# ResearchOnline@JCU

This file is part of the following reference:

# Herbert, Brett (2005) Feeding and growth of Golden perch (Macquaria ambigua), and assessment of its potential for aquaculture. PhD thesis, James Cook University.

Access to this file is available from:

http://eprints.jcu.edu.au/1332/

If you believe that this work constitutes a copyright infringement, please contact <u>ResearchOnline@jcu.edu.au</u> and quote <u>http://eprints.jcu.edu.au/1332/</u>



### FEEDING AND GROWTH OF GOLDEN PERCH (MACQUARIA AMBIGUA), AND ASSESSMENT OF ITS POTENTIAL FOR AQUACULTURE

Thesis submitted by Brett Herbert

For the Degree of Doctor of Philosophy in the School of Marine Biology and Aquaculture James Cook University September 2005

## ELECTRONIC COPY

I, the undersigned, the author of this work, declare that the electronic copy of this thesis provided to the James Cook University Library, is an accurate copy of the print thesis submitted, within the limits of the technology available.

Signature

Date

#### STATEMENT OF ACCESS

I, the undersigned, the author of this thesis, understand that James Cook University will make it available for use within the University Library and, by microfilm or other means, allow access to other users in other approved libraries. All users consulting this thesis will have to sign the following statement:

In consulting this thesis, I agree not to copy or closely paraphrase it in whole or in part without the written consent of the author; and to make proper public written acknowledgement for any assistance which I have obtained from it.

Beyond this, I do not wish to place any restriction on access to this thesis.

------

Brett Herbert

#### STATEMENT OF SOURCES

#### DECLARATION

I declare that this thesis is my own work and has not been submitted in any form for another degree or diploma at any university or other institution of tertiary education. Information derived from the published or unpublished work of others has been acknowledged in the text and a list of references is given.

-----

Brett Herbert

#### ACKNOWLEDGEMENTS

I acknowledge the support and assistance of my colleagues at the Freshwater Fisheries and Aquaculture Centre, Walkamin, for their assistance with sampling and running experiments. In particular, Peter Graham, Neil Harris, Ben Hockings, Dave Bull and Bevan O'Grady assisted with all aspects of day to day running and monitoring of experiments (particularly feeding and water quality monitoring), maintenance of equipment, assistance with measuring and grading fish, and support when things went wrong. I also acknowledge the assistance of DPI&F biometricians Scott Foster, Bob Mayer and Joanne de Faveri for guidance and advice on statistical analysis. Peter Graham is a co-author of the papers due to his dedication and input into experiments, and input into material prepared for publication separately to this thesis. I also thank the veterinary staff at Oonoonba veterinary laboratory for assistance and advice with disease issues, and for providing histological material for me to examine.

This work was conducted as an element of my employment by the Queensland Department of Primary Industries, which provided facilities, infrastructure and administrative support to enable this work to be conducted.

I wish to acknowledge the input of Chris Barlow, my initial co-supervisor who suggested that this study be undertaken. I also acknowledge the input of my supervisory team, Peter Appleford, Paul Southgate, and Rocky de Nys who assisted with commencement and finalisation of the thesis, and provided valuable comments and criticisms of the drafts. Stuart Rowland, Mike Rimmer, Jeff Gooley and Brett Ingram also provided input and suggestions on draft material for publications, and advised on experimental design and direction. Members of the Aquaculture Association of Queensland provided input into the direction of the research (for example focus on weaning fingerlings rather than larvae), and also advised me of requirements for further research when they started farming golden perch.

I also thank my family for their unfailing support during the course of this study.

#### Abstract

Golden perch (*Macquaria ambigua*) is a valuable freshwater fish native to south eastern Australia. The fishery for this species is diminishing and there exists an opportunity to develop aquaculture techniques for commercial production. Mass production techniques for fingerlings have been developed, but the paradigm that weaning of golden perch onto artificial foods essential for aquaculture development was difficult or impossible, impeded investigations into optimising aquaculture techniques for the species. The aims of this study were to develop aquaculture techniques for golden perch, focussing on three major issues: (1) weaning fingerlings on to artificial foods; (2) the nursery phase of production; and (3) growout of the fish to market size. Additional investigation of a destructive epizootic in aquaculture golden perch was also undertaken to develop control techniques for mixed motile aeromonad/ciliate protozoan infections.

