# Climate change impedes scleractinian corals as primary reef ecosystem engineers

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Abstract. Coral reefs are among the most diverse and productive ecosystems on our planet. Scleractinian corals function as the primary reef ecosystem engineers, constructing the framework that serves as a habitat for all other coral reef-associated organisms. However, the coral's engineering role is particularly susceptible to global climate change. Ocean warming can cause extensive mass coral bleaching, which triggers dysfunction of major engineering processes. Sub-lethal bleaching results in the reduction of both primary productivity and coral calcification. This may lead to changes in the release of organic and inorganic products, thereby altering critical biogeochemical and recycling processes in reef ecosystems. Thermal stress-induced bleaching and subsequent coral mortality, along with ocean acidification, further lead to long-term shifts in benthic community structure, changes in topographic reef complexity, and the modification of reef functioning. Such shifts may cause negative feedback loops and further modification of coral-derived inorganic and organic products. This review emphasises the critical role of scleractinian corals as reef ecosystem engineers and highlights the control of corals over key reef ecosystem goods and services, including high biodiversity, coastal protection, fishing, and tourism. Thus, climate change by impeding coral ecosystem engineers will impair the ecosystem functioning of entire reefs.

Additional keywords: bleaching, ecosystem goods and services, ocean warming and acidification.

## Scleractinian corals as reef ecosystem engineers

Ecosystem engineers are organisms that modulate the availability of resources to other species by causing physical state changes in biotic or abiotic materials (Jones *et al.* 1994, 1997). Scleractinian corals act as key reef ecosystem engineers in two main ways: first, they are autogenic engineers, because through their calcification and ensuing reef accretion, they change the physical, chemical, and biological environment and thereby **provide habitats** for associated reef organisms. The generation

of complex, hard, and stable substrates by scleractinian corals underpins the high biodiversity characteristic of tropical coral-reef ecosystems (Bellwood and Hughes 2001).

Second, corals also act as allogenic ecosystem engineers because they **intensively generate and transform inorganic and organic materials**. Coral-derived inorganic calcareous skeletons are transformed into calcareous reef sands by reef-associated bio-eroding organisms (such as molluscs, echinoderms, and sponges) and by other biological, physical, and

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chemical erosion processes (Glynn 1997; Hallock 1997), so that corals, through the production of inorganic materials, control adjacent sediments. These highly permeable biogenic sediments with generally large grain sizes allow intense advective coupling between the water column and the seafloor (Huettel et al. 2003; Rasheed et al. 2003; Wild et al. 2005a). Calcareous reef sands support abundant associated heterotrophic microbes (Wild et al. 2006). Coral-generated calcareous sediments thereby act as biocatalytic filter systems that facilitate rapid processing and recycling of organic matter (Wild et al. 2004a, 2005a, 2005b, 2008). Corals also continuously release large amounts of dissolved and particulate organic materials, which may function as energy carriers and particle traps (Wild et al. 2004a; Huettel et al. 2006; Naumann et al. 2009). This release of organic material promotes the formation of mucus-particle aggregations in the water column that increase the sedimentation and recycling rates by which essential elements are retained (Wild et al. 2004b, 2005b; Huettel et al. 2006). Consequently, a wide range of biogeochemical processes, important in coral reef functioning, are directly controlled by scleractinian corals acting as the principal ecosystem engineers.

This review sets out to expand our understanding of the impacts of global climate change on coral reef ecosystems owing to the direct impediment of the reef's primary ecosystem engineer – the scleractinian corals. Rather than focusing on the plight of corals *per se*, we chose to explore how the impacts of ocean warming and acidification may affect the corals' ability to act as engineers within this complex ecosystem. We discuss the impact of climate change on scleractinian corals at the organism level and how this translates into responses at the reef ecosystem level.

# The impact of global climate change on the coral engineer

As stenothermic and calcifying organisms, corals are particularly sensitive to both ocean acidification and warming. There are several indications that carbon pollution-induced increases in ocean acidity and temperature are impacting the metabolism and growth of reef-building corals (Langdon and Atkinson 2005; De'ath *et al.* 2009; Tanzil *et al.* 2009; Manzello 2010).

# Ocean acidification and coral calcification

Increasing acidity of oceanic waters represents a direct threat to reef-building scleractinian corals, with various implications for their role as ecosystem engineers. Ocean acidification is the consequence of global oceanic uptake of increasing anthropogenic atmospheric CO<sub>2</sub> (e.g. Kleypas *et al.* 1999). Such uptake increases CO<sub>2</sub> partial pressure (pCO<sub>2</sub>) in the water column, decreases seawater pH, increases concentrations of total dissolved CO<sub>2</sub> ([CO<sub>2</sub>] and [HCO<sub>3</sub>]), and reduces concentrations of [CO<sub>3</sub><sup>2</sup>] in seawater (Caldeira and Wickett 2003; Feely *et al.* 2004). Physiological processes (e.g. calcification) in corals may respond to these changes in ocean chemistry (Langdon and Atkinson 2005).

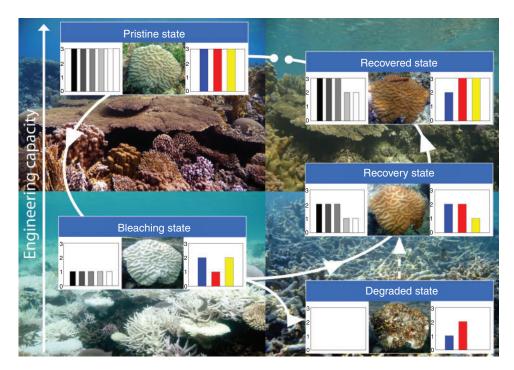
The reduction in  $[CO_3^2]$ , at constant seawater calcium concentration  $[Ca^{2+}]$ , consequently results in the decrease of the saturation state of aragonite ( $\Omega_{arag}$ ), the polymorph of  $CaCO_3$  produced by coral calcification. Presently, tropical

surface waters, with the exception of the eastern Pacific Ocean, are about 4-fold supersaturated with respect to aragonite (Hoegh-Guldberg et~al.~2007). However,  $\Omega_{arag}$  is expected to significantly decrease to levels of 2.5–3.0 by the year 2100 (Feely et~al.~2009). Scleractinian corals generally require seawater that is super-saturated in aragonite for efficient aragonite accretion. In acidified seawater, lowered external  $\Omega_{arag}$  impedes the essential increase of  $\Omega_{arag}$  within the internal calcifying fluid, and causes a corresponding decrease in calcification rate (reviewed in Cohen and Holcomb 2009). This decrease in skeletal growth performance, caused by ocean acidification, directly translates to a decline in the engineering capacity of scleractinian corals to construct essential reef habitats.

Various studies have documented the negative effect of ocean acidification and the consequential reduction in seawater  $\Omega_{arag}$  on coral calcification in both the laboratory (e.g. Anthony et al. 2008; Jokiel et al. 2008) and the field (Bak et al. 2009; De'ath et al. 2009; Tanzil et al. 2009). However, a very recent study (Jury et al., in press) presents significant differences in calcification rates at equal [CO<sub>3</sub><sup>2</sup>] and further suggests [HCO<sub>3</sub><sup>3</sup>] as a potentially more important driver for coral calcification, thereby questioning the reliability of  $\Omega_{arag}$  or  $[CO_3^{2-}]$  as sole predictors of the effect of ocean acidification on coral calcification. Another recent study argues that deleterious effects caused by elevated [CO2], as a result of ocean acidification, may be ameliorated by inorganic nutrient enrichment (Holcomb et al. 2010). These authors conclude that naturally elevated inorganic nutrient levels may thus support increased primary and secondary production, consequently facilitating coral calcification in environmenst with naturally high concentrations of CO2. However, species-specific differences in sensitivity to ocean acidification and thermal stress may occur (Manzello 2010). This could have tremendous effects on the structure of communities in future coral reefs (Loya et al. 2001).

Ocean acidification can also affect coral reproduction by reducing sperm motility (Morita et al. 2009) or settlement and post-settlement development of planula larvae and coral recruits (Albright et al. 2008; Cohen et al. 2009). At experimentally reduced aragonite saturation states ( $\Omega_{arag}$ ), the early skeleton of coral recruits showed progressive changes in aragonite crystal morphology and a decline in crystal growth rate (Cohen et al. 2009). This implies that ocean acidification may significantly affect recruitment rates and the competitive capacity of coral populations, and may consequently lead to a shift in coral community structure. In addition, a recent study showed that spawning female corals of the temperate species Astrangia poculata are more susceptible to the negative effects of ocean acidification than spawning male corals (Holcomb et al., in press). This gender discrimination may be a result of the energetically expensive egg production process, leaving only limited resources to compensate for the effects of acidification on calcification. On a longer time-scale, this lack of energy and growth may reduce recruitment success for gonochoricspawning coral species.

