

JCU ePrints

This file is part of the following reference:

Canyon, Sarah J. (2003) *A novel cardioprotective therapy : adenosine and lidocaine solution in an in vivo rat model of acute myocardial ischemia-reperfusion.* PhD thesis, James Cook University.

Access to this file is available from:

<http://eprints.jcu.edu.au/1642>

A Novel Cardioprotective Therapy:

Adenosine and Lidocaine Solution
in an *In Vivo* Rat Model of Acute
Myocardial Ischemia-Reperfusion

Thesis submitted by Sarah J. Canyon BSc (Hons) JCU
in October 2003

For the degree of Doctor of Philosophy
School of Biomedical sciences
Department of Physiology and Pharmacology
James Cook University of North Queensland, Australia

ELECTRONIC COPY

I, the undersigned, the author of this work, declare that the electronic copy of this thesis provided to the James Cook University Library, is an accurate copy of the print thesis submitted, within the limits of the technology available.

Signature

Date

Statement of Access

Declaration

I, the undersigned, the author of this thesis, understand that James Cook University of North Queensland will make it available for use within the University Library and, by microfilm or other means, allow access to users in other approved libraries. All users consulting this thesis will have to sign the following statement:

In consulting this thesis, I agree not to copy or closely paraphrase it in whole or in part without the written consent of the author; and to make proper public written acknowledgment for any assistance which I have obtained from it.

Beyond this, I do not wish to place any restriction on access to this thesis.

.....
(Signature)

.....
(Date)

Statement of Sources

Declaration

I declare that this thesis is my own work and has not been submitted in any form for another degree or diploma at any university or other institution of tertiary education. Information derived from the published or unpublished work of others has been acknowledged in the text and a list of references is given.

.....
(Signature)

.....
(Date)

Acknowledgements

Immense gratitude is owed to Associate Professor Geoff Dobson for his outstanding knowledge, guidance and infectious enthusiasm regarding scientific discovery. He is a true lover of the fundamentals of basic science from its philosophical roots to its application in education and the laboratory. In our lab, Geoff treats his students and technicians as colleagues and creates an atmosphere of encouragement so great that the smallest achievements are praised with the same zeal as the larger ones. The effect of this is to enhance performance and respect for the power of science and its limitations.

Much appreciation goes to Sam Hitchins for his willingness to come in and provide support with various aspects of the *in vivo* model and especially the NMR and to Uwe Himmelreich who also went out of his way to help me gain NMR confidence. Thanks to the other talented postgrads, Michael Jones, Donna Rudd and Kathy Sloots for friendship, laughs, company and often guidance. Furthermore, special thanks to Mr. John Atkinson, whose special aptitude for solving mechanical puzzles enriched many of the experiments presented in this work.

Further acknowledgements are owed to the School of Biomedical Sciences and School of Pharmacy and Molecular sciences, especially administrative and technical staff, for continued friendly support throughout the course of my doctorate.

Lastly, I wish to acknowledge the tremendous support, understanding and seemingly endless patience of my husband, Deon Canyon and my three children, Chauncy, Kestrel and Rowan. Moreover, I want to thank my father, Professor Cooper Lansing, for it was trust in his wisdom that steered me towards science and academic pursuits early on in life.

Abstract

Background: Recently, our laboratory demonstrated that an adenosine and lidocaine (AL) non-depolarizing cardioplegic arrest solution conferred superior protection during arrest and recovery compared with the hyperkalemic depolarizing St. Thomas' Hospital cardioplegic solution in isolated rat hearts. The aim of this thesis was to extend those findings by applying an adenosine and lidocaine (AL) solution at nonarresting concentrations before and during ischemia in an *in vivo* rat model of acute myocardial ischemia. No study has investigated the effect of AL combination treatment to reduce ischemic injury. Yet, previous studies in the 1990s have used the sequential and separate administration of lidocaine (2 mg/kg i.v.) and adenosine (150 µg/kg/ml/min i.c) as reperfusion therapy with conflicting results.

