearly not independent, although often different, and an investigation of the biology of a species must therefore include both stages, if a broad understanding of it is to be obtained. The biology of larval and juvenile

tropical marine fishes have received increasing attention in recent years (eg. Sale et al., 1980, Williams, 1980, Williams & Sale, 1981, Leis, 1982, Brothers et al., 1983, Doherty, 1983 and Shulman, 1984) but integrated studies of several life history stages are limited.

This thesis is presented in three main parts: morphology (Chapters 1 & 2), systematics (Chapters 3 & 4) and behavioural ecology (Chapters 5 & 6). Each part is further subdivided into adult and juvenile chapters. The descriptions in Chapters 1 and 2 are detailed, but unavoidably so, as they form a prerequisite for: a) the systematic considerations in Chapter 3 and b) the functional analyses in Chapters 1 and 2, which form the basis for designing and interpreting the ecological studies described in Chapters 5 and 6. The lack of published information on the taxonomy of juvenile scarids made the systematic analyses in Chapter 4 necessary, prior to the studies of juvenile scarids in Chapters 2 and 6.