Finally, and possibly most alarmingly, ocean acidification has been identified as a potential trigger for coral bleaching (Anthony *et al.* 2008). According to this study, branching and massive coral species experience an increase in bleaching with decreasing seawater pH (8.4–7.6) at low (25–26°C) and high



**Fig. 1.** Relative potential of key physiological functions and resulting ecosystem engineering capacity of corals under pre-, during and post-bleaching conditions (foreground, organism level; background, ecosystem level). Left columns: physiological functions (black to white, coral–dinoflagellate symbiosis stability, primary productivity, calcification, reproductive output, immune defence). Right columns: coral-generated reef ecosystem engineering (blue, facilitation of associated biodiversity via habitat differentiation; red, generation of biocatalytical filter systems via inorganic sediment production; yellow, impact on particle trapping and conservation of essential elements via organic matter release). Scale: 0–3, none, low, medium, high. Coral photographs are modified from Manzello *et al.* (2007) whereas reef photographs are copyright of M. S. Naumann, except that of bleached reef (copyright: R. Berkelmans; www.reefbase.org).

(28–29°C) seasonal temperatures. These findings suggest that ocean acidification may be another primary bleaching-trigger, causing similar and/or other deleterious consequences for scleractinian corals as reef ecosystem engineers.

# Ocean warming and coral bleaching

Reef-building corals form a primary habitat for zooxanthellae (endosymbiotic dinoflagellates) of the genus Symbiodinium, which are crucial to the physiological integrity and ecological function of reef-building corals within tropical-reef systems. Coral-zooxanthellae interactions reflect a close evolutionary relationship between host and symbiont, with temporal and spatial variability allowing the coexistence of diverse symbionts within the coral host (Rowan et al. 1997). In the eastern Pacific Ocean, Pocillopora verrucosa colonies inhabiting shallow-water habitats contained different clades of zooxanthellae than Pavona gigantea colonies living in deeper water and more shaded habitats (Iglesias-Prieto et al. 2004). Similarly, Symbiodinium spp., within Acropora tenuis appear to be distributed according to light exposure, with two different types of Symbiodinium spp. found in sunny and shady parts of the colony, respectively (Van Oppen et al. 2001). Such tight interaction leads to the conclusion that the two organisms cannot be considered separately (Iglesias-Prieto et al. 2004).

One of the most dramatic impacts of ocean warming on coral reefs is mass coral bleaching, which is the breakdown of the symbiosis between corals and zooxanthellae. Bleaching is associated with a pronounced loss of colour from affected corals caused by the expulsion of the photopigment-rich zooxanthellae (Fig. 1). Mass coral bleaching is strongly associated with anomalously high sea-surface temperatures (Glynn 1993), but it can also be triggered by a range of other environmental stress factors (e.g. altered salinity, light, sedimentation, and toxins). Since the early 1980s, mass coral bleaching events have increased in extent and frequency in response to steadily warming ocean-water temperatures (Hoegh-Guldberg 1999).

Coral bleaching is a reversible process (Fig. 1) if the stressful conditions are relatively mild and short-term, with zooxanthellae populations inside the host tissues returning to pre-bleaching levels after 2–6 months depending on the coral species (Hoegh-Guldberg and Smith 1989; Brown 1997; Hoegh-Guldberg and Jones 1999). Bleached corals, however, are physiologically compromised because of the reduction in the abundance of zooxanthellae and/or their pigments. Under normal conditions, zooxanthellae translocate up to 95% of their photosynthetically fixed carbon to the coral host (Muscatine *et al.* 1984). They also cover 30% of the host's nitrogen requirements for growth, reproduction and maintenance (Bythell 1988) from dissolved nutrient uptake (Bythell 1990). During bleaching, the photosynthetic ability of reef-building corals is greatly reduced

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(Porter *et al.* 1989; Suzuki and Kawahata 2003; Rodrigues and Grottoli 2006), leading to the depletion of energy stores, which results in 'food shortage' and starvation (Porter *et al.* 1989; Fitt *et al.* 1993).

As calcification strongly depends on the photosynthetic efficiency of the zooxanthellae within the tissues of corals (Gattuso et al. 1999; Allemand et al. 2004; Colombo-Pallotta et al. 2010), bleached and/or thermally stressed corals have lower growth rates (Jokiel and Coles 1977; Leder et al. 1991) and tissue regeneration capacities (Meesters and Bak 1993). In Montastraea faveolata, one of the major reef-builders of the Caribbean, bleaching reduced photosynthetic efficiency by 4.6-fold and capacity by up to 4.8-fold (Rodríguez-Román et al. 2006). These changes in photosynthetic activity translate directly into a decline in calcification. Calcification rates of bleached M. annularis colonies were only 37% of the mean annual calcification observed before bleaching (Leder et al. 1991). Laboratory experiments revealed that calcification rates in Montastraea faveolata, immediately after bleaching, can be as low as 22% of unbleached control corals (F. Colombo-Pallotta, pers. comm.).

The physiological consequences of bleaching and the time required to return to normal conditions is species-specific. Many branching corals of the genus Acropora die within 10 to 14 days after bleaching, whereas some massive species may show greater resistance and recover after several months (Rodrigues and Grottoli 2006; Rodrigues et al. 2008). Early life stages of corals seem to be particularly vulnerable to climate change stressors (Edmunds 2007), thus affecting the demographics of coral reefs. Bleaching may impact the reproductive output and planula larvae development of corals (Baird and Marshall 2002) for up to several years (Ward et al. 2002). The observed reduction in reproductive output of scleractinian corals that survived bleaching indicates that recovery to former levels is slow (Baird and Marshall 2002), suggesting that sub-lethal bleaching leads to long-term effects on corals and associated ecosystem functioning. Re-establishment of reef corals after major disturbance events with mass coral mortality usually takes years to decades (e.g. Loya 1976, 2004; Done 1992; Loch et al. 2004).

#### The effect of light

For corals under thermal stress, ambient light conditions can significantly affect the onset and severity of bleaching events. Low or reduced irradiance during times of high water temperature reduces photoinhibition and suppresses coral bleaching (Iglesias-Prieto et al. 1992; Mumby et al. 2001; Brown et al. 2002; Takahashi et al. 2004). By contrast, reductions in cloudiness exacerbate thermal stress. Nonetheless, the response of clouds to climate change remains one of the largest sources of uncertainty in modelling climate at regional and local scales. Some studies show that warming oceans could reduce upper-level cloudiness, particularly in the tropics, because the higher temperatures increase turnover efficiency (Mumby et al. 2001; Lau and Wu 2003; Lau et al. 2005). A cause for concern is the modelled simulations of doubled atmospheric CO2 that predict reductions in total cloudiness, leading to greater absorption of shortwave and long-wave radiation by the oceans (Sud et al. 2008).

### Coral immunity and diseases

Several studies have correlated climate change, in general (Harvell *et al.* 2001), or temperature anomalies, in particular (Willis *et al.* 2004; Miller *et al.* 2006; Bruno *et al.* 2007; Muller *et al.* 2008), to the increased prevalence of coral disease, and disease incidence has been shown to increase following coral bleaching (Brandt and McManus 2009). Invertebrate immunity can be reduced under starvation conditions (Feder *et al.* 1997; Moret and Schmid-Hempel 2000; Seppälä *et al.* 2008) similar to those induced during coral bleaching. Compromised coral immunity during and following a bleaching event (Fig. 1) (Mydlarz *et al.* 2009; Reed *et al.* 2010) will therefore limit the activation and efficacy of a response to invading organisms or physical injury (Meszaros and Bigger 1999; Rinkevich 1999; Stedman 2000), leading to an increase in disease prevalence following coral bleaching (Harvell *et al.* 2001; Miller *et al.* 2006).

There is also growing consensus that microbes interact strongly with corals, forming an integral part of a symbiosis that includes coral, zooxanthellae, and associated microflora. This has been referred to as the 'holobiont' to connote a biological 'unit' that operates as one, within both ecological and evolutionary time frames (Rohwer et al. 2002). The concept of the coral holobiont emerged following several research advances using culture-based and culture-independent molecular techniques. Coral-associated bacterial communities show greater similarity between distantly-separated colonies of the same host species than between adjacent colonies of different host species (Rohwer et al. 2002), implying that bacterial communities are host-coral specific. Healthy and diseased corals show distinct differences in microbial community composition (Cooney et al. 2002; Frias-Lopez et al. 2002, 2004; Ritchie 2006), including the apparently healthy tissues remote from the disease lesion of infected corals (Pantos et al. 2003; Pantos and Bythell 2006). This indicates that in a changing climate, there will be pronounced changes in community composition and diversity of microbes associated with corals.

Such microbial associates may perform several different functions, including that of N-fixation (Williams *et al.* 1987; Shashar *et al.* 1994; Lesser *et al.* 2004), and as an antibiotic agent in the case of bacteria associated with coral mucus (Ritchie 2006). The experimental evidence on selective properties of antibacterial substances in the mucus of the coral *Acropora palmata* (Ritchie 2006) indicates a probiotic role of bacteria in corals (Knowlton and Rohwer 2003; Pantos *et al.* 2003; Reshef *et al.* 2006; Rypien *et al.* 2010), whereby the normal microbial flora inhibits the colonisation and growth of potential pathogens via competition and release of antimicrobial compounds (Ritchie 2006; Rypien *et al.* 2010).