Methods: In all four studies, ischemia-reperfusion was achieved by placing a reversible tie around the left coronary artery of anaesthetized (sodium pentobarbital, 60 mg/ml/kg i.p.) and ventilated male Sprague-Dawley rats (300 - 400 g). The ischemic period lasted 30 min while reperfusion times were maintained for either 40 min or 120 min. Where applicable, a lead II electrocardiogram, heart rate, and systolic and diastolic pressures were recorded and mean arterial pressure and rate-pressure product calculated. The primary end points included infarct size, episodes and durations of ventricular arrhythmias, pH and changes in the concentration of ATP ([ATP]) and phosphocreatine ([PCr]) during ischemia-reperfusion recorded every 5 min. The level of statistical significance was $P < 0.05$.

Experimental design: The first two studies examined the cardioprotective potential of adenosine and lidocaine using the following treatment strategies: i) three AL solutions with varying concentrations of adenosine (A: 152, 305 and 457 µg/kg/min

Abstract

plus L: 608 µg/kg/min i.v., n = 18) compared to saline controls (0.9% saline, n = 12), adenosine only (adeno-only, 305 µg/kg/min i.v., n = 8) and lidocaine only (lido-only, 608 µg/kg/min i.v., n = 8); all of these treatments were given 5 before ischemia and continued throughout 30 min ischemia but not reperfusion; and ii) the separate and sequential administration of adenosine (150 µg/kg/min i.v.) and lidocaine (2 mg/kg i.v.) during reperfusion (n = 7); the sequential administration of AL solution (A: 305 µg/kg/min plus L: 608 µg/kg/min i.v.) of 5 min pretreatment and again 5 min before and during 30 min reperfusion, n = 6 and a 5 min pretreatment of AL solution (A: 305 µg/kg/min plus L: 608 µg/kg/min i.v.) continued throughout ischemia and 30 min of reperfusion (n = 6). In the third study, ^{31}P nuclear magnetic resonance was used to investigate the changes in [ATP], [PCr] and pH during ischemia (30 min) and reperfusion (40 min) with AL solution treatment (n = 6) or in controls (n = 7). In the fourth study, AL solution (A: 305 µg/kg/min plus L: 608 µg/kg/min i.v.) was compared to that of ischemic preconditioning (three 3 min cycles of ischemia-reperfusion) (IPC) (n = 6), the adenosine receptor A₁ agonist, 2-chloro-N6-cyclopentyladenosine (CCPA) (5 µg/kg i.v.) plus lidocaine (n = 6), and CCPA alone (n = 7).

Results: Seven of the 12 saline-control rats and 4 of the 8 Adeno-only treated rats died during the ischemic period from an episode of ventricular fibrillation. No deaths occurred in the Lido-only treated rats (n = 6) or in any group where AL solution was infused. Ventricular tachycardia (VT) occurred in 100% of saline controls (18 ± 9 episodes), 50% of the adeno-only group (11 ± 7 episodes), and 83% of lido-only treatment (23 ± 11 episodes). VT was also experienced in 60% of low-dose AL treated rats (2 ± 1 episodes) ($P < 0.05$), 57% of mid-dose AL (2 ± 1 episodes) ($P < 0.05$), and 67% of high-dose AL treated rats (6 ± 3 episodes). Ventricular fibrillation

Abstract

(VF) occurred in 75% of saline controls (4 ± 3 episodes), 100% of adeno-only (3 ± 2 episodes), and in 33% lido-only treated rats (2 ± 1 episodes). Low-dose AL and mid-dose AL completely prevented VF from occurring during ischemia. The mean infarct size of mid dose-AL ($38 \pm 6\%$) was significantly reduced from saline controls ($61 \pm 5\%$), adeno-only ($56 \pm 4\%$), and lido-only ($66 \pm 8\%$) ($P < 0.05$) but not from low-dose AL ($45 \pm 9\%$) and high-dose AL animals ($45 \pm 6\%$).