Weaning of fingerling golden perch was investigated after discussions with industry indicated that reliable, cheap, mass production of fingerlings was regularly undertaken and there was no real need to wean larvae or fry. Fingerlings used in weaning experiments (18-31 mm TL, 0.1-0.5 g) are produced by commercial hatcheries for recreational fishery enhancement. A weaning technique using frozen *Artemia* nauplii and formulated crumble food ice blocks was developed, in which the frozen *Artemia* was gradually replaced by crumble food particles. *Artemia* nauplii slurry was replaced with crumble food particles, with the proportion of crumble increasing by ten percent each day until 100% crumble food was being fed to the fingerlings after ten days. Iceblocks of pure *Artemia* slurry, presented in a mesh bag, were fed over an acclimation period to habituate fish. Subsequently prepared co-feeding (*Artemia*/crumble) iceblocks were fed using the same method. This method was successful resulting in 77% weaning success.

Due to the relatively high cost of *Artemia*, frozen zooplankton was tested as an alternative and produced better weaning results than the control *Artemia* weaning treatment in terms of survival (16% better) and growth (1.57 g  $\pm$  0.55 g on zooplankton against 1.11  $\pm$  1.07 g on *Artemia*). This was adopted as the control treatment and the method of choice for mass weaning of fingerlings for nursery and

growout trials. After requests from industry for alternatives to plankton or *Artemia* in weaning fingerlings, commercially available seafood products were tested as co-feeding diets in the weaning process. Squid, fish roe, prawn, mussel, scallop, and fish were tested against a zooplankton control. The seafood materials were processed to be a similar size to the plankton control ( $500 \ \mu\text{m} - 1.5 \ \text{mm}$ ) and co-fed with crumble as iceblocks in 10% increasing increments and compared to a zooplankton control. Weaning using zooplankton as a co-feeding diet was far superior to other treatments tested in terms of both survival and growth. The best survival rates of the seafood treatments (the fish roe co-feeding treatment) was 39% and the poorest survival rate of zooplankton co-fed controls was 87% in the first trial. End weights of golden perch on all weaning treatments were significantly different (P<0.05) to the control. The olfactory and organoleptic properties of the co-feeding weaning diets were suggested to be of great importance in success of weaning.

Weaning techniques were further refined by investigating the effect of the duration of the transition period, the size of fingerlings, and light intensity on weaning success in fingerling golden perch. Attempts to reduce the co-feeding period or alter the ten-day transition period reduced weaning success significantly, from 62% feeding fish in the control treatment to between 36 and 46% for abbreviated treatments, suggesting that a minimum ten day transition period was essential for golden perch fingerlings. Mean weights at the end of the trial were also significantly different (P<0.05) ( $2.1 \pm 0.97$  g for controls, and  $1.6 \pm 0.83$  g to  $1.8 \pm 0.99$  g for treatments). The effect of size of fingerlings at weaning was tested (using fingerlings of the same age but different sizes), and no significant differences in weaning success were found between sizes from 0.1-0.24 g, suggesting that early weaning may provide significant benefits through a rapid transition to artificial diets.

The effect of light intensity on weaning of golden perch fingerlings was tested, with bright light, low light level and no light treatments. Golden perch weaned best in low light ( $1.79\pm0.081$  lux) and bright light ( $73.64\pm0.55$  lux), but over 6% of those in the dark treatment ( $0.00965\pm0.00275$  lux) were weaned successfully. The low light treatment produced significantly better results in terms of condition ( $3.45\pm0.01$  compared to the bright treatment ( $2.9\pm0.01$ ) (P<0.05), but was not significantly different in terms of survival or growth.