#### Reef ecosystem-level responses

Habitat provisioning by corals and effects on associated organisms

Climate change has the capacity to change reef landscapes and benthic communities. Coral-generated production of framework structures will substantially decrease with climate change-induced bleaching and subsequent reduction of coral calcification. Such declines will influence associated organisms and significantly alter the reef's biogeochemical functioning (Fig. 1).

On a larger scale, impeding coral ecosystem engineers may cause phase shifts from scleractinian corals to other invertebrates such as gorgonians, soft corals, ascidians and sponges (Bak et al. 1996; Maliao et al. 2008) or even benthic algae (Hughes 1994; McCook 1999; Diaz-Pulido and McCook 2002; Bellwood et al. 2006) with obvious negative effects on provisioning of three-dimensional surfaces and habitats for associated high biodiversity. Climate change will contribute to such 'phase shifts' and potentially to long-term destruction of reef framework and changes in reef functioning (Glynn 1993; Aronson and Precht 1997; Diaz-Pulido and McCook 2002; Aronson et al. 2004). Dead corals are rapidly colonised by a diverse community of algae (Diaz-Pulido and McCook 2002), and living corals are directly overgrown or damaged by macroalgae (Smith et al. 2006; Haas et al. 2010). Algal colonisation of surfaces of dead or impaired corals therefore likely influences the recovery of neighbouring corals from climate changeinduced stress such as bleaching of the coral engineer. This may also include dramatic changes of biogeochemical processes such as the fixation of nitrogen via coral-associated cyanobacteria. On a reef-wide scale, within weeks of a major bleaching event and mass coral mortality, the production of new nitrogen can be 30-fold greater than that associated with living corals under undisturbed conditions (Davey et al. 2008). Thermallyinduced mortality of the coral engineer therefore has the potential to significantly alter the reef systems' overall dynamics and integrity. In addition, the loss of live tissue will affect natural recycling of nutrients within the system.

On a smaller scale, this involves changes in the scleractinian coral community. Branching, framework-building scleractinian corals including the genera *Acropora*, *Seriatopora*, *Pocillopora*, and *Stylopora* are examples of morphologies that are more sensitive to thermal stress than massive and encrusting growth forms (Marshall and Baird 2000; Loya *et al.* 2001; McClanahan *et al.* 2002). In addition, large colonies of these genera were more susceptible to thermal stress than small colonies (Loya *et al.* 2001; Mumby *et al.* 2001; Nakamura and van Woesik 2001; Bena and van Woesik 2004). Therefore, climate change will reduce the framework-building and habitat-generation capacity of reef-coral communities.

Calcifying activities of reef-building corals ultimately result in a three-dimensional matrix that provides space, shelter, and food for a multitude of organisms (Srinivasan 2003; Sale et al. 2005; Raes et al. 2007). High reef rugosity, for example where large massive Porites coral colonies dominate the seascape, is associated with high abundances of reef fishes (McClanahan and Shafir 1990). Similarly, a high abundance of microstructures within coral colonies is strongly and positively correlated with high fish and invertebrate diversity (Nanami and Nishihira 2004; Nanami et al. 2005). Reduction in rugosity reduces the availability of habitat space at a variety of scales and leads to a considerable reduction in reef biodiversity (McClanahan and Shafir 1990). In the Caribbean, the architectural complexity of reefs has already declined non-linearly over the last 40 years (Alvarez-Filip et al. 2009) as a combined result of climate-change effects and local disturbances.

The fate of reef corals and their community composition is not only fundamental for future reefs to maintain their ability to accrete and form coastal barriers, but also in determining whether or not they will continue to support the plethora of coral-associated diversity through time. Loss of corals and shifts in species composition reduces the variety of habitats available at macro- and micro-scales, which likely leads to subsequent losses of habitat complexity and associated biodiversity of invertebrates and fish (Reaka-Kudla 1997; Loya *et al.* 2001; Wilson *et al.* 2006). This includes obligate associates such as corallivorous fishes in the short term, but also subsequent reductions in overall fish diversity 3–4 years following a thermal stress event (Pratchett 2007; Munday *et al.* 2008).

#### Control of biocatalytical sand filters

The commonly observed phase-shifts from the dominance of corals to that of fleshy algae will additionally decrease the production of inorganic material in the long term, but will increase sediment-forming erosion processes in the short term, because of the breakdown of the reef framework (Fig. 1). The extent of this reef degradation is obviously dependent on the functional group of algae involved in phase shift. In areas dominated by crustose coralline algae, fragmentation of the reef framework can be mitigated by cementation and binding of sediments and loose coral fragments (Kennedy et al. 2002; Payri 1988). Some coralline algae species can even induce coral larval settlement, thereby facilitating new recruitment and reef recovery (Fabricius and De'ath 2001; Harrington et al. 2004; Golbuu and Richmond 2007). This beneficial effect may, however, depend on the extent of climate change, as crustose coralline algae are particularly sensitive to ocean warming and acidification (Anthony et al. 2008). Calcareous green and articulated corallines contribute less to sediment binding, although their sediment production, in particular by calcareous green algae (e.g. *Halimeda* spp.), can be high (Bach 1979; Drew 1983; Payri 1988).

Carbonate production by *Halimeda* spp. is likely quantitatively lower than that by scleractinian corals (Bosence 1989). In addition, sand and coralline sediments derived from *Halimeda* spp. are different in density and chemistry from coral-derived sediments (Borowitzka and Larkum 1977; Braga *et al.* 1996; Bosence and Wilson 2003). Therefore, these algae-generated sediments will very likely have shorter residence times, and therefore will most likely not function as biocatalytical filter systems as described for coral-generated sediments (Wild *et al.* 2005*a*, 2009; Werner *et al.* 2006).

### Generation and fate of coral-derived organic material

Under undisturbed conditions, scleractinian corals release particulate organic carbon (POC) in quantities of 0.3–7.1 mg POC m<sup>-2</sup> coral surface h<sup>-1</sup> (Wild *et al.* 2004*a*, 2005*b*; Naumann *et al.* 2010). To date, there are no available studies on the effect of acidification on coral-derived organic matter release. However, during thermal-induced bleaching, two different kinds of organic matter are released in increased quantities: (1) zoo-xanthellae, and (2) coral-derived particulate organic matter (POM) (Niggl *et al.* 2009). Degradation of these substrates by reef microbes differs considerably, with very low rates observed

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for symbiotic dinoflagellates (<1% h<sup>-1</sup>), and high rates for coral-derived POM (>5% h<sup>-1</sup>) (Wild *et al.* 2005*a*). This could indicate that a major fraction of the symbiotic dinoflagellate-composed POM does not remain in the reef system long enough to be recycled, and thus likely involves a considerable loss of essential nutrients. In contrast, the coral-derived POM may function as an energy carrier and particle trap (Wild *et al.* 2004*a*; Huettel *et al.* 2006). This material potentially stays within the system and is recycled, particularly by the benthic community. A short-term pulse of labile organic matter can result, and studies have demonstrated that this organic matter can be recycled by the planktonic and benthic reef community within two weeks (Wild *et al.* 2004*c*, 2008; Eyre *et al.* 2008; Glud *et al.* 2008).

Histological analyses (Fitt *et al.* 2009), however, indicate that internal mucus reservoirs in the coral tissue are depleted during bleaching, so that mucus–POM release by corals is most likely only stimulated during the early phase of bleaching, dropping to low levels the longer the bleaching event lasts (Fig. 1). This dynamic flux of POM was confirmed by Piggot *et al.* (2009), who demonstrated that the number of mucusproducing cells (mucocytes) in the coral tissue increases with increasing sea-surface temperatures, but declines to low levels after the initial bleaching response.

Coral bleaching thereby largely reduces the metabolic exchange between corals and associated organisms, as well as reducing the capacity of corals to trap organic matter. This may lead to further loss of POM from the reef system. Therefore, not only are corals affected by thermal stress, but the entire reef system changes and functions differently during and after a bleaching event, particularly when bleaching results in extensive coral mortality. Corals can recover from bleaching, which allows for the return of the described reef engineering functions through production of organic matter by the coral engineer (Fig. 1). The respective recovery time scales range from weeks to months (Gates 1990; Jokiel and Coles 1990), so that short- to mid-term effects on biogeochemical processes can be expected from a brief impediment of the coral engineer. However, these effects may include long-term changes in the reef's recycling capacity.

# Value of coral engineer-driven reef ecosystem goods and services

Reef-building corals act as ecosystem engineers because they provide the physical structure as well as biotic and biogeochemical services to reefs (Moberg and Folke 1999). Such services are functionally similar to other ecosystem engineers such as rainforest trees, mangroves or seagrasses (Guitierrez et al., in press), but usually much more persistent because of the calcareous material of the coral skeleton. However, corals will most likely be the ecosystem engineers most affected by climate change because they are comparatively slow-growing, stenothermic and calcifying organisms, and therefore particularly susceptible to ocean warming and acidification. Global climate change, especially ocean warming that includes coral bleaching, impedes the engineering capacity of scleractinian corals (Fig. 1). This also includes the loss of reef ecosystem services for coastal protection, tourism, and productivity.