The separate and sequential administration of lidocaine and adenosine resulted in 2 out of 7 deaths and ischemia-induced VT (6 ± 3 episodes, 4 ± 2 sec) was not prevented while VF (1 ± 0 episodes, 1 ± 0 sec) was reduced. Infarct size ($52 \pm 5\%$) was not significantly different from saline-controls ($61 \pm 5\%$). When AL was given at pretreatment, stopped for ischemia and resumed 5 min before reperfusion, infarct size reduction ($67 \pm 8\%$) and protection from ventricular arrhythmias (VT: 39 ± 23 episodes, 84 ± 49 sec; VF: 2 ± 1 episodes, 21 ± 8 sec) were lost; though, there were no deaths in this group. AL solution given continuously from pretreatment through ischemia and reperfusion provided similar protection to AL infusion during pretreatment and ischemia ($41 \pm 10\%$ vs. $38 \pm 6\%$, 2 ± 1 VT and 0 VF).

During ischemia, control [ATP] fell to 61% of baseline at 15 min and recovered 68% - 88% of baseline during reperfusion. AL treatment maintained [ATP] in a steady state throughout ischemia and reperfusion with changes ranging of $95 \pm 7\%$ to $117 \pm 10\%$ of baseline. Control [PCr] was significantly reduced compared to AL treated hearts during ischemia at 10 min (62 ± 7 vs. $89 \pm 9\%$), 15 min ($45 \pm 4\%$ vs $81 \pm 7\%$), 20 min ($44 \pm 9\%$ vs. $92 \pm 9\%$) and 30 min ($45 \pm 8\%$ vs. $77 \pm 7\%$) and during reperfusion at 10 min ($44 \pm 19\%$ vs. $92 \pm 9\%$) and 15 min ($50 \pm 8\%$ vs. $90 \pm 7\%$) ($P < 0.05$). The pH

Abstract

of AL and control hearts were similar throughout ischemia ranging from pH 7.6 to 6.4 in control and pH 7.5 to 6.8 in AL hearts. Controls maintained a mean pH below baseline for the first 20 min of reperfusion (pH 7.1) while AL hearts pH recovered to baseline within the first 5 min of reperfusion (pH 7.4 ± 0.1).

Pretreating the heart before and during ischemia with AL or with CCPA plus lidocaine resulted in no deaths, and no lethal arrhythmias. Infarct size reduction in CCPA plus lidocaine treated rats ($12 \pm 4\%$) was similar to ischemic preconditioning ($11 \pm 3\%$), whereas in AL- and CCPA- treated rats, the infarct size was $38 \pm 6\%$ and $42 \pm 7\%$ respectively.

Conclusions: i) The intravenous infusion of AL solution before or during 30 min was more cardioprotective than adenosine alone, lidocaine alone, or the separate and sequential infusion of adenosine and lidocaine; ii) the AL combination led to no death, virtually no episodes of VF, few episodes of VT, and a significantly reduced infarct size; iii) AL cardioprotection appears to be associated with preservation of high energy phosphates and a better balance between supply and demand during ischemic conditions; iv) low pH was not an indicator of myocardial damage in AL treated rats, and v) when adenosine was substituted with an adenosine A₁ receptor agonist, CCPA, plus lidocaine cardioprotection was significantly enhanced and similar to IPC. In summary, targeting adenosine receptors, especially the A₁ adenosine receptor, with lidocaine Na⁺ fast channel modulation may offer a new combination therapy to delay myocardial damage during ischemia and prevent ischemia-induced arrhythmias.

