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OSTEOPROTEGERIN:
A PATHOLOGICAL ROLE IN HUMAN ABDOMINAL AORTIC
ANEURYSM

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Corey Stephen Moran

October 2006

Declaration on Ethics

The research presented and reported in this thesis was conducted within the guidelines for research ethics outlined in the *National Statement on Ethics Conduct in Research Involving Human* (1999), the *Joint NHMRC/AVCC Statement and Guidelines on Research Practice* (1997), the *James Cook University Policy on Experimentation Ethics. Standard Practices and Guidelines* (2001), and the *James Cook University Statement and Guidelines on Research Practice* (2001). The proposed research methodology received clearance from the James Cook University Experimentation Ethics Review Committee (approval numbers H1464 and A964)

Corey Moran

October 16, 2006

(Date)

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Communications & Awards

1. CONFERENCES

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Moran CS, McCann M, Karan M, Quigley F, Viridi I, Lam AKY, Ketheesan N, Golledge J. *Osteoprotegerin: A key cytokine in abdominal aortic aneurysm.* XIIIth International Vascular Biology Meeting (IVBM); Toronto, Canada; June 2004. (Poster presentation)

Moran CS, Karan M, Quigley F, Ketheesan N, Golledge J. *Therapeutic relevance of osteoprotegerin (OPG) in human abdominal aortic aneurysm (AAA).* XIth meeting of the Australian Vascular Biology Society (AVBS); Barossa Valley, South Australia; September 2004. (Oral presentation)

Moran CS, Golledge J. *Interaction between ang-II, OPG, and PPAR γ in human AAA: A therapeutic pathway?* XIIIth meeting of the Australian Vascular Biology Society (AVBS); Gold Coast, Queensland; September 2006. (Poster presentation)

2. PAPERS

Moran CS, McCann M, Karan M, Norman P, Ketheesan N, Golledge J. Association of osteoprotegerin with human abdominal aortic aneurysm progression. *Circulation* 2005;111:3119-3125

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3. AWARDS

- Chemicon Prize in Biology and Genetics, Poster presentation, 3rd JCU Natural Sciences Festival, 2006
- John Shaw Biomedical Postdoctoral Fellowship, National Heart Foundation, Australia (2005).
- Doctoral Merit Research Prize, Faculty of Medicine, JCU (2004).
- Student Award, Oral presentation, AVBS Meeting, Barossa Valley (2004).

Abstract

Rupture of Abdominal Aortic Aneurysm (AAA) is the end-stage, catastrophic failure of the aneurysmal aortic wall and is associated with a mortality rate of up to 95 percent. Presently, surgery is the only treatment option available but carries with it a mortality rate of up to five percent and is usually reserved for repair of aneurysms showing high probability of rupture. What is required for the treatment of AAA, and essentially the basis of research in this area, is to understand the pathology of the disease well enough so that non-surgical intervention aimed at inhibiting small aneurysm progression can be developed.

The lack of non-invasive medical treatment for the disease, especially at the initial stages of development, stems from an incomplete understanding of its pathogenesis. Despite extensive laboratory and clinical research, the precise mechanisms leading to aneurysm formation remain unclear. The hallmark features of an aneurysmal aortic wall are degradation and fragmentation of the medial extracellular matrix (ECM), and significant reduction in smooth muscle cell (SMC) density, believed to be associated with the marked cellular inflammatory response also observed in the aneurysmal tissue.

A newly identified member of the tumour necrosis factor receptor superfamily known as osteoprotegerin (OPG) is constitutively expressed within the human artery wall and, under pathological conditions, is upregulated and associated with vascular disease. Elaboration on the involvement OPG of in AAA will determine its potential as a pharmacological target for the treatment of aneurysmal disease.

The focus of this study was to understand whether OPG might be important in the development of AAA. Two hypotheses were proposed:

1. Expression of OPG is upregulated in the aneurysmal aorta
2. Osteoprotegerin actively promotes aneurysm phenotype within the aortic wall

The specific aims of the study were to:

- a) Assess relationship between aortic concentration of OPG and the presence of aneurysm
- b) Define possible mechanism(s) by which OPG may be functionally active in the promotion of aneurysm development
- c) Modulate aortic expression of OPG and assess the effect on aneurysm development