Protection of coastal areas and their high associated biodiversity strongly depend on the production of inorganic material by scleractinian corals. These reef structures can absorb 70–90% of wind-generated wave energy (Wells *et al.* 2006) and differentiate habitats and physical environments that influence biological diversity at all scales. The attraction of coral reefs for tourism is probably a reflection of such coral-driven biodiversity.

The total annual economic value of coral reefs via ecosystem services has been estimated at least USD100 000–600 000 km<sup>-2</sup> (Wells *et al.* 2006). The impediment of corals as reef engineers will drastically reduce these economic values, even without consideration of the ecological values (e.g. high biodiversity) if reefs are further degraded under climate change.

Coral engineer-controlled reef biodiversity also has an increasing economic importance for the delivery of new biologically active molecules (e.g. Fung *et al.* 1997; Fung and Ding 1998) that are being developed as pharmaceuticals. Reef productivity and ensuing fishery biomass largely depend on the production of scleractinian corals, but also on their reefengineering functions including the initiation of biogeochemical processes, leading to rapid recycling, and conservation of essential elements, such as nitrogen and phosphorus.

#### **Conclusions**

This review indicates that anthropogenic climate change has the potential to drive major changes across the entire reef ecosystem and indeed is already doing so (Fig. 1). These fundamental changes have the ability to cause both reversible and irreversible changes to reef ecosystem functioning. As part of these interactions, several related aspects are likely to feed back on the role that corals play as ecosystem engineers. Bleaching-induced death of the coral engineers includes the generation of bare skeletons that are particularly sensitive to physical, chemical and biological erosion processes (Stoddart 1969). In addition, colonisation of these stable surfaces by microbial biofilms, algae or other invertebrates may not only reduce recruitment success (e.g. Webster *et al.* 2004), but also change the biogeochemical processes such as nitrogen fixation (Davey *et al.* 2008).

The initiation of carbon and nutrient cycles by coral-derived organic matter will likely be significantly reduced in warmer oceans (Fig. 1), which may alter nutrient dynamics to favour other organisms such as cyanobacteria rather than corals.

Production of calcareous reef sands will probably increase immediately after bleaching-induced mass coral mortality as a result of increased erosion. However, this is a short-term effect lasting only until the carbonate supply diminishes. At large scales, a net loss in carbonate accretion will hypothetically influence the biocatalytic filter systems so that the processing of imported organic matter, for example via eutrophication or river discharge, is reduced (Fig. 1). This may ultimately decrease reef resilience and increase vulnerability of coral-reef ecosystems.

Similar, but not as pronounced, feedback effects on the coral engineer can be expected during and after sub-lethal bleaching (Fig. 1). As the energetic status of corals will be affected, less energy may be invested in the production of inorganic and

organic matter, including carbonates, secondary metabolites, and antibiotic substances (Ritchie 2006) that prevent overgrowth by other organisms and promote resistance against pathogens. Such reductions may change the diversity and activity of coral-associated organisms, which consequently may overgrow or infect corals so that recovery from bleaching is suppressed.

The impacts of global climate change on fundamental physiological processes such as scleractinian coral growth, calcification, defence, maintenance and reproduction result in broad-scale consequences for ecosystem functions and services provided by the reef-building coral engineers. Reduced growth and reproduction translate directly to a reduced resilience of coral-dominated reef communities. Similarly, compromised immune systems and reduction in competitive abilities (Mydlarz et al. 2009; Reed et al. 2010) lead to fundamental changes in the community structure of tropical benthic assemblages (Hoegh-Guldberg et al. 2007). The consequences of reduced calcification rates as a result of ocean warming and acidification will manifest as a reduction in skeletal extension (Jokiel et al. 2008), changes in skeletal crystal morphology (Cohen et al. 2009) or, hypothetically, the formation of less dense skeletons that are highly susceptible to rapid physico-chemical and biological erosion.

Such deleterious impacts of climate change will thereby modify the ability of scleractinian corals to maintain their engineering roles in coral-reef ecosystems. Global influences will also interact with more local and direct disturbances to exacerbate the adverse effects of overfishing and declining water quality (Hughes *et al.* 2007) that additionally impede the functioning of corals as reef ecosystem engineers.

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#### References

- Albright, R., Mason, B., and Langdon, C. (2008). Effect of aragonite saturation state on settlement and post-settlement growth of *Porites astreoides* larvae. *Coral Reefs* 27, 485–490. doi:10.1007/S00338-008-0392-5
- Allemand, D., Ferrier-Pages, C., Furla, P., Houlbreque, F., Puverel, S., et al. (2004). Biomineralisation in reef-building corals: from molecular mechanisms to environmental control. Comptes Rendus Palevol 3, 453–467. doi:10.1016/J.CRPV.2004.07.011
- Alvarez-Filip, L., Dulvy, N. K., Gill, J. A., Cote, I. M., and Watkinson, A. R. (2009). Flattening of Caribbean coral reefs: region-wide decline in architectural complexity. *Proceedings of the Royal Society of London, Series B. Biological Sciences* 276, 3019–3025. doi:10.1098/RSPB.2009. 0339
- Anthony, K. R. N., Kline, D. I., Diaz-Pullido, G., Dove, S., and Hoegh-Guldberg, O. (2008). Ocean acidification causes bleaching and productivity loss in coral reef builders. *Proceedings of the National Academy of Sciences of the United States of America* 105, 17442–17446. doi:10.1073/PNAS.0804478105

- Aronson, R. B., and Precht, W. F. (1997). Stasis, biological disturbance, and community structure of a Holocene coral reef. *Paleobiology* 23, 326–346.
- Aronson, R. B., Macintyre, I. G., Wapnick, C. M., and O'Neill, M. W. (2004). Phase shifts, alternative states, and the unprecedented convergence of two reef systems. *Ecology* 85, 1876–1891. doi:10.1890/03-0108
- Bach, S. D. (1979). Standing crop, growth and production of calcareous Siphonales (Chlorophyta) in a South Florida Lagoon. *Bulletin of Marine Science* 29, 191–201.
- Baird, A. H., and Marshall, P. A. (2002). Mortality, growth and reproduction in scleractinian corals following bleaching on the Great Barrier Reef. *Marine Ecology Progress Series* 237, 133–141. doi:10.3354/MEPS237133
- Bak, R. P. M., Lambrechts, D. Y. M., Joenje, M., Nieuwland, G., and Van Veghel, M. L. J. (1996). Long-term changes on coral reefs in booming populations of a competitive colonial ascidian. *Marine Ecology Pro*gress Series 133, 303–306. doi:10.3354/MEPS133303
- Bak, R. P. M., Nieuwland, G., and Meesters, E. H. (2009). Coral growth rates revisited after 31 years: what is causing lower extension rates in *Acropora palmata? Bulletin of Marine Science* **84**, 287–294.
- Bellwood, D. R., and Hughes, T. P. (2001). Regional-scale assembly rules and biodiversity of coral reefs. *Science* **292**, 1532–1535. doi:10.1126/SCIENCE.1058635
- Bellwood, D. R., Andrew, S. H., Ackerman, J. H., and Depczynski, M. (2006). Coral bleaching, reef fish community phase shifts and the resilience of coral reefs. *Global Change Biology* **12**, 1587–1594. doi:10.1111/J.1365-2486.2006.01204.X
- Bena, C., and van Woesik, R. (2004). The impact of two bleaching events on the survival of small coral colonies (Okinawa, Japan). *Bulletin of Marine Science* **75**, 115–125.
- Borowitzka, M. A., and Larkum, A. W. D. (1977). Calcification in the green alga *Halimeda*. I. An ultrastructure study of thallus development. *Journal of Phycology* **13**, 6–16.
- Bosence, D. (1989). Biogenic carbonate production in Florida Bay. *Bulletin of Marine Science* **44**, 419–433.
- Bosence, D., and Wilson, J. (2003). Maerl growth, carbonate production rates and accumulation rates in the NE Atlantic. *Aquatic Conservation: Marine and Freshwater Ecosystems* 13, S21–S31. doi:10.1002/AOC.565
- Braga, J. C., Martín, J. M., and Riding, R. (1996). Internal structure of segment reefs: *Halimeda* algal mounds in the Mediterranean Miocene. *Geology* **24**, 35–38. doi:10.1130/0091-7613(1996)024<0035:ISOSRH> 2.3.CO;2
- Brandt, M. E., and McManus, J. W. (2009). Disease incidence is related to bleaching extent in reef-building corals. *Ecology* 90, 2859–2867. doi:10.1890/08-0445.1
- Brown, B. E. (1997). Coral bleaching: causes and consequences. *Coral Reefs* **16**, S129–S138. doi:10.1007/S003380050249
- Brown, B. E., Dunne, R. P., Goodson, M. S., and Douglas, A. E. (2002). Experience shapes the susceptibility of a reef coral to bleaching. *Coral Reefs* 21, 119–126.
- Bruno, J. F., Selig, E. R., Casey, K. S., Page, C. A., Willis, B. L., *et al.* (2007). Thermal stress and coral cover as drivers of coral disease outbreaks. *PLoS Biology* 5, e124. doi:10.1371/JOURNAL.PBIO.0050124
- Bythell, J. C. (1988). A total nitrogen and carbon budget for the elkhorn coral *Acropora palmata* (Lamarck). *Proceedings of the 6th International Coral Reef Symposium* **6**, 535–540.
- Bythell, J. C. (1990). Nutrient uptake in the reef-building coral *Acropora* palmata at natural environmental conditions. *Marine Ecology Progress* Series **68**, 65–69. doi:10.3354/MEPS068065
- Caldeira, K., and Wickett, M. E. (2003). Anthropogenic carbon and ocean pH. Nature 425, 365. doi:10.1038/425365A

Cohen, A. L., and Holcomb, M. C. (2009). Why corals care about ocean acidification: uncovering the mechanism. *Oceanography (Washington, D.C.)* 22, 118–127.