Table of Contents

TITLE.....	I
STATEMENT OF ACCESS	III
STATEMENT OF SOURCES.....	IV
ACKNOWLEDGEMENTS	V
ABSTRACT.....	VI
ABBREVIATIONS.....	XV
LIST OF FIGURES.....	XVI
LIST OF ILLUSTRATIONS	XIX
CHAPTER 1. INTRODUCTION.....	1
1.1. CARDIOPROTECTION	2
1.2. A BRIEF HISTORICAL PERSPECTIVE OF THE HEART STRUCTURE AND FUNCTION.....	4
1.3. ENERGY SUPPLY TO THE MYOCARDIUM.....	7
1.4. ACUTE MYOCARDIAL ISCHEMIA-REPERFUSION INJURY	9
<i>1.4.1 Acute ischemia</i>	<i>10</i>
1.4.1.1 Development of cell death: The transition from reversible to irreversible injury.....	14
1.4.1.2. Ischemic electrophysiology.....	15
1.4.1.3. Arrhythmias arising from the ischemic myocardium.....	17
<i>1.4.2. Reperfusion injury.....</i>	<i>19</i>
1.4.2.1. The ‘oxygen paradox’	20
1.4.2.2. Free radical formation	21
1.4.2.3. Myocardial stunning.....	22
1.4.2.4. Inflammation.....	24
1.5. ISCHEMIC PRECONDITIONING- A PARADIGM FOR CARDIOPROTECTION	25
<i>1.5.1. Possible mechanisms of ischemic preconditioning.....</i>	<i>26</i>
1.5.1.1. ATP-sensitive potassium channels	27
<i>1.5.2. Phases of ischemic preconditioning cardioprotection</i>	<i>31</i>
<i>1.5.3. Protective outcomes of ischemic preconditioning</i>	<i>31</i>
1.6. PHARMACOLOGICAL TARGETS BASED ON THE KNOWN MECHANISMS OF ISCHEMIA-REPERFUSION INJURY AND ISCHEMIC PRECONDITIONING.....	32

Table of Contents

1.7. ADENOSINE	33
1.7.1. Adenosine synthesis during normoxia, hypoxia and ischemia	35
1.7.1.1. Vascular Adenosine Synthesis	36
1.7.1.2. Adenosine metabolism	37
1.7.2. Pharmacology of adenosine and adenosine receptors	40
1.7.2.1. Adenosine A ₁ receptor.....	41
1.7.2.2. The A _{2a} and A _{2b} receptors	43
1.7.2.3. Adenosine A ₃ receptor.....	44
1.8. LIDOCAINE	46
1.8.1. Antiarrhythmic Pharmacological effects of lidocaine	48
1.8.2. Additional cardioprotective properties of lidocaine.....	52
1.9. SIMULTANEOUS ADENOSINE AND LIDOCAINE FOR REPERFUSION THERAPY	53
1.10. SUMMARY OF RESEARCH AIMS	55
CHAPTER 2. MATERIALS AND METHODS	57
2.1. INTRODUCTION	58
2.2. ANIMALS AND REAGENTS	58
2.3. <i>IN VIVO</i> RAT MODEL OF ACUTE MYOCARDIAL ISCHEMIA SURGICAL PROTOCOL ...	59
2.4. MEASUREMENT OF ISCHEMIC AREA AT RISK AND INFARCT SIZE	63
2.4.1. Introduction to the main concepts of infarct size measurement.....	63
2.4.2 The protocol used for infarct size measurement.....	64
2.5. HEMODYNAMIC MEASUREMENTS	67
2.6. IDENTIFICATION AND ANALYSIS OF ARRHYTHMIAS	67
2.7. <i>IN VIVO</i> ³¹ P MAGNETIC RESONANCE SPECTROSCOPY OF THE RAT HEART	70
2.7.1. Modifications to surgical protocol.....	70
2.7.2. Calibration of surface coil sampling depth.....	71
2.7.3. NMR spectroscopy.....	71
2.7.4. Phosphorus Quantification.....	74
2.7.5. Intracellular pH.....	76
2.7.6. Free Magnesium (Mg ²⁺).....	77
2.8. GENERAL EXPERIMENTAL DESIGN	77
2.9. STATISTICAL ANALYSIS	78