Two experiments were conducted on weaned golden perch to determine effects of density and diet on growth of golden perch in tanks. Firstly, in order to test the effect of density on growth of fingerlings, and to determine if density used in the weaning trials was suitable, golden perch fingerlings were grown in tanks at densities of 1000, 2000,7500 and 10000/m<sup>3</sup> for 82 days. At high stocking density there was less heterogeneity in growth than at low density, but overall growth was slower. Fish in the highest density treatment weighed significantly less ( $5.9 \pm 0.3$  g) than other groups ( $7.1\pm 0.3$  g to  $7.9 \pm 0.4$  g). Secondly, due to perceptions in industry that pellet texture was an impediment to golden perch feeding, a soft pellet was prepared using gelatine as a binding and moistening agent, and tested against the three commercially available dry pelleted feeds. Growth of golden perch feed on moist pellets ( $3.4 \pm 0.07$  g to  $2.6 \pm 0.07$  g) was significantly less than that of fish fed dry pellets ( $3.4 \pm 0.09$  g and  $3.83 \pm 0.09$  g).

After the issues in weaning golden perch on to artificial foods were resolved, trials to assess growth rates of golden perch in pond culture conditions were undertaken. The initial trial was conducted at two densities (105, 000 and 31, 250 fish/ha) for 220 days, with two replicates for each treatment. There were differences between the treatments in terms of growth (low densities  $96.86 \pm 9.62$  g and  $121.92 \pm 10.61$  g; and high densities  $83.75 \pm 10.01$  g and  $89.02 \pm 10.65$  g), but only the heaviest high density treatment was significantly different to the others. The size frequency distribution of high density treatment was skewed to the left (i.e. a high proportion of small fish) and bimodal, whereas in low density treatments it was more normal. To determine the reason for the skewed distribution the density experiment was repeated with greater replication (3 replicates of each treatment) and fish were sampled regularly to determine the role of diet in growth patterns. The results showed that a large proportion (67-70%) of fish reverted back to eating natural foods and that these were generally much smaller (mean weight about 10g) than those which retained pellet eating behaviour (mean weight around 80g). For every percentage point of pellet in the gut the weight was on average increased by 0.6423%. Analysis of natural diets determined that golden perch are more selective feeders than previously thought with smaller fish selecting *Moina* as prey over copepods, and larger fish feeding on chironomids or Trichoptera but not on Ephemeroptera or Odonata.

In order to test whether exposure to formulated food had a major influence on retention of weaned golden perch on pellets, a further experiment was run to test the effect of broadcast feeding. The results indicated that broadcast feeding significantly enhanced retention of fingerlings on pellets (42.5% retention in broadcast fed treatments against 25% in point feed treatments) and overall growth rates were therefore improved. Broadcast fed fish (15.639  $\pm$  1.07 g) were significantly larger than point fed fish (10.74  $\pm$  0.52 g and 10.899 $\pm$  1.14 g) at the end of a four month nursery period. In addition, a commercial probiotic product was concurrently trialed to determine whether probiotics had positive effects on water quality or health of fish. The results were too variable to permit meaningful analysis, due to the inherent variability of pond based production systems.

Growout of golden perch to market size after nursery phase was also conducted. To determine whether the smallest golden perch did have growth potential in a commercial setting, the entire contents of six ponds of fish were graded after nursery phase into the smallest 50% and the remainder. The different groups were then restocked into separate ponds. Ungraded controls (at the original density of 1265 fish/pond, approximately 4 fish  $/m^2$ ) were maintained as a control group. Sex ratios of the respective populations suggested that there was selective mortality of the fastest growing females due to grading (70% males in graded treatments compared to 62% males in the ungraded treatments). The majority of the small size class of fish did not reach market size in the six months after grading. Small fish started at  $6.2 \pm 0.4$  g finished at mean weight of  $107.6 \pm 10.83$  g, compared to large fish stocked at  $15.7 \pm$ 0.2 g which grew to a mean weight of  $235.1 \pm 20.56$  g. Ungraded fish averaged  $10.9 \pm$ 1.14 g at the start of the experiment and averaged  $165.7 \pm 22.43$  g at the end. Small fish did not appear to grow rapidly when separated from potentially dominant, larger fish, suggesting that factors other than behaviour influenced the size frequency distribution of golden perch cultured in ponds

Finally, when a mixed motile Aeromonad and hymenostome ciliate infection destroyed fish in early growout trials during this study, the aetiology and pathology of the disease was documented and an effective treatment devised. It was determined that *Tetrahymena corlissii* is a primary pathogen to naïve golden perch, and that motile aeromonad bacteria were probably secondary invaders. An effective treatment using a systemic protozooicide was administered which halted mortalities. Previously, *Tetrahymena* had not been reported in Australia, or in food fish, as a primary pathogen of fish in well managed ponds.