Serum OPG was correlated with aneurysm growth rate in 146 men with small AAA followed by ultrasound for 3 years ($R=0.20$; $P=0.04$), and a demonstrated predictor of aneurysm expansion on multiple-regression analysis ($P=0.02$; coefficient 1.33, SE 0.51) in a model consisting of patient age, diabetic status, smoking history, initial aortic diameter, serum cholesterol, and C-reactive protein. Western analysis showed 3-fold, 8-fold, and 12-fold greater OPG concentrations in human AAA biopsies compared to age and gender-matched atherosclerotic narrowed aorta (AOD; 1.4 ± 0.1 ng/mg tissue vs 0.5 ± 0.1 ng/mg tissue; $P=0.002$), post-mortem non-diseased abdominal aorta (PAA; 1.4 ± 0.1 ng/mg tissue vs 0.2 ± 0.1 ng/mg tissue; $P<0.001$), and non-diseased thoracic aorta (TA; 1.4 ± 0.1 ng/mg tissue vs 0.1 ± 0.06 ng/mg tissue; $P<0.001$), respectively. Resident vascular smooth muscle cells (VSMC) and infiltrating macrophages were identified as primary sources for OPG within the aneurysmal aortic media. The association between aortic expression of OPG and the presence of AAA was confirmed in an animal model of experimental aneurysm formation, in which levels of OPG protein were 4-fold greater in aneurysmal aortic tissue compared to non-aneurysmal tissue. Furthermore, aortic tissue levels of OPG in this model correlated strongly with vessel diameter.

Healthy human aortic VSMC incubated with recombinant human OPG (0-20 ng rhOPG/ 10^5 cells/ml/24h) developed an aneurysmal phenotype defined by dose-dependent impaired cell proliferation ($P<0.001$), increased apoptosis ($P<0.01$), decreased interleukin (IL)-6 expression ($P<0.001$), and increased matrix metalloproteinase (MMP)-9 activity ($P=0.01$). Gene expression in OPG-treated

VSMC reflected these results exhibiting downregulation of genes associated with cell growth and survival, and upregulation of genes that negatively regulate cell growth and promote cell death.

Incubation of human monocytic cells with OPG (0-20 ng rhOPG/10⁵ cells/ml/24h) resulted in up to a 2-fold dose-dependent increase in IL-6 production in lipopolysaccharide (LPS)-activated cells ($P=0.005$). In addition, OPG (1 ng/10⁵ cells/ml/24h) acted to induce a 2-fold increase in MMP-9 expression ($P<0.001$), with a 1.5-fold increase in MMP-2 production ($P=0.01$) in resting human monocytic cells.

Treatment of human AAA tissue in culture with the angiotensin II receptor blocker, Irbesartan, and the peroxisome proliferator-activated receptor gamma (PPAR γ) ligands, Pioglitazone and Rosiglitazone, inhibited OPG production by up to 50%, as well as reducing inflammatory cytokine, and proteolytic enzyme production. The effects produced by thiazolidinedione treatment on aneurysm tissue *ex vivo* were reproduced *in vivo*. Both aortic expression of OPG and MMP activity within aortic tissue from a mouse model of experimental aneurysm formation were down-regulated significantly with Pioglitazone medication.

This study demonstrates for the first time the association of OPG with AAA and identifies a possible key role for the protein in the promotion of an aneurysmal phenotype within the normal aortic wall. The ability of existing medication to limit this action potentially opens a therapeutic pathway through which to limit aneurysm expansion in humans by targeting arterial expression of OPG.