Table of Contents

CHAPTER 3. PROTECTION AGAINST VENTRICULAR ARRHYTHMIAS AND CARDIAC DEATH USING AN ADENOSINE AND LIDOCAINE (AL) SOLUTION DURING ACUTE MYOCARDIAL ISCHEMIA-REPERFUSION.....	80
3.1. INTRODUCTION	81
3.2. EXPERIMENTAL DESIGN.....	83
3.4. RESULTS.....	84
3.4.1. <i>Mortality</i>	84
3.4.2. <i>Arrhythmias during Ischemia</i>	85
3.4.3 <i>Early Reperfusion Arrhythmias</i>	89
3.4.4. <i>Infarct Size</i>	89
3.4.5. <i>Systemic Hemodynamics</i>	91
3.5. DISCUSSION.....	94
3.5.1. <i>AL Solution's antiarrhythmic actions and survival benefit</i>	94
3.5.1.1. <i>Ischemia-induced arrhythmias</i>	95
3.5.1.2. <i>Reperfusion-induced Arrhythmias</i>	97
3.5.2. <i>Proarrhythmic effects of adenosine and lidocaine alone</i>	97
3.5.3. <i>Effect on infarct size</i>	99
3.5.4. <i>Possible mechanisms of action for AL solution in ischemia and reperfusion</i>	100
3.5.5. <i>Conclusion</i>	101
3.5.6. <i>Limitations of the Study</i>	102
CHAPTER 4. AL CARDIOPROTECTION: ISCHEMIC VS REPERFUSION THERAPY.....	104
4.1. INTRODUCTION	105
4.2. EXPERIMENTAL DESIGN.....	107
4.3. RESULTS.....	109
4.3.1 <i>Mortality</i>	109
4.3.2. <i>Arrhythmias during Ischemia: episodes and durations</i>	109
4.3.2.1. <i>Early Reperfusion Arrhythmias</i>	112
4.3.3. <i>Infarct Size</i>	112
4.3.4. <i>Systemic Hemodynamics</i>	114

4.4. DISCUSSION.....	116
4.4.1 <i>Conclusions and Interpretation</i>	119
CHAPTER 5. ^{31}P NMR SPECTROSCOPIC ANALYSIS OF THE EFFECT OF AL SOLUTION ON ENERGETIC METABOLISM AND INTRACELLULAR PH DURING ACUTE MYOCARDIAL ISCHEMIA	122
5.1. INTRODUCTION	123
5.2. EXPERIMENTAL DESIGN	124
5.3. RESULTS.....	125
5.3.1. <i>Hemodynamics</i>	125
5.3.2. <i>Effects of AL solution on bioenergetic responses to ischemia-reperfusion</i>	128
5.3.2.1. ATP Concentration ([ATP]).....	128
5.3.2.2. PCr concentration ([PCr])	129
5.3.2.3. Inorganic phosphate ($[\text{P}_i]$)	131
5.3.2.4. Intracellular pH.....	132
5.3.2.5. Free magnesium ($[\text{Mg}^{2+}]$).....	133
5.4. DISCUSSION.....	134
5.4.1. <i>Metabolic features of AL treated hearts compared to controls</i>	134
5.4.2. <i>Myocardial protection by AL treatment: maintenance of a more balanced energetic steady-state</i>	135
5.4.3. <i>Acidosis with cardioprotection in AL treated hearts</i>	138
5.4.4. <i>Intracellular free magnesium ($[\text{Mg}^{2+}]$)</i>	141
5.4.5. <i>Conclusions</i>	142
5.4.6. <i>Limitations with NMR sampling of in vivo rat NMR</i>	142
CHAPTER 6. PHARMACOLOGICAL PRECONDITIONING: CONCOMITANT TARGETING OF THE ADENOSINE A₁ RECEPTOR AND SODIUM CHANNELS SURPASSES AL SOLUTION CARDIOPROTECTION.....	145
6.1. INTRODUCTION	146
6.2. EXPERIMENTAL DESIGN.....	147
6.3 RESULTS.....	149
6.4. DISCUSSION.....	154

Table of Contents

CHAPTER 7. DISCUSSION.....	158
7.1. RESTATEMENT OF THE HYPOTHESIS	159
7.2. SUMMARY OF PRIMARY FINDINGS.....	159
7.3. POSSIBLE MECHANISMS OF ACTION RESPONSIBLE FOR AL CARDIOPROTECTION	
.....	161
<i>7.3.1. Regulation of transmembrane ion distribution and improved Na⁺ and Ca²⁺ handling.....</i>	161
<i>7.3.2. Possible role for AL protection from inflammation injury.....</i>	164
<i>7.3.3. Possible clinical significance and limitations of AL infusion therapy....</i>	166
7.4. CONCLUDING REMARKS.....	168
REFERENCES	170
APPENDIX.....	A1