In summary, the results of this study indicate that golden perch can be weaned on to artificial foods and do have potential for aquaculture, although there are still issues regarding feeds and feeding (particularly retention of artificial foods), and handling of fish (and subsequent losses due to infections), which require further research for the industry to develop rapidly. However, their potential rapid growth, high market price, and tolerance of poor water quality engender them to profitable aquaculture production systems.

# **Table of Contents**

ABS	TRACT
List	of tablesx
List	of Figuresxv
СНА	PTER 1 1
INTF	RODUCTION AND LITERATURE REVIEW 1
1.1	Scope of Review1
1.2	Importance of golden perch1
1.3	Biology of golden perch
1.3	3.1 Distribution
1.3	B.2 Breeding
1.3	B.3 Feeding in fingerlings and juveniles
1.3	6.4 Growth rates
1.3	Ecology
1.4	Taxonomy and Genetics of Percichthyidae in Australia7
1.5	Golden perch in experimental and aquaculture situations
1.5	5.1 Induced breeding and fingerling production
1.5	5.2 Feeding of fingerlings in aquaculture ponds
1.5	5.3 Nutrition and feeding
1.5	5.4 Diseases
1.5	5.5 Physiology
1.6	Weaning of fish in aquaculture13
1.7	Size heterogeneity and grading of fish in aquaculture15
1.8	Aims of this Study16
СНА	PTER 2
DEV	ELOPMENT OF A WEANING PROTOCOL FOR GOLDEN PERCH FINGERLINGS 18
2.1	Introduction
2.1	.1 Development of weaning protocols for golden perch fingerlings
2.2	Materials and Methods
2.2	2.1 Experimental facilities
	2.2.1.1 Experiment 2.1. Weaning golden perch onto formulated diets using abrupt weaning,
;	and co-feeding with Artemia
	2.2.1.2 Experiments 2.2 and 2.3. Weaning golden perch using <i>Artemia</i> , zooplankton, and
-	commercially available seafood products
2.2	22 Diets and weaning regime
	2.2.2.1 Experiment 2.1. weaning golden perch onto formulated diets using abrupt weaning,
	and co-recently Artenna
	weaning fingerlings of golden perch.

2 a	2.2.2.3 Experiment 2.3. Weaning golden perch fingerlings using zooplankton and commerc vailable seafood products.	ally 25
<b>2.3</b> 2.3 co-1	<b>Results</b> 1 Experiment 2.1. Weaning golden perch onto formulated diets using abrupt weaning, ar feeding Artemia	<b>27</b> Id 27
2.3. wea 2.3. ava	<ul> <li>aning fingerlings of golden perch</li> <li>Experiment 2.3. Weaning golden perch fingerlings using zooplankton and commercial ilable seafood products.</li> </ul>	29 ly 30
2.4	Discussion	35
2.4. co-:	1 Experiment 2.1. Weaning golden perch onto formulated diets using abrupt weaning, a feeding Artemia.	nd 35
2.4. wea 2.4.	<ul> <li>Experiment 2.2. Comparison of Artenna, nozen zooplankton and formulated roods to</li> <li>fingerlings of golden perch.</li> <li>Experiment 2.3. Weaning golden perch fingerlings using zooplankton and commercia</li> </ul>	36 lly
ava	ilable seafood products	37
2.5	Conclusion	39
CHAI	PTER 3	41
FURT PERI 3.1	THER REFINEMENTS OF WEANING TECHNIQUES-EFFECTS OF TRANSITION OD, SIZE AT WEANING AND LIGHT INTENSITY ON WEANING SUCCESS	41 41
2.2		
3.2 3.2 3.2 3.2	<ol> <li>Experiment 3.1. Effect of transition period length on weaning of golden perch</li> <li>Experiment 3.2. Weaning of golden perch fingerlings of different sizes</li> <li>Experiment 3.3 Effect of Light on Weaning</li> </ol>	40 46 48 49
2.2	Decreter	12
3.3 3.3 3.3	<ol> <li>Experiment 3.1. Effect of transition period length on weaning of golden perch</li> <li>Experiment 3.2. Weaning of golden perch fingerlings of different sizes</li> <li>Experiment 3.3. Effect of Light On Weaning</li> </ol>	51 52 54
3.4	Discussion.	55
3.4. 3.4. 3.4.	<ol> <li>Experiment 3.1. Effect of transition period length on weaning of golden perch</li> <li>Experiment 3.2. Weaning of golden perch fingerlings of different sizes</li> <li>Experiment 3.3. Effect of Light on Weaning</li> </ol>	55 57 58
3.5	Conclusion	60
CHAI	PTER 4	61
GRO	WTH OF GOLDEN PERCH FINGERLINGS IN EXPERIMENTAL TANKS	61
4.1	General introduction	61
4.2	Materials and Methods	66
4.2. 4.2.	<ol> <li>Experiment 4.1. Effect of density on growth</li> <li>Experiment 4.2. Growth of golden perch fingerlings fed moist and dry formulated diet</li> </ol>	66 s. 68
4.3	Results	71
4.3.	A.1.1 Experiment 4.1. Effect of density on growth	71 71