Contents

DECLARATION	i
STATEMENT OF ACCESS	i
DECLARATION ON ETHICS	ii
ACKNOWLEDGEMENTS	iii
COMMUNICATIONS & AWARDS	iv
ABSTRACT	vi
LIST OF TABLES	xiv
LIST OF FIGURES	xv
ABBREVIATIONS	xviii
CHAPTER 1 INTRODUCTION	1
CHAPTER 2 LITERATURE REVIEW	4
2.1 AORTIC STRUCTURE IN HEALTH	4
2.1.1 The Aortic Wall	4
2.1.2 Aortic Extracellular Matrix: Tunica Media	6
2.1.2.1 <i>Collagen</i>	6
2.1.2.2 <i>Elastin</i>	6
2.1.3 Aortic Response to Change: Vascular Remodeling	9
2.1.3.1 <i>Extracellular Proteolytic Systems</i>	10
(i) Plasminogen/Plasmin System	10
(ii) The Matrix Metalloproteinase (MMP) System	12
2.1.3.2 <i>Vascular Cell (SMC) Apoptosis</i>	15
2.1.3.3 <i>Vasculopathology of Ageing</i>	20
2.2 THE ANEURYSMAL AORTA	23
2.2.1 Clinical Background	23
2.2.1.1 <i>Definition</i>	23
2.2.1.2 <i>Prevalence</i>	23
2.2.1.3 <i>Risk Factors</i>	24
2.2.1.4 <i>Natural History</i>	25
2.2.1.5 <i>Management Options</i>	26
2.2.2 Pathogenesis of Abdominal Aortic Aneurysms	27
2.2.2.1 <i>Altered Matrix Biology</i>	28
(i) Elastin	29
(ii) Collagen	30
a. Collagen Synthesis	31
b. Collagen Metabolism	31
(iii) Other ECM Components	33
2.2.2.2 <i>Proteolytic Degradation of the Aortic Media</i>	34
(i) MMP-2	34
(ii) MMP-9	37
(iii) MMP-12	37
2.2.2.3 <i>Aberrant Remodeling and AAA</i>	39
2.2.2.4 <i>Inflammation and AAA</i>	42
(i) Proinflammatory Cytokines	42
(ii) Inflammatory-cell Recruitment	43
(iii) Angiotensin II	44
(iv) Hypoxia-induced Inflammation	45

2.2.2.5	<i>Smooth Muscle Cell Apoptosis in AAA</i>	45
2.3	OSTEOPROTEGERIN	47
2.3.1	Characterization	47
2.3.2	Gene Organization and Protein Structure	48
2.3.2.1	<i>The OPG Gene</i>	48
2.3.2.2	<i>The OPG Protein</i>	49
2.3.3	OPG and the Vascular System	54
2.3.3.1	<i>The Skeletal-Vascular Link</i>	54
2.3.3.2	<i>Association of OPG with Vascular Disease</i>	55
2.4	SUMMARY	58
 CHAPTER 3 GENERAL MATERIALS AND METHODS		 60
3.1	HUMAN TISSUE STUDIES	60
3.1.1	Preparation and Storage of Human Serum	60
3.1.2	Collection of Vascular Tissue	60
3.1.3	Preparation and Storage of Biopsies	61
3.1.4	Histology	61
(i)	Tissue and Slide Preparation	61
(ii)	Haematoxylin and Eosin Stain	62
(iii)	Immunohistochemistry	62
3.1.5	Tissue Protein Extraction and Quantification	63
3.1.6	Western Blot Analysis	63
(i)	Protein Separation and Transfer	64
(ii)	Protein Detection and Visualization	64
3.1.7	Gelatin Zymography	65
3.2	IN VITRO STUDIES	65
3.2.1	Cell Culture	65
(i)	Human Vascular Smooth Muscle Cells	65
(ii)	Monocytic THP-1 Cells	66
(iii)	Human Aortic Macrophages and Peripheral Blood Mononuclear Cells	66
(iv)	Cell Passaging	67
(v)	Cell Storage	67
(vi)	Trypan Blue Exclusion Test of Cell Viability	68
(vii)	Cell Culture Immunocytochemistry	69
3.2.2	Explant Culture	69
3.2.3	Enzyme-linked Immunoassay	70
3.2.4	Fluorescence-activated Cell Scanning	70
(i)	Cell Fixation	70
(ii)	Cell-surface Staining	71
(iii)	Intracellular Staining	71
3.2.5	Assessment of Cell Proliferation	72
3.2.6	Assessment of Cell Apoptosis	72
(i)	Plasma Membrane Asymmetry (Annexin V Labeling)	72
(ii)	DNA Fragmentation	72
3.2.7	Extraction of Cellular mRNA	73
(i)	Purification	73
(ii)	Assessment of Purification, Yield, and Stability	73
3.3	MOUSE MODEL OF AAA	74
3.3.1	Animals	74
3.3.2	Aneurysm Formation	74
3.3.3	Assessment of Aneurysm Development	75