Abbreviations

³¹ P	phosphorus-31
adeno	adenosine
ADP	adenosine diphosphate
AL	adenosine and lidocaine
AL solution	adenosine and lidocaine solution
AMP	adenosine monophosphate
APD	action potential duration
ATP	adenosine triphosphate
bpm	beats per minute
Ca ²⁺	calcium ion
CCPA	2-chloro-N6-cyclopentyladenosine
CK	creatine kinase
Cr	creatinine
D ₂ O	deuterium oxide
ECG	electrocardiogram
FID	free induction decay
H ⁺	hydrogen ion
HR	heart rate
Hrs	hours
i.c.	intracoronary
i.p.	intraperitoneal
i.v.	intravenous
K _{ATP}	ATP sensitive potassium channel
Lido	lidocaine hydrochloride
Map	mean arterial pressure
Mg ²⁺	free magnesium
min	minutes
Mito	mitochondrial
Na _i	intracellular sodium
NMR	nuclear magnetic resonance
o.d.	outer diameter
PCr	phosphocreatine
pH	-log ₁₀ [H ⁺]
P _i	inorganic phosphate
PKC	protein kinase C
PPA	phenylphosphoric acid
Ppm	parts per million
PVB	premature ventricular beat
rpp	rate pressure product
Sarc	sarcolemmal
SCF	saturation correction factor
Sec	seconds
solution	solution
T	tesla
TTC	triphenyltetrazolium chloride
VF	ventricular fibrillation
VT	ventricular tachycardia
VT+VF	sum of VT and VF

List of figures

FIGURE 1.1. HIGH ENERGY PHOSPHATE, ATP AND PHOSPHOCREATINE (PCr), UTILIZATION IN NORMAL MYOCARDIUM.....	8
FIGURE 1.2. EFFECT OF INCREASED INTRACELLULAR CALCIUM ON THE GENESIS OF ISCHEMIA-INDUCE ARRHYTHMIAS.	19
FIGURE 1.3. SCHEMATIC DIAGRAM OF SOME CURRENT IDEAS ON IPC SIGNAL TRANSDUCTION.....	28
FIGURE 1.4. ADENOSINE FORMATION, METABOLISM AND RECEPTOR-EFFECTOR COUPLING.....	38
FIGURE 1.5. EFFECT OF LIDOCAINE ON Na^+ CHANNEL RECOVERY FROM INACTIVATION IN VENTRICULAR MYOCYTES.	49
FIGURE 1.6. DIAGRAM OF THE MAJOR FEATURES OF LIDOCAINE'S EFFECT ON VENTRICULAR ACTION POTENTIALS.....	50
FIGURE 2.1. PHOTOGRAPH OF THE <i>IN VIVO</i> RAT MODEL OF ACUTE MYOCARDIAL ISCHEMIA.....	60
FIGURE 2.2. DIAGRAM OF THE SURGICAL PREPARATION OF RAT <i>IN VIVO</i> MODEL OF MYOCARDIAL ISCHEMIA.....	61
FIGURE 2.3 PHOTOGRAPHS OF THE <i>IN VIVO</i> MODEL SET UP.	62
FIGURE 2.4. THE <i>IN VIVO</i> RAT MODEL OF ACUTE MYOCARDIAL ISCHEMIA COUPLED TO MACLAB CHART RECORDING SOFTWARE.	63
FIGURE 2.5. PHOTOGRAPHS OF INFARCT SIZE PROCEDURE COMPONENTS.	66
FIGURE 2.6. EXAMPLES OF SLICES DEMARCATED FOR INFARCT SIZING.....	67
FIGURE 2.7. EXAMPLES OF CATEGORIZED ARRHYTHMIAS TAKEN FROM SAMPLES USED IN THESE STUDIES.	69
FIGURE 2.8. PHOTOGRAPH OF THE OXFORD 7.05-T SUPERCONDUCTING MAGNET.	70
FIGURE 2.9. PHOTOGRAPHS OF SURFACE COIL PLACEMENT UPON PERSPEX CRADLE....	72
FIGURE 2.10. MULTILAYERED AND MULTI-CHAMBERED DEVICE FOR COIL SAMPLE DEPTH CALIBRATION.	73
FIGURE 2.11. EXAMPLE OF TYPICAL PHOSPHORUS SPECTRA FROM A NON-ISCHEMIC RAT HEART USED IN THIS STUDY.	73
FIGURE 3.1 CHAPTER 3 TREATMENT PROTOCOL.....	84