- Cohen, A. L., McCorkle, D. C., de Putron, S., Gaetani, G. A., and Rose, K. A. (2009). Morphological and compositional changes in the skeletons of new coral recruits reared in acidified seawater: insights into the biomineralization response to ocean acidification. *Geochemistry Geophysics Geosystems* 10, Q07005. doi:10.1029/2009GC002411
- Colombo-Pallotta, M. F., Rodriguez-Roman, A., and Iglesias-Prieto, R. (2010). Calcification in bleached and unbleached *Montastraea faveolata*: evaluating the role of oxygen and glycerol. *Coral Reefs*. doi:10.1007/S00338-010-0638-X
- Cooney, R. P., Pantos, O., Le Tissier, M. D., Barer, M. R., O'Donnell, A. G., et al. (2002). Characterization of the bacterial consortium associated with black band disease in coral using molecular microbiological techniques. *Environmental Microbiology* 4, 401–413. doi:10.1046/J.1462-2920.2002.00308.X
- Davey, M., Holmes, G., and Johnstone, R. (2008). High rates of nitrogen fixation (acetylene reduction) on coral skeletons following bleaching mortality. *Coral Reefs* 27, 227–236. doi:10.1007/S00338-007-0316-9
- De'ath, G., Lough, J. M., and Fabricius, K. E. (2009). Declining coral calcification on the Great Barrier Reef. *Science* 323, 116–119. doi:10.1126/SCIENCE.1165283
- Diaz-Pulido, G., and McCook, L. J. (2002). The fate of bleached corals: patterns and dynamics of algal recruitment. *Marine Ecology Progress Series* 232, 115–128. doi:10.3354/MEPS232115
- Done, T. J. (1992). Phase shifts in coral reef communities and their ecological significance. *Hydrobiologia* 247, 121–132. doi:10.1007/ BF00008211
- Drew, E. A. (1983). *Halimeda* biomass, growth rates and sediment generation on reefs in the Central Great Barrier Reef province. *Coral Reefs* 2, 101–110. doi:10.1007/BF02395280
- Edmunds, P. J. (2007). Evidence for a decadal-scale decline in the growth rates of juvenile scleractinian corals. *Marine Ecology Progress Series* **341**, 1–13. doi:10.3354/MEPS341001
- Eyre, B., Glud, R. N., and Patten, N. (2008). Mass coral spawning: a natural large-scale nutrient addition experiment. *Limnology and Oceanography* **53**, 997–1013. doi:10.4319/LO.2008.53.3.0997
- Fabricius, K. E., and De'ath, G. (2001). Environmental factors associated with the spatial distribution of crustose coralline algae on the Great Barrier Reef. Coral Reefs 19, 303–309. doi:10.1007/S003380000120
- Feder, D., Mello, C., Garcia, E., and Azambuja, P. (1997). Immune responses in *Rhodnius prolixus*: influence of nutrition and ecdysone. *Journal of Insect Physiology* 43, 513–519. doi:10.1016/S0022-1910(97) 00010-3
- Feely, R. A., Sabine, C. L., Lee, K., Berelson, W., Kleypas, J., *et al.* (2004). The impact of anthropogenic CO<sub>2</sub> on the CaCO<sub>3</sub> system in the oceans. *Science* **305**, 362–366. doi:10.1126/SCIENCE.1097329
- Feely, R. A., Doney, S. C., and Cooley, S. R. (2009). Ocean acidification: present conditions and future changes in a high-CO<sub>2</sub> world. *Oceano-graphy* 22, 36–47.
- Fitt, W. K., Spero, H. J., Halas, J. C., White, M. W., and Porter, J. W. (1993).

  Recovery of the coral *Montastrea annularis* in the Florida Keys after the 1987 Caribbean 'bleaching event'. *Coral Reefs* 12, 57–64. doi:10.1007/BF00302102
- Fitt, W. K., Gates, R. D., Hoegh-Guldberg, O., Bythell, J. C., Jatkar, A., et al. (2009). Response of two species of Indo–Pacific corals, *Porites cylindrica* and *Stylophora pistillata*, to short-term thermal stress: the host does matter in determining the tolerance of corals to bleaching. *Journal of Experimental Marine Biology and Ecology* 373, 102–110. doi:10.1016/J.JEMBE.2009.03.011
- Frias-Lopez, J., Zerkle, A. L., Bonheyo, G. T., and Fouke, B. W. (2002).

  Partitioning of bacterial communities between seawater and healthy, black band diseased, and dead coral surfaces. *Applied and*

- Environmental Microbiology **68**, 2214–2228. doi:10.1128/AEM.68.5. 2214-2228.2002
- Frias-Lopez, J., Klaus, J. S., Bonheyo, G. T., and Fouke, B. W. (2004). Bacterial community associated with black band disease in corals. Applied and Environmental Microbiology 70, 5955–5962. doi:10.1128/ AEM.70.10.5955-5962.2004
- Fung, F. M. Y., and Ding, J. L. (1998). A novel antitumour compound from the mucus of a coral, *Galaxea fascicularis*, inhibits topoisomerase I and II. *Toxicon* 36, 1053–1058. doi:10.1016/S0041-0101(97)00098-6
- Fung, F. M. Y., Tachibana, S., Chou, L. M., and Ding, J. L. (1997). Cytotoxic and anticancer agents in mucus of *Galaxea fascicularis*: purification and characterization. *Journal of Marine Biotechnology* 5, 50–57.
- Gates, R. D. (1990). Seawater temperature and sublethal coral bleaching in Jamaica. Coral Reefs 8, 193–197. doi:10.1007/BF00265010
- Gattuso, J. P., Frankignoulle, M., and Smith, S. V. (1999). Measurement of community metabolism and significance in the coral reef CO<sub>2</sub> source– sink debate. *Proceedings of the National Academy of Sciences of the United States of America* 96, 13 017–13 022. doi:10.1073/PNAS.96.23. 13017
- Glud, R. N., Eyre, B., and Patten, N. (2008). Biogeochemical responses on mass coral spawning at the Great Barrier Reef: effects on respiration and primary production. *Limnology and Oceanography* 53, 1014–1024. doi:10.4319/LO.2008.53.3.1014
- Glynn, P. W. (1993). Coral reef bleaching ecological perspectives. *Coral Reefs* 8, 181–191. doi:10.1007/BF00265009
- Glynn, P. W. (1997). Bioerosion and coral reef growth: a dynamic balance. In 'Life and Death of Coral Reefs'. (Ed. C. Birkeland.) pp. 68–95. (Springer: Berlin.)
- Golbuu, Y., and Richmond, R. H. (2007). Substratum preferences in planula larvae of two species of scleractinian corals, *Goniastrea retiformis* and *Stylaraea punctata*. *Marine Biology* 152, 639–644. doi:10.1007/ S00227-007-0717-X
- Guitierrez, J. L., Jones, C. G., Byers, J. E., Arkema, K. K., Berkenbusch, K., et al. (in press). Physical ecosystem engineers and the functioning of estuaries and coasts. In 'Functioning of Estuaries and Coastal Ecosystems'. (Eds C. H. R. Heip, C. J. M. Philippart and J. J. Middelburg.) (Elsevier: Amsterdam.)
- Haas, A. F., al-Zibdah, M., and Wild, C. (2010). Seasonal in-situ monitoring of coral–algae interaction stability in fringing reefs of the Northern Red Sea. Coral Reefs 29, 93–103. doi:10.1007/S00338-009-0556-Y
- Hallock, P. (1997). Reefs and reef limestones in Earth history. In 'Life and Death of Coral Reefs'. (Ed. C. Birkeland.) pp. 13–42. (Springer: Berlin.)
- Harrington, L., Fabricius, K., De'ath, G., and Negri, A. (2004). Recognition and selection of settlement substrata determine post-settlement survival in corals. *Ecology* 85, 3428–3437. doi:10.1890/04-0298
- Harvell, C. D., Kim, K., Quirolo, C., Weir, J., and Smith, G. (2001). Coral bleaching and disease: contributors to 1998 mass mortality in *Briareum* asbestinum (Octocorallia, Gorgonacea). *Hydrobiologia* 460, 97–104. doi:10.1023/A:1013169331913
- Hoegh-Guldberg, O. (1999). Climate change, coral bleaching and the future of the world's coral reefs. *Marine and Freshwater Research* 50, 839–866. doi:10.1071/MF99078
- Hoegh-Guldberg, O., and Jones, R. J. (1999). Photoinhibition and photoprotection in symbiotic dinoflagellates from reef-building corals. *Marine Ecology Progress Series* 183, 73–86. doi:10.3354/MEPS183073
- Hoegh-Guldberg, O., and Smith, G. J. (1989). The effect of sudden changes in temperature, light and salinity on the population density and export of zooxanthellae from the reef corals Stylophora pistillata (Esper) and Seriatopora hystrix (Dana). Journal of Experimental Marine Biology and Ecology 129, 279–303. doi:10.1016/0022-0981(89)90109-3
- Hoegh-Guldberg, O., Mumby, P. J., Hooten, A. J., Steneck, R. S., Greenfield, P., et al. (2007). Coral reefs under rapid climate change and ocean acidification. Science 318, 1737–1742. doi:10.1126/SCIENCE. 1152509