3.4	STATISTICS	75
3.4.1	Human Studies	75
3.4.2	Animal Model	76

CHAPTER 4 FEASIBILITY STUDIES AND PROTOCOL

	OPTIMIZATIONS	77
4.1	INTRODUCTION	77
4.2	R&D DuoSet® ELISA for OPG and IL-6	77
4.2.1	Study Design	77
4.2.2	Results and Conclusion	78
4.3	ANTIGEN EPITOPE RETRIEVAL FOR OPG IMMUNOSTAINING	79
4.3.1	Study Design	79
4.3.2	Results and Conclusion	80
4.4	WESTERN BLOT ANALYSIS FOR OPG	81
4.4.1	Study Design	81
4.4.2	Results and Conclusion	82
4.5	ISOLATION AND CULTURE OF VSMC	83
4.5.1	Study Design	84
4.5.2	Results and Conclusion	84
4.6	MACROPHAGE ISOLATION FROM AAA TISSUE	86
4.6.1	Study Design	86
4.6.2	Results and Conclusion	86
4.7	DETERMINATION OF VSMC PROLIFERATION	87
4.7.1	Aim of Study	87
4.7.2	Results and Conclusion	88
4.8	LPS-ACTIVATION OF THP-1 CELLS	89
4.8.1	Study Design	89
4.8.2	Results and Conclusion	90
4.9	EXTRACTION OF VSMC RNA	90
4.9.1	Study Design	91
4.9.2	Results and Conclusion	91
4.10	EXPLANT CULTURE OF HUMAN AAA TISSUE	92
4.10.1	Study Design	92
4.10.2	Results and Conclusion	92
4.11	ANIMAL MODEL FOR AAA	93
4.11.1	Study Design	93
4.11.2	Results and Conclusion	94
4.12	DETERMINATION OF DOSE-RANGE OF rhOPG FOR <i>IN VITRO</i> STUDIES	95
4.12.1	Study Design	96
4.12.2	Results and Conclusion	96

CHAPTER 5 OSTEOPROTEGERIN AND THE PRESENCE OF AORTIC ANEURYSM

5.1	INTRODUCTION	98
5.2	EXPERIMENTAL METHODS	99
5.2.1	Relationship between serum levels of OPG and AAA	99
5.2.2	Comparison of OPG levels in aneurysmal versus non-aneurysmal aortic tissue	99

5.2.3	Secretion of OPG by vascular and inflammatory cells within the aneurysm wall	99
5.2.4	Expression of aortic OPG in experimental AAA	100
5.3	RESULTS	100
5.3.1	Serum levels of OPG are weakly with aneurysm growth rate	100
5.3.2	OPG is upregulated in human aneurysmal aorta compared with non-aneurysmal aorta	102
5.3.3	OPG is secreted at high levels by medial smooth muscle cells and inflammatory cells within the human aortic aneurysm wall	104
5.3.4	OPG concentration is higher in aneurysmal aorta compared to non-aneurysmal aorta and correlates with aortic diameter in a mouse model of AAA.	107
5.4.	DISCUSSION	108
 CHAPTER 6 BIOLOGICAL ACTION OF OPG IN AAA		
PATHOGENESIS		110
6.1	INTRODUCTION	110
6.2	EXPERIMENTAL METHODS	111
6.2.1	OPG and proliferation of normal human aortic VSMC	111
6.2.2	OPG and apoptosis in normal human aortic VSMC	111
6.2.3	Effect of OPG on IL-6 production and gelatinase activity in normal human aortic VSMC	112
6.2.4	Effect of OPG on IL-6 production and gelatinase activity in human Monocytic cells	112
6.3	RESULTS	112
6.3.1	Recombinant human OPG inhibits proliferation in normal human VSMC	112
6.3.2	Recombinant human OPG promotes apoptosis in normal human VSMC	113
6.3.3	Recombinant human OPG inhibits IL-6 production and augments MMP-9 activity in normal human VSMC	116
6.3.1.	Recombinant human OPG stimulates IL-6 production and MMP-9 activity in THP-1 cells	117
6.4.	DISCUSSION	120
 CHAPTER 7 MECHANISMS OF OPG-INDUCED ANEURYSMAL		
PHENOTYPE IN HUMAN ABDOMINAL AORTIC VSMC		122
7.1	INTRODUCTION	122
7.2	EXPERIMENTAL METHODS	123
7.2.1	Preparation of control and OPG-treated VSMC	123
7.2.2	Gene Expression	123
7.3	RESULTS	124
7.3.1	Yield and purity of VSMC mRNA	124
7.3.2	OPG regulates expression of genes governing VSMC growth and survival	126
7.4	DISCUSSION	128