FIGURE 3.2. EPISODES OF PREMATURE VENTRICULAR BEATS (PVB) AND SALVOS IN SALINE- CONTROLS AND THE FIVE TREATMENT GROUPS DURING 30 MIN ISCHEMIA.	86
FIGURE 3.3. EPISODES AND DURATIONS OF BIGEMINY IN SALINE-CONTROLS AND THE FIVE TREATMENT GROUPS DURING 30 MIN ISCHEMIA.....	86
FIGURE 3.5. SUM OF DURATIONS OF VENTRICULAR TACHYCARDIA (VT) AND VENTRICULAR FIBRILLATION (VF) IN SALINE CONTROLS AND THE FIVE TREATMENT GROUPS DURING 30 MIN ISCHEMIA.	88
FIGURE 3.6. EFFECTS OF TREATMENTS ON LEFT VENTRICLE NECROSIS AND INFARCT SIZE.	90
FIGURE 3.7. HEMODYNAMIC CHANGES IN SALINE- CONTROLS AND THE FIVE TREATMENT GROUPS AT BASELINE, BEFORE OCCLUSION, 20 MIN ISCHEMIA AND 30, 60 AND 120 MIN REPERFUSION. ..	93
FIGURE 4.1. CHAPTER 4 TREATMENT PROTOCOL.....	108
FIGURE 4.2. THE EPISODES AND DURATION OF VENTRICULAR TACHYCARDIA (VT) AND VENTRICULAR FIBRILLATION (VF) AND VT+VF DURING ISCHEMIA FOR SURVIVING RATS. ..	111
FIGURE 4.3. EFFECTS OF AL SOLUTION AND SEQUENTIAL ADMINISTRATION OF ADENOSINE AND LIGNOCAINE DURING ISCHEMIA AND/OR REPERFUSION ON INFARCT SIZE ..	113
FIGURE 4.4. HEMODYNAMIC CHANGES FOR ALL SURVIVING ANIMALS DURING THE COURSE OF THE FIRST EXPERIMENT. MEASUREMENTS WERE RECORDED THROUGHOUT PRETREATMENT/PREOCLUSION, ISCHEMIA AND REPERFUSION. ..	115
FIGURE 5.1. CHAPTER 5 TREATMENT PROTOCOL.....	125
FIGURE 5.2. HEMODYNAMIC PARAMETERS IN CONTROL (NO PRETREATMENT) AND AL SOLUTION RATS AT BASELINE (0 MIN AL SOLUTION; 5 MIN CONTROL), PRETREATMENT (5 MIN AL SOLUTION), DURING ISCHEMIA (30 MIN) AND REPERFUSION (30 MIN). ..	127
FIGURE 5.3. PERCENT CHANGES IN [ATP] FROM BASELINE IN CONTROL (NO PRETREATMENT) AND AL SOLUTION RATS AT BASELINE (0 MIN AL SOLUTION; 5	