4.3.1	.2 Growth	71
4.3.2	Experiment 4.2. Growth of golden perch fingerlings fed moist and dry formulated	diets.74
<b>4.4 D</b> i	scussion	77
4.4.1	Experiment 4.1. Effect of density on growth	
4.4.2	Experiment 4.2. Growth of golden perch fingerlings fed moist and dry formulated	l diets. 79
4.5 C	onclusion	
0		
CHAPTE	:R 5	83
POND N	URSERY TRIALS-EFFECTS OF DENSITY AND FEEDING	83
5.1 G	eneral introduction	
5.2 M	aterials and Methods	87
5.2.1	Pond and cage facilities	
5.2.2	Data collection techniques	
5.2.3	Measurements and handling of fish	
5.2.4	Water Quality	90
5.2.5	Experiment 5.1. First Pond Nursery and Growout Trial	90
5.2.6	Experiment 5.2. Nursery production of golden perch at two densities in ponds	92
5.2.7	Experiment 5.3. Effect of broadcast feeding and probiotics on growth of golden p	erch in
nursery	y. 94	
5.3 R	esults	
531	Experiment 5.1 First Pond Nursery and Growout Trial	96
531	1 Growth	96
531	2 Survival	102
531	3 FCR	102
531	4 Behaviour	102
531	5 Water Quality	102
5.3.2	Experiment 5.2. Nursery production of golden perch at two densities in ponds.	102
5.3.2	C1 Growth at different densities.	102
5.3.2	2 Reversion to natural production	105
5.3.2	Growth of fish on formulated or natural food	
5.3.2	.4 Water quality	
5.3.3	Experiment 5.3. Effect of broadcast feeding and probiotics on growth of golden p	erch in
nursery	. IIU 1 Dread aget/maint faceding	110
5.5.5	2 Water Quality/Drohiotica	110
3.3.3	5.2 water Quality/Problotics	
54 Di	scussion	115
541	Experiment 5.1 First Pond Nursery and Growout Trial	115
542	Experiment 5.1. First Fond Adjustry and Glowout That two densities in ponds	118
5.4.3	Experiment 5.3. Effect of broadcast feeding and probiotics on growth of golden p	erch in
nursery	v. 123	
5.4.3	.1 Broadcast/Point Feeding.	
5.4.3	2.2 Probiotics	
5.5 C	onclusion	
CHAPTE	:К б	130
	UT OF GRADED AND UNGRADED GOLDEN PERCH, AND SEXUAL	400
		130
6.1 In	troduction	