CHAPTER 8 MODULATION OF OPG IN THE ANEURYSMAL AORTA	132
8.1 INTRODUCTION	132
8.2 EXPERIMENTAL METHODS	133
8.2.1 Effect of AT ₁ R blockade on OPG production in AAA tissue	133
8.2.2 Effect of PPAR γ activation on OPG production in AAA tissue	133
8.2.3 Effect of activation on aortic expression of PPAR γ in experimental AAA	134
8.2.4 Effect of PPAR γ activation on expression of aortic OPG in experimental AAA	134
8.3 RESULTS	135
8.3.1 AT ₁ R blockade suppresses OPG production in AAA tissue	135
8.3.2 PPAR γ activation downregulates OPG production in AAA tissue	138
8.3.3 Pioglitazone increases PPAR γ within the aorta of Angiotensin II-infused mice	141
8.3.4 PPAR γ -activator therapy <i>in vivo</i> decreases OPG expression in the experimental aneurysmal aorta	141
8.4 DISCUSSION	143
CHAPTER 9 GENERAL DISCUSSION	146
APPENDIX 1 REPRODUCIBILITY DATA OF DuoSet[®] OPG ELISA	160
APPENDIX 2 REGULATION OF GENE EXPRESSION IN HEALTHY HUMAN ABDOMINAL AORTIC VSMC BY OPG	163
APPENDIX 3 BUFFERS, GELS, AND SOLUTIONS	177
APPENDIX 4 ETHICS APPROVALS	181
BIBLIOGRAPHY	185

List of Tables

Table 2.1	MMP subclasses and their extracellular substrates	13
Table 2.2	Feature differences between Apoptosis and Oncosis (Necrosis)	16
Table 2.3	Risk factors for abdominal aortic aneurysms	24
Table 2.4	Differences in MMP expression/activity in AAA and AOD	41
Table 2.5a	Regulation of OPG production (upregulation)	52
Table 2.5b	Regulation of OPG production (downregulation)	53
Table 4.1	Spectrometric analysis of extracted THP-1 RNA	91
Table 4.2	Average quantity of OPG (ng) per milligram of AAA tissue	96
Table 4.3	Average number of VSMC per milligram AAA tissue	97
Table 7.1.	Total RNA yield from control and rhOPG-treated VSMC	124
Table 7.2	OPG-induced change in expression of genes associated with cell-cycle regulation and survival in healthy human aortic VSMC	126
Table 7.3	OPG-induced change in expression of genes associated with growth and extracellular matrix synthesis in healthy human aortic VSMC	127

List of Figures

Figure 2.1	Cross-section through the normal aortic wall	5
Figure 2.2	Lamellar Unit	8
Figure 2.3	Morphological sequence of apoptosis	17
Figure 2.4	Schematic illustrating apoptosis pathway	18
Figure 2.5	Schematic illustrating factors potentially involved in the pathogenesis of aortic aneurysmal disease	28
Figure 2.6	Structure and amino acid sequence motifs of the OPG protein	50
Figure 4.1	Assessment of reproducibility in the R&D DuoSet [®] OPG ELISA	78
Figure 4.2	Effect of antigen epitope retrieval on the immunodetection of OPG in human AAA tissue	80
Figure 4.3	Quantification of immunostain: epitope retrieval versus no epitope retrieval	81
Figure 4.4	Validation of protein extraction and western blot protocols	82
Figure 4.5	Comparison of polyclonal primary antibody versus monoclonal antibody for detection of OPG in human AAA tissue.	83
Figure 4.6	Ratio of CD68-positive cells to α -actin-positive cells from primary culture through to third passage	85
Figure 4.7	Immunohistochemical validation of VSMC isolated by enzymic extraction.	85
Figure 4.8	FACS detection of the macrophage-specific cell surface marker CD71	87
Figure 4.9	DNA synthesis in VSMC isolated from nondiseased and aneurysmal human aorta over 24 hours	88