- Holcomb, M. C., McCorkle, D. C., and Cohen, A. L. (2010). Long-term effects of nutrient and CO<sub>2</sub> enrichment on the temperate coral Astrangia poculata (Ellis and Solander, 1786). Journal of Experimental Marine Biology and Ecology 386, 27–33. doi:10.1016/J.JEMBE.2010.02.007
- Holcomb, M. C., Cohen, A. L., and McCorkle, D. C. (in press). Gender bias in the coral response to ocean acidification. Ocean Sciences Meeting Portland, Oregon, USA; 22–26 February 2010 (American Geophysical Union, Washington, DC.)
- Huettel, M., Røy, H., Precht, E., and Ehrenhauss, S. (2003). Hydrodynamical impact on biogeochemical processes in aquatic sediments. *Hydrobiolo*gia 494, 231–236. doi:10.1023/A:1025426601773
- Huettel, M., Wild, C., and Gonelli, S. (2006). The mucus trap in coral reefs: formation and temporal evolution of aggregates caused by coral mucus. *Marine Ecology Progress Series* 307, 69–84. doi:10.3354/MEPS307069
- Hughes, T. P. (1994). Catastrophes, phase shifts, and large-scale degradation of a Caribbean coral reef. Science 265, 1547–1551. doi:10.1126/ SCIENCE.265.5178.1547
- Hughes, T., Rodrigues, M., Bellwood, D. R., Ceccarelli, D., Hoegh-Guldberg, O., et al. (2007). Phase shifts, herbivory, and the resilience of coral reefs to climate change. Current Biology 17, 360–365. doi:10.1016/J.CUB.2006.12.049
- Iglesias-Prieto, R., Matta, J. L., Robins, W. A., and Trench, R. K. (1992). Photosynthetic response to elevated temperature in the symbiotic dinoflagellate Symbiodinium microadriaticum in culture. Proceedings of the National Academy of Sciences of the United States of America 89, 302–305.
- Iglesias-Prieto, R., Beltran, V. H., LaJeunesse, T. C., Reyes-Bonilla, H., and Thome, P. E. (2004). Different algal symbionts explain the vertical distribution of dominant reef corals in the eastern Pacific. *Proceedings* of the Royal Society of London, Series B. Biological Sciences 271, 1757–1763. doi:10.1098/RSPB.2004.2757
- Jokiel, P. L., and Coles, S. L. (1977). Effects of temperature on the mortality and growth of Hawaiian reef corals. *Marine Biology* 43, 201–208. doi:10.1007/BF00402312
- Jokiel, P. L., and Coles, S. L. (1990). Response of Hawaiian and other Indo– Pacific reef corals to elevated temperature. *Coral Reefs* 8, 155–162. doi:10.1007/BF00265006
- Jokiel, P. L., Rodgers, K. S., Kuffner, I. B., Andersson, A. J., Cox, E. F., et al. (2008). Ocean acidification and calcifying reef organisms: a mesocosm investigation. *Coral Reefs* 27, 473–483. doi:10.1007/S00338-008-0380-9
- Jones, C. G., Lawton, J. H., and Shachak, M. (1994). Organsims as ecosystem engineers. Oikos 69, 373–386. doi:10.2307/3545850
- Jones, C. G., Lawton, J. H., and Shachak, M. (1997). Positive and negative effects of organisms as physical ecosystem engineers. *Ecology* 78, 1946– 1957. doi:10.1890/0012-9658(1997)078[1946:PANEOO]2.0.CO;2
- Jury, C. P., Whitehead, R. F., and Szmant, A. M. (in press). Effects of variations in carbonate chemistry on the calcification rates of *Madracis* mirabilis (Duchassaing 1861): bicarbonate concentrations best predict calcification rates. Global Change Biology. doi:10.1111/J.1365-2486. 2009.02057.X
- Kennedy, D. M., Woodroffe, C. D., Jones, B. G., Dickson, M. E., and Phipps, C. V. G. (2002). Carbonate sedimentation on subtropical shelves around Lord Howe Island and Balls Pyramid, Southwest Pacific. *Marine Geology* 188, 333–349. doi:10.1016/S0025-3227(02)00406-1
- Kleypas, J. A., Buddemeier, R. W., Archer, D., Gattuso, J. P., Langdon, C., et al. (1999). Geochemical consequences of increased atmospheric carbon dioxide on coral reefs. Science 284, 118–120. doi:10.1126/ SCIENCE.284.5411.118
- Knowlton, N., and Rohwer, F. (2003). Multispecies microbial mutualisms on coral reefs: the host as a habitat. *American Naturalist* 162, S51–S62. doi:10.1086/378684
- Langdon, C., and Atkinson, M. J. (2005). Effect of elevated pCO2 on photosynthesis and calcification of corals and interactions with seasonal

- change in temperature/irradiance and nutrient enrichment. *Journal of Geophysical Research* **110**, C09S07. doi:10.1029/2004JC002576
- Lau, K. M., and Wu, H. T. (2003). Warm rain processes over tropical oceans and climate implications. *Geophysical Research Letters* 30, 2290. doi:10.1029/2003GL018567
- Lau, K. M., Wu, H. T., Sud, Y. C., and Walker, G. K. (2005). Effects of cloud microphysics on tropical atmospheric hydrologic processes and intraseasonal variability. *Journal of Climatology* 18, 4731–4751. doi:10.1175/JCLI3561.1
- Leder, J. J., Szmant, A. M., and Swart, P. K. (1991). The effect of prolonged 'bleaching' on skeletal banding and stable isotopic composition in Montastrea annularis. Coral Reefs 10, 19–27. doi:10.1007/BF00301902
- Lesser, M. P., Mazel, C. H., Gorbunov, Y., and Falkowski, P. G. (2004). Discovery of symbiotic nitrogen-fixing cyanobacteria in corals. *Science* **305**, 997–1000. doi:10.1126/SCIENCE.1099128
- Loch, K., Loch, W., Schumacher, H., and See, W. R. (2004). Coral recruitment and regeneration on a Maldivian Reef four years after the coral bleaching event of 1998. Part 2: 2001–2002. *Marine Ecology* 25, 145–154. doi:10.1111/J.1439-0485.2004.00021.X
- Loya, Y. (1976). Recolonization of Red Sea corals affected by natural catastrophes and man-made perturbations. *Ecology* 57, 278–289. doi:10.2307/1934816
- Loya, Y. (2004). The coral reefs of Eilat past, present and future: three decades of coral community structure studies. In 'Coral Health and Disease'. (Eds E. Rosenberg and Y. Loya.) pp. 1–34. (Springer: New York.)
- Loya, Y., Sakai, K., Yamazato, K., Nakano, Y., Sambali, H., et al. (2001).
  Coral bleaching: the winners and the losers. Ecology Letters 4, 122–131.
  doi:10.1046/J.1461-0248.2001.00203.X
- Maliao, R. J., Turingan, R. G., and Lin, J. (2008). Phase-shift in coral reef communities in the Florida Keys National Marine Sanctuary (FKNMS), USA. Marine Biology 154, 841–853. doi:10.1007/S00227-008-0977-0
- Manzello, D. P. (2010). Coral growth with thermal stress and ocean acidification: lessons from the eastern tropical Pacific. *Coral Reefs* **29**, 749–758. doi:10.1007/S00338-010-0623-4
- Manzello, D. P., Brandt, M., Smith, T. B., Lirman, D., Hendee, J. C., et al. (2007). Hurricanes benefit bleached corals. Proceedings of the National Academy of Sciences of the United States of America 104, 12 035– 12 039. doi:10.1073/PNAS.0701194104
- Marshall, P. A., and Baird, A. H. (2000). Bleaching of corals on the Great Barrier Reef: differential susceptibilities among taxa. *Coral Reefs* **19**, 155–163. doi:10.1007/S003380000086
- McClanahan, T. R., and Shafir, S. H. (1990). Causes and consequences of sea urchin abundance and diversity in Kenyan coral reef lagoons. *Oecologia* **83**, 362–370.
- McClanahan, T., Maina, J., and Pet-Soede, L. (2002). Effects of the 1998 coral morality event on Kenyan coral reefs and fisheries. *Ambio* 31, 543–550.
- McCook, L. J. (1999). Macroalgae, nutrients and phase shifts on coral reefs: scientific issues and management consequences for the Great Barrier Reef. *Coral Reefs* **18**, 357–367. doi:10.1007/S003380050213
- Meesters, E. H., and Bak, R. P. M. (1993). Effects of coral bleaching on tissue regeneration potential and colony survival. *Marine Ecology Progress Series* 96, 189–198. doi:10.3354/MEPS096189
- Meszaros, A., and Bigger, C. (1999). Qualitative and quantitative study of wound healing processes in the coelenterate *Plexaurella fusifera*: spatial, temporal, and environmental (light attenuation) influences. *Journal of Invertebrate Pathology* 73, 321–331. doi:10.1006/JIPA.1999.4851
- Miller, J., Waara, R., Muller, E., and Rogers, C. (2006). Coral bleaching and disease combine to cause extensive mortality on reefs of the US Virgin Islands. *Coral Reefs* **25**, 418. doi:10.1007/S00338-006-0125-6
- Moberg, F., and Folke, C. (1999). Ecological goods and services of coral reef ecosystems. *Ecological Economics* **29**, 215–233. doi:10.1016/S0921-8009(99)00009-9