6.2	Materials and Methods	
6.3	Results	
6.3	.1 Growth	
6.3	.2 Survival	
6.3	.3 Sex ratio	141
6.4	Discussion	145
6.5	Conclusion	148
СНА	PTER 7	
TETF SEPT	RAHYMENOSIS, COLUMNARIS DISEASE AND MOTILE AERON TICAEMIA IN GOLDEN PERCH, FROM AUSTRALIA	10NAD 149
7.1	Introduction	149
7.2	Methods and Materials	
7.2	.1 Grow out environment and subject clinical history	
7.2	.2 Gross pathology, histology and microbiology	
7.2	Pond treatments and chemotherapeutic tests	
7.3	Results	
7.3	.1 Growout mortality and clinical history	
7.3	.2 Gross pathology, histology and microbiology	
7.3	.3	
7.3	.4 Pond treatments and chemotherapeutic tests	
7.4	Discussion	
СНА	PTER 8	168
GEN	ERAL CONCLUSION	168
LITE	RATURE CITED	174
APPI	ENDICES	202
Appe	endix 1 Animal Ethics Certifications.	
Appe	endix 2 Publications arising from this thesis	

## List of tables

Table 1.1. Golden perch production figures in Australia. Tonnage sold is exclusively wild caught fish, except for 2003/2004 which is aquaculture product	2
Table 1.2. Growth of golden perch in rivers and impoundments in NSW. Total length in mm unless indicated otherwise. Weight measurements where calculated are in parentheses.	6
Table 1.3. Diseases and parasites of golden perch	11
Table 2.1. Feeding schedule for golden perch weaning Experiment 2.1	23
Table 2.2. Analysis of diets used in weaning trials	23
Table 2·3. Mean ( $\pm$ SE) weight and length of golden perch in different weaning treatments. The same superscript indicates no significant difference between means in the same column (LSD P>0.05). Data are means of treatments	28
Table 2·4. Mean ( $\pm$ SE) weight, survival and weaning success of golden perch fed different weaning diets. Data are means of four replicates. Means with the same superscript are not significantly different (P<0.001). Mean starting weight was 0.16 $\pm$ 0.0097 g	29
Table 2.5. Mean ( $\pm$ SE) percentage survival and final weights of juvenile golden perch weaned using alternative fresh foods to zooplankton in Trial 1 and Trial 2. Different superscripts indicate significant differences between means (P<0.05).	32
Table 2.6. Predicted proportions of survivors and results of pairwise	
indicate significant differences between means (P<0.001)	33

Table 2.7. The analysis of standard length and weight showed a significant $1 + 2$	
weaning diet effect on growth. Different superscripts indicate significant	24
differences within columns between means at the indicated P levels	34
Table 3.1. Effects of light intensity on juvenile fish of various species	45
Table 3.2. Experimental layout of abbreviated weaning trials. Numerical	
values are percentage of crumble diet mixed with zooplankton for weaning	47
Table 3.3. Mean (± SE) light intensity readings for treatments. Means of light	
intensity in lux from 06:00 to 17:59 (day) and 18:00 to 05:59 (night)	50
Table 3.4. Mean (± SE) survival, final weight and percentage of non-feeding	
fish at end of experiment. Means with the same superscript were not	
significantly different at the 0.05 level	51
Table 3.5. Mean ( $\pm$ SE) start size, survival, SGR and final size of golden	
perch weaned at different sizes	53
Table 3.6. Mean ( $\pm$ SE) final weight, length, survival, condition index and	
starvation rates of golden perch weaned at three different light intensities; dark	
(0.00965 lux), ambient (1.79 lux) and light (73.64 lux)	55
Table 4.1. Effect of density on selected fish species	63
Table 4.2. Proximate analysis and indicator of rancidity (peroxide value) in	
moist and dry diets. Dry diets fed to fish were oven treated. All measures are	
on a dry matter basis. K = Kinta, R = Ridley's and P = Pivot	70
Table 4.3 Regression tests for density effects in golden perch density trials	
from the final measurements	71