MIN CONTROL), PRETREATMENT (5 MIN AL SOLUTION), DURING ISCHEMIA (30 MIN) AND REPERFUSION (30 MIN)	128
FIGURE 5.4. CHANGES IN [PCR] IN CONTROL (NO PRETREATMENT) AND AL SOLUTION RATS AT BASELINE (0 MIN AL SOLUTION; 5 MIN CONTROL), PRETREATMENT (5 MIN AL SOLUTION), DURING ISCHEMIA (30 MIN) AND REPERFUSION (30 MIN).....	129
FIGURE 5.5. CHANGES IN P _i PEAK INTEGRALS IN CONTROL (NO PRETREATMENT) AND AL SOLUTION RATS AT BASELINE (0 MIN AL SOLUTION; 5 MIN CONTROL), PRETREATMENT (5 MIN AL SOLUTION), DURING ISCHEMIA (30 MIN) AND REPERFUSION (30 MIN).	131
FIGURE 5.6. CHANGES IN INTRACELLULAR pH IN CONTROL (NO PRETREATMENT) AND AL SOLUTION RATS AT BASELINE (0 MIN AL SOLUTION; 5 MIN CONTROL), PRETREATMENT (5 MIN AL SOLUTION), DURING ISCHEMIA (30 MIN) AND REPERFUSION (30 MIN).	132
FIGURE 5.7. CHANGES IN [Mg ²⁺] IN CONTROL (NO PRETREATMENT) AND AL SOLUTION RATS AT BASELINE (0 MIN AL SOLUTION; 5 MIN CONTROL), PRETREATMENT (5 MIN AL SOLUTION), DURING ISCHEMIA (30 MIN) AND REPERFUSION (30 MIN).....	133
FIGURE 5.8. SCHEMATIC OF THE POSSIBLE MECHANISMS OF MAINTAINING ATP AND PCR IN A METABOLIC STEADY-STATE DURING AL INFUSION THROUGHOUT ISCHEMIA-REPERFUSION.	136
FIGURE 6.1. CHAPTER 6 TREATMENT PROTOCOL.....	148
FIGURE 6.2. THE EPISODES AND DURATION OF VENTRICULAR TACHYCARDIA (VT) AND VENTRICULAR FIBRILLATION (VF) AND VT+VF DURING ISCHEMIA FOR SURVIVING RATS IN ALL TREATMENT GROUPS.....	149
FIGURE 6.3. EFFECTS OF IPC, AL SOLUTION, CCPA PLUS LIDOCAINE, CCPA ALONE ON LEFT VENTRICLE NECROSIS AND INFARCT SIZE.	151

List of illustrations

PLATE I. "APOPLEXIE DU COEUR." ILLUSTRATION BY JEAN CRUVEILHIER (1791-1874) IN <i>ANATOMIE PATHOLOGIQUE DU CORPS HUMAN</i> (LEIBOWITZ, 1970).....	1
PLATE II. PHOTOGRAPH OF STRING GALVANOMETER, 1903, INVENTED BY WILLEM EINHOVEN (1860-1927) (LEIBOWITZ, 1970).....	57
PLATE III. ANATOMICAL SKETCHES OF A "TRANSPARENT" TORSO BY LEONARDO DA VINCI (1452-1519) (HERRLINGER, 1970).....	80
PLATE IV. THE FIRST DEPICTION OF THE ORIGIN OF CORONARY VESSELS FROM THE CORONARY SINUS BY LEONARDO DA VINCI (1452-1519) (LEIBOWITZ, 1970).....	80
PLATE V. ILLUSTRATION OF THE RUPTURED HEART OF KING GEORGE II (1683-1766) (LEIBOWITZ, 1970).....	104
PLATE VI. HEART WITHIN PERICARDIUM SURROUNDED BY THE LUNGS AND PART OF THE DIAPHRAGM (1543), BY ANDREAS VESALIUS (1514-1564) (HERRLINGER, 1970).....	121
PLATE VII. DIAGRAMMATIC RELATIONSHIP OF THE NORMAL ELECTROCARDIOGRAM (ECG) AND THE SEVEN TRANSMEMBRANE ACTION POTENTIALS OF EACH CELL TYPE BY FRANK NETTER (NETTER, 1969).....	144
PLATE VIII. WILLIAM HARVEY (1578-1657) USED THESE WOODCUT PRINTS TO DEMONSTRATE THE CONTINUOUS CIRCULATION OF BLOOD WITHIN A CONTAINED SYSTEM (LYONS, 1987).....	157