Moret, Y., and Schmid-Hempel, P. (2000). Survival for immunity: the price of immune system activation for bumblebee workers. *Science* 290, 1166–1168. doi:10.1126/SCIENCE.290.5494.1166

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- Morita, M., Suwa, R., Iguchia, A., Nakamura, M., Shimada, K., et al. (2009). Ocean acidification reduces sperm flagellar motility in broadcast spawning reef invertebrates. Zygote 18, 103–107. doi:10.1017/S0967199409990177
- Muller, E. M., Rogers, C. S., Spitzack, A. S., and van Woesik, R. (2008).
  Bleaching increases likelihood of disease on *Acropora palmata* (Lamarck) in Hawksnest Bay, St John, US Virgin Islands. *Coral Reefs* 27, 191–195. doi:10.1007/S00338-007-0310-2
- Mumby, P. J., Chisholm, J. R. M., Edwards, A. J., Clark, C. D., Roark, E. B., et al. (2001). Unprecedented bleaching-induced mortality in *Porites* spp. at Rangiroa Atoll, French Polynesia. *Marine Biology* 139, 183–189. doi:10.1007/S002270100575
- Munday, P. L., Jones, G. P., Pratchett, M. S., and Williams, A. J. (2008). Climate change and the future for coral reef fishes. *Fish and Fisheries* 9, 261–285. doi:10.1111/J.1467-2979.2008.00281.X
- Muscatine, L., Falkowski, P. G., and Porter, J. (1984). Fate of photosynthetic fixed carbon in light and shade-adapted colonies of the symbiotic coral Stylophora pistillata. Proceedings of the Royal Society of London, Series B. Biological Sciences 222, 181–202. doi:10.1098/RSPB.1984.0058
- Mydlarz, L. D., Couch, C. S., Weil, E., Smith, G., and Harvell, C. D. (2009). Immune defenses of healthy, bleached and diseased *Montastraea faveolata* during a natural bleaching event. *Diseases of Aquatic Organisms* 87, 67–78. doi:10.3354/DAO02088
- Nakamura, T., and van Woesik, R. (2001). Water-flow rates and passive diffusion partially explain differential survival of corals during the 1998 bleaching event. *Marine Ecology Progress Series* 212, 301–304. doi:10.3354/MEPS212301
- Nanami, A., and Nishihira, M. (2004). Microhabitat association and temporal stability in reef fish assemblages on massive *Porites* microatolls. *Ichthyological Research* 51, 165–171. doi:10.1007/S10228-004-0213-Y
- Nanami, A., Nishihira, M., Suzuki, T., and Yokochi, H. (2005). Species-specific habitat distribution of coral reef fish assemblages in relation to habitat characteristics in an Okinawan coral reef. *Environmental Biology of Fishes* 72, 55–65. doi:10.1007/S10641-004-4188-3
- Naumann, M., Richter, C., el-Zibdah, M., and Wild, C. (2009). Coral mucus as an efficient trap for picoplanktonic cyanobacteria – implications for pelagic–benthic coupling in the reef ecosystem. *Marine Ecology Progress Series* 385, 65–76. doi:10.3354/MEPS08073
- Naumann, M. S., Haas, A., Struck, U., Mayr, C., el-Zibdah, M., et al. (2010). Organic matter release by the dominant hermatypic corals of the Northern Red Sea. *Coral Reefs* **29**, 649–659. doi:10.1007/S00338-010-0612-7
- Niggl, W., Glas, M., Laforsch, C., Mayr, C., and Wild, C. (2009). First evidence of coral bleaching stimulating organic matter release by reef corals. In 'Proceedings of the 11th International Coral Reef Symposium'. (Ed. B. Riegl.) pp. 905–910. (International Society for Reef Studies: Ft. Lauderdale, FL, USA.)
- Pantos, O., and Bythell, J. C. (2006). Bacterial community structure associated with white band disease in the elkhorn coral Acropora palmata determined using culture-independent 16S rRNA techniques. Diseases of Aquatic Organisms 69, 79–88. doi:10.3354/DAO069079
- Pantos, O., Cooney, R. P., Le Tissier, M. D. A., Barer, M. R., O'Donnell, A. G., et al. (2003). The bacterial ecology of a plague-like disease affecting the Caribbean coral Montastrea annularis. Environmental Microbiology 5, 370–382. doi:10.1046/J.1462-2920.2003.00427.X
- Payri, C. E. (1988). *Halimeda* contribution to organic and inorganic production in a Tahitian reef system. *Coral Reefs* 6, 251–262. doi:10.1007/BF00302021
- Piggot, A. M., Fouke, B. W., Sivaguru, M., Sanford, R. A., and Gaskins, H. R. (2009). Change in zooxanthellae and mucocycte tissue density as an adaptive response to environmental stress by the coral, *Montastrea*

- annularis. Marine Biology 156, 2379–2389. doi:10.1007/S00227-009-1267-1
- Porter, J. W., Fitt, W. K., Spero, H. J., Rogers, C. S., and White, M. N. (1989).
  Bleaching in reef corals: physiological and stable isotopic responses.
  Proceedings of the National Academy of Sciences of the United States of America 86, 9342–9346. doi:10.1073/PNAS.86.23.9342
- Pratchett, M. S. (2007). Dietary selection by coral-feeding butterfly fishes (Chaetodontidae) on the Great Barrier Reef, Australia. *The Raffles Bulletin of Zoology* 14, 171–176.
- Raes, M., De Troch, M., Ndaro, S. G. M., Muthumbi, A., Guilini, K., et al. (2007). The structuring role of microhabitat type in coral degradation zones: a case study with marine nematodes from Kenya and Zanzibar. Coral Reefs 26, 113–126. doi:10.1007/S00338-006-0184-8
- Rasheed, M., Badran, M., and Huettel, M. (2003). Particulate matter filtration and seasonal nutrient dynamics in permeable carbonate and silicate sands of the Gulf of Aqaba, Red Sea. *Coral Reefs* 22, 167–177. doi:10.1007/S00338-003-0300-Y
- Reaka-Kudla, M. L. (1997). The global biodiversity of coral reefs: a comparison with rain forests. In 'Biodiversity II: Understanding and Protecting our Biological Resources'. (Eds M. L. Reaka-Kudla, D. E. Wilson and E. O. Wilson.) pp. 83–108. (Joseph Henry Press: Washington, DC.)
- Reed, K. C., Muller, E. M., and van Woesik, R. (2010). Coral immunology and resistance to disease. *Diseases of Aquatic Organisms* 90, 85–92. doi:10.3354/DAO02213
- Reshef, L., Koren, O., Loya, Y., Zilber-Rosenberg, I., and Rosenberg, E. (2006). The coral probiotic hypothesis. *Environmental Microbiology* **8**, 2068–2073. doi:10.1111/J.1462-2920.2006.01148.X
- Rinkevich, B. (1999). Invertebrates versus vertebrates innate immunity: in the light of evolution. *Scandinavian Journal of Immunology* **50**, 456–460. doi:10.1046/J.1365-3083.1999.00626.X
- Ritchie, K. B. (2006). Regulation of microbial populations by coral surface mucus-associated bacteria. *Marine Ecology Progress Series* 322, 1–14. doi:10.3354/MEPS322001
- Rodrigues, L. J., and Grottoli, A. G. (2006). Calcification rate and stable carbon, oxygen, and nitrogen isotopes in the skeleton, host tissue, and zooxanthellae of bleached and recovering Hawaiian corals. *Geochimica* et Cosmochimica Acta 70, 2781–2789. doi:10.1016/J.GCA.2006.02.014
- Rodrigues, L. J., Grottoli, A. G., and Pease, T. K. (2008). Lipid class composition of bleached and recovering *Porites compressa* (Dana, 1846) and *Montipora capitata* (Dana, 1846) corals from Hawaii. *Journal of Experimental Marine Biology and Ecology* 358, 136–143. doi:10.1016/J.JEMBE.2008.02.004
- Rodríguez-Román, A., Hernández-Pech, X., Thomé, P., Enríquez, S., and Iglesias-Prieto, R. (2006). Photosynthesis and light utilization in the Caribbean coral *Montastraea faveolata* recovering from a bleaching event. *Limnology and Oceanography* 51, 2702–2710. doi:10.4319/ LO.2006.51.6.2702
- Rohwer, F., Seguritan, V., Azam, F., and Knowlton, N. (2002). Diversity and distribution of coral-associated bacteria. *Marine Ecology Progress Series* 243, 1–10. doi:10.3354/MEPS243001
- Rowan, R., Knowlton, N., Baker, A., and Jara, J. (1997). Landscape ecology of algal symbionts creates variation in episodes of coral bleaching. *Nature* 388, 265–269. doi:10.1038/40843
- Rypien, K. L., Ward, J. R., and Azam, F. (2010). Antagonistic interactions among coral-associated bacteria. *Environmental Microbiology* 12, 28–39. doi:10.1111/J.1462-2920.2009.02027.X
- Sale, P. F., Danilowicz, B. S., Doherty, P. J., and Williams, D. M. (2005). The relation of microhabitat to variation in recruitment of young-of-year coral reef fishes. *Bulletin of Marine Science* 76, 123–142.
- Seppälä, O., Liljeroos, K., Karvonen, A., and Jukka, J. (2008). Host condition as a constraint for parasite reproduction. *Oikos* 117, 749–753. doi:10.1111/J.0030-1299.2008.16396.X