Table 4·4. Mean ( $\pm$  SE) weight, CV of weight, survival, condition and SGR

of juvenile golden perch cultured for 82 days at four different densities. The	
same superscript indicates no significant difference (P<0.05)	73
Table 4.5. Mean ( $\pm$ SE) weights of golden perch throughout the experimental	
period. The same superscripts indicate no significant difference (P<0.001,	
except at day 98 P<0.007) between diet formulations (K.P.R) or texture	
(moist/dry) and interactions	75
Table 4.6 Survival of fish fed moist and dry diets after 98 days. The same	
superscripts indicate no significant difference in survival between treatments	77
Table 5.1. Experimental work on silver perch at different densities. Most	
experiments indicated small differences in growth between the densities	
trialed	85
Table 5.2. Food conversion ratio, specific growth rate (as $\%$ increase per	
day), survival, mean (±SE) final weight and weight gain up to day 221 for	
golden perch at two densities in ponds	97
Table 5.3 Size frequencies of golden perch from ponds at harvest	100
Table 5.4. Differences of size classes between high and low density	
treatments. Values are percentages, and are the mean of three replicate ponds.	
LSDs are derived from the raw data. Also presented are mean FCR, SGR and	
percentage survival of golden perch in nursery phase	103
Table 5.5 Percentage of golden perch eating pellets (pellets comprised >25%	
of stomach contents) on each sampling occasion	106
Table 5.6. Mean percentage of fish sampled with over 25% of gut contents	
being pellets (P), Chironomids (C), Moina (M), Ostracods.(O), or Trichoptera	
(T). Mean is of all sampling events	106

Table 5.7. Mean ( $\pm$ SE) weights of fish in three treatments at each sampling

period. Superscripts indicate significant difference at P< $0.05$ . LSD at 5% =	
1.727 except when comparing within the same treatment, then LSD $5\%$ =	
1.456	111
Table 5.8. Mean ( $\pm$ SE) pH levels in probiotic and non probiotic ponds	114
Table 6.2 Weights for golden perch for wild caught golden perch (Battaglene,1991)	132
Table 6.3 Mean ( $\pm$ SE) weights (g) and history (feeding treatment in nursery), and number of replicate ponds of golden perch fingerlings in this trial	133
Table 6.4. Percentages of size classes at end of trial. Combined Small plus large $(S+L)$ is small and large graded fish combined for comparison with ratios of ungraded fish	135
Table 6.5. Weight and length of fish (means $\pm$ SE, standard deviation and	
coefficient of variation) of at the end of the grading experiment. Different superscripts indicate significant differences at P< 0.001	137
Table 6.6. Survival of fish in ponds. All mortalities occurred immediately after grading. Mortalities due to operator error (B2) and tetrahymenosis (B6) are not included. Sg = small graded, Ug = ungraded and Lg = large graded treatments	140
	140
Table 6.7. Means (±SE) of variables measured in males and females in graded and ungraded treatments of pond grown golden perch	141
Table 6.8. The effect of prior treatment history on mean ( $\pm$ SE) growth of golden perch. The sex ratio is number of males per female	143
Table 6.9. Predicted weights of golden perch of Battaglene (1991)(B) and the current study (cs).	146

Table 7.10. Mean (± SE) weights for group A	151
fish	
Table 7.11. Mean (±SE) weights and size classes of group B	152
fish	
Table 7.12. Chemotherapeutic trials. As all fish had severe lesions, not all	
survived for the full length of the trial. At least two fish from every trial were	
alive at the end	154

Table 7.13 Morphometric characterization of <i>Tetrahymena corlissi</i> ( $X = mean$ ,	
SE = standard error, min = minimum, max = maximum, n = number of	
observations). Provided by Peter O'Donoghue, University of Queensland. All	
length measurements in µm	161

# List of Figures

Figure 3.1. Mean weight ( $\pm$ SE) of golden perch in abbreviated weaning	
trials	52
Figure 3.2. Coefficient of variation of golden perch weaned at different sizes (large, medium and small). The CV of large fish is lower than that of the small fish.	54
Figure 4·1 Regressions for density against mean weight, condition and percentage of 'feeding fish' for each replicate treatment. Each data point represents the values of a single replicate for fish measured on day 52	72
Figure 4·2. Size frequency distributions of golden perch fingerlings cultured for 82 days at four different densities (750, 500, 200 and 100 per 100 L tank)	73
Figure 4.3. Coefficient of variation (%) and growth rate (mg/day) of golden perch cultured for 82 days at four different densities. Error bars represent SE between replicates.	74
Figure 4·4. Mean (± SE) weights of fish fed dry diets (solid lines) or moist diets (broken lines)	76
Figure 5.1. Mean ( $\pm$ SE) growth of golden perch in ponds at two densities, low density (105,000 fish/ha) or high density (31,250 fish/ha)	97
Figure $5.2$ Specific growth rate and mean daily temperatures over the trial	98
Figure 5.3. Size frequency distributions of golden perch at two different densities (B1 and B3 low density, B2 and B4 high density)	99
Figure 5.4 Size frequency distributions during the last three months of the	