Figure 4.10	Proliferation in NASMC exposed to increasing concentration of FBS	89
Figure 4.11	Secretion of IL-6 from LPS-stimulated THP-1 cells	90
Figure 4.12	Stability of ribosomal RNA following storage at -20°C for 14 days	91
Figure 4.13	Viability of tissue explants pre-culture and after six days incubation in the absence or presence of treatment	93
Figure 4.14	Suprarenal aortic aneurysms in angiotensin II-infused ApoE ^{-/-} mice	94
Figure 4.15	MMP-9 activity in aortic segments from angiotensin II-infused ApoE ^{-/-} mice and correlation with aortic diameter	95
Figure 5.1	Correlation between serum OPG concentration and aortic diameter and aneurysm growth rate	101
Figure 5.2	Localization and distribution of OPG in healthy human aorta, human aneurysmal aorta, and human occluded aorta	102
Figure 5.3	Over-expression of OPG in human AAA	103
Figure 5.4	OPG expression in human aortic medial VSMC	105
Figure 5.5	Comparison of cellular secretion of OPG	105
Figure 5.6	Detection of intracellular OPG in AAA-derived VSMC by FACS	106
Figure 5.7	Detection of intracellular OPG in AAA-derived macrophages by FACS	106
Figure 5.8	Tissue OPG associated with aneurysm formation in the aorta of angiotensin II-infused ApoE ^{-/-} mice and correlated with aortic diameter	107
Figure 6.1	Inhibition of proliferation in healthy human aortic VSMC by OPG	113
Figure 6.2	Apoptosis in healthy human aortic VSMC induced by rhOPG	114

Figure 6.3	DNA fragmentation induced in normal human aortic VSMC by rhOPG	115
Figure 6.4	Upregulation of MMP-9 and downregulation of IL-6 in normal human aortic VSMC by rhOPG	116
Figure 6.5	Effect of rhOPG-treatment on IL-6 production in resting and LPS-activated THP-1 cells	117
Figure 6.6	IL-6 production in LPS-stimulated THP-1 cells in the presence of rhOPG	118
Figure 6.7	Upregulation of MMP-9 and MMP-2 in LPS-stimulated THP-1 cells	119
Figure 7.1	Stability of extracted VSMC mRNA	125
Figure 7.2	Purity of extracted VSMC mRNA	125
Figure 8.1	Effect of AT ₁ R blocker Irbesartan on MMP-2, 9, and IL-6 production in human AAA tissue explants	136
Figure 8.2	Downregulation of OPG expression in AAA tissue explants by AT ₁ R blocker Irbesartan	137
Figure 8.3	Downregulation of MMP-9 and IL-6 in human AAA tissue explants by PPAR γ activation	139
Figure 8.4	Downregulation of OPG production in human AAA tissue explants by PPAR γ activation	140
Figure 8.5	Upregulation of aortic PPAR γ in angiotensin II-infused mice by pioglitazone	141
Figure 8.6	Effect of pioglitazone pre-treatment on aneurysm formation in the suprarenal aorta of angiotensin II-infused ApoE ^{-/-} mice	142
Figure 8.7	Downregulation of OPG and MMP-9 production within the suprarenal aorta of pioglitazone-treated angiotensin II-infused ApoE ^{-/-} mice	143
Figure 9.1	Postulated role of OPG in AAA pathogenesis and progression in humans.	154

Abbreviations

AAA	Abdominal Aortic Aneurysm
AAMϕ	Aortic Aneurysm-derived Macrophage(s)
AASMC	Aneurysm Aortic Smooth Muscle Cell(s)
ACE	Angiotensin Converting Enzyme
AOD	Aortic Occlusive Disease
ApoE^(-/-)	Apolipoprotein E Gene (homozygous deletion)
AT₁R	Angiotensin II Receptor Type 1
DMEM	Dulbecco's Modified Eagles Media
DNA	Deoxyribonucleic Acid
ECM	Extracellular Matrix
ELISA	Enzyme-linked Immunoassay
FACS	Fluorescence-activated Cell Scanning
FBS	Foetal Bovine Serum
IFN	Interferon
IL	Interleukin
IRA	Infra-renal Aorta
LPS	Lipopolysaccharide
MMP	Matrix Metalloproteinase(s)
Mϕ	Macrophage(s)
NASMC	Normal Aortic Smooth Muscle Cell(s)
OPG	Osteoprotegerin
PAA	Post-mortem non-diseased Abdominal Aorta
PBM	Peripheral Blood Monocyte(s)
PPAR	Peroxisome Proliferator-activated Receptor
RANK	Receptor Activator of NF κ B
RANKL (Rkl)	RANK Ligand
rhOPG	recombinant human Osteoprotegerin
RNA	Ribonucleic Acid
SEM	Standard Error of the Mean
SMC	Smooth Muscle Cell(s)
SRA	Supra-renal Aorta
TA	Thoracic Aorta
TNF	Tumour Necrosis Factor
VSMC	Vascular Smooth Muscle Cell(s)