- Shashar, N., Cohen, Y., Loya, Y., and Sar, N. (1994). Nitrogen fixation (acetylene reduction) in stony corals: Evidence for coral-bacteria interactions. *Marine Ecology Progress Series* 111, 259–264. doi:10.3354/ MEPS111259
- Smith, J. E., Shaw, M., Edwards, R. A., Obura, A., Pantos, O., et al. (2006). Indirect effects of algae on coral: algae-mediated, microbe-induced coral mortality. Ecology Letters 9, 835–845. doi:10.1111/J.1461-0248.2006. 00937 X
- Srinivasan, M. (2003). Depth distributions of coral reef fishes: the influence of microhabitat structure, settlement, and post-settlement processes. *Oecologia* 137, 76–84. doi:10.1007/S00442-003-1320-6
- Stedman, T. L. (2000). 'Stedman's Medical Dictionary.' (Lippincott, Williams and Wilkins: Baltimore.)
- Stoddart, D. R. (1969). Ecology and morphology of recent coral reefs. Biological Reviews of the Cambridge Philosophical Society 44, 433–498. doi:10.1111/J.1469-185X.1969.TB00609.X
- Sud, Y. C., Walker, G. K., Zhou, Y. P., Schmidt, G. A., and Lau, K. M. (2008). Effects of doubled CO<sub>2</sub> on tropical sea surface temperatures (SSTs) for onset of deep convection and maximum SST: Simulations based inferences. *Geophysical Research Letters* 35, L12707. doi:10.1029/2008GL033872
- Suzuki, A., and Kawahata, H. (2003). Carbon budget of coral reef systems: an overview of observations in fringing reefs, barrier reefs and atolls of the Indo–Pacific regions. *Tellus* 55B, 428–444.
- Takahashi, S., Nakamura, T., Sakamizu, M., van Woesik, R., and Yamasaki, H. (2004). Repair machinery of symbiotic photosynthesis as the primary target of heat stress for reef-building corals. *Plant & Cell Physiology* 45, 251–255. doi:10.1093/PCP/PCH028
- Tanzil, J. T. I., Brown, B. E., Tudhope, A. W., and Dunne, R. P. (2009). Decline in skeletal growth of the coral *Porites lutea* from the Andaman Sea, South Thailand, between 1984 and 2005. *Coral Reefs* 28, 519–528. doi:10.1007/S00338-008-0457-5
- Van Oppen, M. J. H., Palstra, F. P., Piquet, A. M., and Miller, D. J. (2001).
  Patterns of coral-dinoflagellate associations in *Acropora*: significance of local availability and physiology of *Symbiodinium* strains and host-symbiont selectivity. *Proceedings of the Royal Society of London, Series B. Biological Sciences* 268, 1759–1767. doi:10.1098/RSPB.2001.
- Ward, S., Harrison, P., and Hoegh-Guldberg, O. (2002). Coral bleaching reduces reproduction of scleractinian corals and increases susceptibility to future stress. In 'Proceedings of the 9th International Coral Reef Symposium'. pp. 1123–1128. (International Society for Reef Studies: Bali, Indonesia.)
- Webster, N. S., Smith, L. D., Heyward, A. J., Watts, J. E. M., Webb, R. I., et al. (2004). Metamorphosis of a scleractinian coral in response to microbial biofilms. Applied and Environmental Microbiology 70, 1213–1221. doi:10.1128/AEM.70.2.1213-1221.2004

- Wells, S., Ravilious, C., and Corcoran, E. (2006). 'In the Front Line Shoreline Protection and Other Ecosystem Services from Mangroves and Coral Reefs.' (UNEP-WCMC: Cambridge.)
- Werner, U., Bird, P., Wild, C., Ferdelman, T., Polerecky, L., et al. (2006). Spatial variability of aerobic and anaerobic mineralization in coral reef sediments (Heron Island, Australia). Marine Ecology Progress Series 309, 93–105. doi:10.3354/MEPS309093
- Wild, C., Huettel, M., Klueter, A., Kremb, S. G., Rasheed, M., et al. (2004a).
  Coral mucus functions as an energy carrier and particle trap in the reef ecosystem. Nature 428, 66–70. doi:10.1038/NATURE02344
- Wild, C., Rasheed, M., Werner, U., Franke, U., Johnstone, R., et al. (2004b). Degradation and mineralization of coral mucus in reef environments. Marine Ecology Progress Series 267, 159–171. doi:10.3354/MEPS267159
- Wild, C., Tollrian, R., and Huettel, M. (2004c). Rapid recycling of coral mass spawning products in permeable reef sediments. *Marine Ecology Progress Series* 271, 159–166. doi:10.3354/MEPS271159
- Wild, C., Rasheed, M., Jantzen, C., Cook, P., Struck, U., et al. (2005a).
  Benthic metabolism and degradation of natural particulate organic matter in silicate and carbonate sands of the Northern Red Sea. Marine Ecology Progress Series 298, 69–78. doi:10.3354/MEPS298069
- Wild, C., Woyt, H., and Huettel, M. (2005b). Influence of coral mucus release on nutrient fluxes in carbonate sands. *Marine Ecology Progress Series* **287**, 87–98. doi:10.3354/MEPS287087
- Wild, C., Laforsch, C., and Huettel, M. (2006). Detection and enumeration of microbial cells in highly porous carbonate reef sands. *Marine and Freshwater Research* 57, 415–420. doi:10.1071/MF05205
- Wild, C., Jantzen, C., Struck, U., Hoegh-Guldberg, O., and Huettel, M. (2008). Biogeochemical responses on coral mass spawning at the Great Barrier Reef: pelagic-benthic coupling. *Coral Reefs* 27, 123–132. doi:10.1007/S00338-007-0298-7
- Wild, C., Naumann, M. S., Haas, A., Struck, U., Mayer, F. W., et al. (2009).
  Coral sand O<sub>2</sub> uptake and pelagic–benthic coupling in a subtropical fringing reef, Aqaba, Red Sea. Aquatic Biology 6, 133–142. doi:10.3354/AB00181
- Williams, W. M., Viner, A. B., and Broughton, W. J. (1987). Nitrogen fixation (acetylene reduction) associated with the living coral *Acropora* variabilis. Marine Biology 94, 531–535. doi:10.1007/BF00431399
- Willis, B. L., Page, C. A., and Dinsdale, E. A. (2004). Coral disease on the Great Barrier Reef. In 'Coral Health and Disease'. (Eds E. Rosenberg and Y. Loya.) pp. 69–104. (Springer: New York.)
- Wilson, S. K., Graham, N. A. J., Pratchett, M. S., Jones, J. P., and Polunin, N. V. C. (2006). Multiple disturbances and the global degradation of coral reefs: are reef fishes at risk or resilient? *Global Change Biology* 12, 2220–2234. doi:10.1111/J.1365-2486.2006.01252.X

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