initial growout trial. Proportions of larger fish increased immediately prior to

harvest	101
Figure 5.5. Growth of golden perch at two densities. Mean (± SE) weight (g)	104
Figure 5.6 Mean ( $\pm$ SE) size frequency of golden perch, stocked at two different levels (High = 95,300 fish/ha; Low = 32,800 fish/ha) at the end of a 126 day pond-based nursery trial	105
Figure 5.7. Change in mean ( $\pm$ S.E.) monthly weight of golden perch stocked at two densities (High = 95,300 fish/ha; Low 32,800 fish/ha) in pond growout.	108
Figure 5.8. Scatter plots and trend lines of size of golden perch dependent on food type eaten (pellet or natural food)	109
Figure 5.9. Mean, maximum and minimum temperatures from hourly logging data	110
Figure 5.10. Mean weight ( $\pm$ SE) of golden perch in three treatments (broadcast fed and probiotics applied; point fed without probiotics, and point fed with probiotics applied) at each sampling period over three month nursery phase.	111
Figure $5 \cdot 11$ Size frequency distribution of fish feeding on pellets or natural foods at the end of the experiment	113
Figure 5.12. Size frequency distributions of fish at harvest after three months of nursery. All size classes are in grams	114
Figure 5.13. Growth of golden perch in different nursery trials. No account is made for differences in seasonal influences due to starting times. The 1999 trials were fed different diets to the others. Experiments in 1999 are	107
experiment 3.1, 2001 is experiment 3.2 and 2002 is experiment 3.3	12/

Figure 5.14. Growth of golden perch accounting for season of stocking. The 1999 trials were fed different diets to the others. Experiments in 1999 are experiment 5.1, 2001 is experiment 5.2 and 2002 is experiment 5.3	129
Figure 6.2 Mean ( $\pm$ SE) weight of graded and ungraded golden perch. Large ungraded and small ungraded are subsamples of the samples from ungraded ponds. Large graded and small graded are the respective graded fish	138
Figure 6.3 Mean ( $\pm$ SE) specific growth rate (percentage growth per day) of graded and ungraded golden perch over 35 weeks from April to December 2002. Means are of all fish sampled	139
Figure 6.3. Fitted (lines) and observed relationship of weight against standard length for male (green) and female (red) golden perch from growout trials. Total $n = 493$ male, 255 female	142
Figure 6.4. Size frequency histogram of golden perch - expressed as a percentage of each sex in each size class	143
Figure 6.5. Size frequency distribution of large, graded golden perch after nine months growout. Expressed as percentage of total population	144
Figure 6.6. Size frequency histogram of small, graded male and female golden perch after nine months growout. Expressed as percentage of total population.	144
Figure 6.7. Size frequency histogram of ungraded male and female golden perch after nine months growout. Expressed as percentage of total population	145
Figure 7.4 Mortalities caused by Tetrahymena in a typical pond, and temperature, in the 2001 epizootic (Group B)	156

Figure 7.5 Mortalities in the single pond affected in 2002 (Group C)	157
Figure 7.3. Location of lesion on the side of the fish	158
Figure 7.4. Early lesion on side of a golden perch. Note deep lesion into	
skeletal muscle and early haemorrhage around margins. There appears to be	
almost no immune response (oedema, erythremia) at this point	159
Figure 7.5. Early lesion with haemorrhagic margin on one side. Scales in	
middle of lesion are lifting	159
Figure 7.7. Advanced lesion. Haemorrhagic margin of lesion with skeletal	
muscle necrosis and secondary infection with fungi and bacteria. Extremely	
high numbers of Tetrahymena were found at the margins of such lesions, and	
fewer throughout the necrotic and dead tissue	160